A RODENT BRAIN ORTHOTOPIC MODEL TO STUDY HUMAN MALIGNANT GLIOMA

Uroš Rajčević

Department of Genetic Toxicology and Cancer Biology, National Institute of Biology, Večna pot 111, 1000 Ljubljana, Slovenia E-mail: uros.rajcevic@nib.si

Summary: Glioblastomas are fatal brain tumors. They have a relatively low incidence, but the fact that regardless of the advances in therapy there hasn't been a major progress in survival over several decades urges the scientists to search for novel diagnostic tools, anti-tumor drug targets and therapies. It appears that our inability to successfully treat brain cancer mostly stems from the lack of understanding of the underlying brain tumor biology. Many rodent orthotopic models have been developed to address issues in drug development as well as biological origins of malignant gliomas. Establishing clinically relevant animal models of glioblastoma multiforme (GBM) remains a challenge, and many commonly used cell line-based models do not recapitulate the invasive growth patterns of patient GBMs. A novel orthotopic rat model of glioblastoma-the most malignant glioma in human-was recently developed, showing some stem-cell properties. The model is based on xenotransplantation of biopsy spheroids from human tumor tissue, into the brain of immunodeficient rats, where they initiated the growth of primary in most cases invasive and angiogenesis-independent glioblastomas. After serial passaging of tumors via spheroids in the subsequent generation of animals, the phenotype of the tumor changed. The most dramatic change was observed in approximately 1/3 of initially invasive tumors that changed into highly angiogenic and very aggressive. Some tumors though, remained invasive even after serial passages, while some were angiogenic from the start. The model thus provides combinations of angiogenic and invasive phenotypes and represents a good alternative to in vitro propagated cell lines for dissecting mechanisms of brain tumor progression. In vivo passaging of patient GBM biopsies produced tumors representative of the patient tumors, with high take rates and a reproducible disease course. The model has also been adapted to eGFP expressing immunodeficient mouse in which fluorescently marked tumors can be established in vivo.

Key words: malignant brain tumors; glioblastoma; nude rat; e-GFP NOD/SCID mice; xenograft; orthotopic transplantation; biopsy spheroids; translational medicine; molecular neuro-oncology

Introduction

Gliomas

Gliomas are brain tumors with some characteristics of brain support tissue – the glia. Malignant gliomas (anaplastic astrocytoma and glioblastoma multiforme-grade III and IV astrocytomas on WHO scale respectively) are predominant malignant brain tumors in human and have a poor prognosis with an average patient survival under current treatment regimens ranging between 12-14 months.

Received: 25 October 2010 Accepted for publication: 17 November 2010 Tumors are characterized by rapid cell growth, extensive neovascularization and diffuse cellular infiltration of normal brain structures (1). They have a relatively low overall incidence, but the fact that in spite of major progress in neurosurgery and oncology (including chemo-, radio- and, more recently biological-therapy) there hasn't been a major progress in malignant gliomas survival over several decades (2-7), urges the scientists to search for novel diagnostic tools, anti-tumor drug targets and therapies.

Animal Tumor Models

In vivo animal modeling provides essential tumorhost interactions and is a more accurate way of modeling human cancer than *in vitro*. We subdivide rodent models into xenograft tumor models – addressed in particular in this review - and models of spontaneous tumor formation in genetically engineered rodents. These models help us address important issues in drug development: toxicity and *in vivo* antitumor effectiveness (8, 9) (where the models are a matter of highly standardized procedures) as well as other basic phenomena in tumor origin and function such as angiogenesis, invasion and many others (3, 4, 10, 11) in the field of cancer research. *In vivo* modeling of drug efficacy is a gold standard required by a majority of pharmaceutical companies (12).

Xenograft tumors are initiated by implantation or injection of primary tumor cells or cell lines s.c. (under the skin) or orthotopically (into native tumor site) of syngeneic (genetically identical or closely related, so as to allow tissue transplant; immunologically compatible.), xenogeneic (derived or obtained from an organism of a different species, as a tissue graft), allogeneic (genetically different although belonging to or obtained from the same species, as in tissue grafts) or, immunosuppressed, immunodeficient, or newborn immunonaive animals (8, 9, 13, 14).

Rodent glioblastoma (GBM) models have been used for over 40 years but the extent to which they recapitulate the characteristics encountered in human GBMs remains controversial (8, 9). The advantages of xenograft glioma models are their highly efficient gliomagenesis, reproducible growth rates, and an accurate knowledge of the location of the tumor (6, 15, 23). However, in xenograft models stepwise genetic changes thought to occur during tumor progression are often missing; injected or implanted cells or cell lines have often been altered by culture or isolation conditions and lack their native tumor stroma. Murine models (e.g. (18) of GBM appear to recapitulate several of the human GBM histopathological features and, considering their reproducibility and availability, they constitute a valuable in vivo system for preclinical studies (8). On the other hand, many xenograft tumors lack histologicaly accurate vascularization, and rarely recapitulate tumor-of-origin phenotype (17). Immunodeficient rodents do not show antitumor immune effects and can produce false positives during drug trials (9). Syngeneic murine models, such as GL26 mouse glioma cells in C57BL6 mice (18) and CNS1 rat glioma cells in Lewis rats (19, 20) are nonimmunogenic. Thus, syngeneic glioma models are excellent for studying the response of brain tumors to immunotherapy (19, 20). Establishing clinically relevant animal models of gliomas that would fully reflect the situation in human malignant glioma remains a challenge.

Two novel rat and mouse models to study glioblastoma

An orthotopic rat xenograft model of glioblastoma was developed (21) to address, among others, the tumor-host interaction issues leading to tumor circumscription, histologically accurate vascularization and to recapitulate accurately the tumor-oforigin phenotype. The model is based on a serial xenotransplantation of glioma biopsy spheroids, generated from glioma tissue (22) into the brain of nude rats. There the glioma spheroids induce the growth of primary, in most cases very invasive glioblastomas in 4-6 months time. These tumors coopted the host vasculature and presented as an aggressive disease without signs of angiogenesis. The malignant cells expressed neural stem cell markers and showed a migratory behavior similar to normal human neural stem cells. When the rats became ill, they were sacrificed and spheroids were generated from their (human) tumors and implanted in the next generation of nude rats. This way, the tumors were passed in a total of 4-6 consecutive generations of nude rats (Figure 1). The most dramatic change was observed in approximately 1/3 of initially invasive tumors that changed into highly angiogenic and very aggressive, less invasive (more circumscribed), but which grew much faster due to good blood supply, killing a rat in 2-3 months. This switch to angiogenic phenotype was characterized by a reduction in stem cells markers (21, 23). Some tumors though, remained invasive even after serial passages, while some were angiogenic from the start. At the level of gene expression and immunoblotting proinvasive genes were up-regulated and angiogenesis signaling genes were down-regulated in invasive tumors. In contrast, proinvasive genes were down-regulated in the angiogenesis-dependent tumors derived from the invasive tumors (21). Uncoupling of invasion and angiogenesis, represented by the stemlike cancer cells and the cells derived from them respectively, points at two different mechanisms that drive tumor progression. Although the mechanism behind the phenotypic shift is not fully understood, HIF-1 expression seems to be triggered by hypoxia, because it was not constitutively expressed by highgeneration tumor spheroids cultured under normoxic conditions (21).



Figure 1: The orthotopic rat glioblastoma model

Thus, by serial passaging, this model uncouples and recapitulates for the first time the two major phenotype characteristics of human glioblastoma which makes it one of the best available glioblastoma models and opens the door to multiple applications in basic, translational and pre-clinical research. The major difference and - possibly - the advantage of this model over other described rodent models of glioblastoma (8, 9) arises from the fact that in the biopsy spheroids, which are structures of heterogenic cellular population, at least part of original glioma microenvironment is conserved, a part essential in inducing the tumor upon transplantation. This microenvironment is absent in xenografts generated from immortalized glioma cell lines (8, 9), but may be conserved to some extent in primary glioma cell lines derived from glioma patients and in glioma tissue grafts. The phenotypes, derived from the same tumor sample, may develop through a selection process where most cells in the biopsy specimens die following implantation in the rat brain. The cells that survive show stem cell-like properties, and are able to adapt to the new microenvironment where they divide and produce new tumor clones that show rapid growth and angiogenesis (21, 24).

The concept of the Rodent Glioblastoma model

Samples of glioblastoma, resected from the patients are finely minced and grown in cell culture medium. There the cells organize in tumor biopsy spheroids of about 300μ m in size. Spheroids are orthotopically implanted into the brain of immunodeficient rodents. There, in most cases, they induce the growth of primary glioblastomas of invasive nature, but non-angiogenic in 4-6 months. With serial passaging in the subsequent generations of rodents via spheroids, the tumor phenotype will change in 1/3 of cases to fully angiogenic in the last generation xenograft with an onset of tumor in 2-3 months.

eGFP NOD/Scid mouse model

This glioblastoma model was recently adapted to a NOD/Scid mouse expressing enhanced green fluorescent protein (eGFP) (Figure 2). In this model human and mouse tumors marked with red fluorescent protein can be established in vivo, both at subcutaneous and orthotopic locations (25). Using modern microscopy techniques, the intricate co localization of tumor and host cells in situ can be visualized in detail. Technology of fluorescence-activated cell sorting (FACS), allows for complete and reliable separation of the host cells from the tumor cells, thus providing a system for detailed cellular and molecular analysis of tumor-host cell interactions. The fact that tumor and host cells can be reliably identified also allows for detection of double-positive cells, possibly arising from cell fusion events or horizontal gene transfer (two possible origins of tumor initiating - tumor stem cells; (26)). Similarly, the model can be applied for the detection of circulating metastatic cells and for detailed studies on the vascular compartments within tumors, including vasculogenic mimicry (25). Thus, the model should provide significant insight into how tumor cells communicate with their microenvironment.



Figure 2: Tumor host interaction studies in an eGFP NOD/Scid mouse model

U87 dsRed expressing human glioblastoma cells were implanted in the flank of eGFP NOD/Scid mouse. (A-C) Endogenous fluorescence on a frozen histological section from a U87 subcutaneous tumor showing eGFP-expressing host cells within the tumor bed (direct eGFP and dsRed fluorescence). Scale bars = $5 \mu m$ (A); 10 μm (B, C). (D) a newborn eGFP NOD/Scid mouse glowing in green fluorescence under the UV light

Applications of the rodent glioblastoma model

Since its establishment, the rat model of glioblastoma was characterized in detail on several levels and used in multiple assays ranging from basic research to drug testing.

MR spectroscopy

In this study the metabolic properties of the two experimental phenotypes were analyzed. The early generation (EG) showed an intact blood-brain barrier and normal vascular morphology. In contrast, the high generation (HG) exhibited leaky vessels and necrosis. The rats with HG tumor had raised concentrations of choline and myo-inositol, and decreased concentrations of glutamate and N-acetylaspartate. In the LG tumor group, similar changes in metabolic concentrations were detected, although the alterations were more pronounced. The LG tumors also had higher concentrations of choline, taurine, and lactate. Results of this study showed that metabolic profiles could be used to distinguish between two glioblastoma phenotypes. More pronounced anaerobic metabolism was present in the LG stem-cell-like tumors, suggesting a more malignant phenotype (27).

Table	1: Applic	ations of	of the	rodent	gliobl	astoma	model
-------	-----------	-----------	--------	--------	--------	--------	-------

Applications		Model Reference		Comments			
Metabolite	Determination of meta-	Nude rat glioma	Thorsen F., et al.,	Metabolic profiles produced by MRS could be used to distinguish			
analysis	bolic properties of the	xenograft	2008 (27)	between two distinct glioblastoma phenotypes			
	xenografts by Magnetic						
	(MRS)						
Tumor initiation.	Othotopic, stereotactic	Nude rat glioma	Sakariassen PO et	Separation of early /invasive tumor phenotypes and late/angiogenic			
take and repro-	glioblastoma xenograft	xenograft	al., 2006 (21)	tumors by serial passaging in nude rats			
ducibility	model construction and	0		V 1 0 0			
	characterization						
	Tumorigenesis; neu-	Nude rat glioma	Wang J et al.,	In vivo passaging of patient GBM biopsies produced tumors represen-			
	ropathological and	xenograft	2009 (23)	tative of the patient tumors, with high take rates and a reproducible			
	dioblastoma xenograft			uisease course.			
	model						
	Tumor-host interaction	eGFP expressing	Niclou SP et al.,	Fluorescence-based intricate co localization			
	studies	NOD/SCID mouse	2008 (25)	of tumor and host cells in situ.			
Anti-tumor drug	Radio surgery	glioblastoma spher-	Thorsen FA et al,	Radio surgery of malignant gliomas might be effective in controlling			
testing and		oids, nude rat glioma	2007 (30)	tumor progression in selected glioblastoma patients.			
ment studies		xenogran					
	The response of the two	Nude rat glioma	Johannessen T-CA et	Highly invasive tumors shown to be more chemo resistant than angio-			
	phenotypes to doxoru-	xenograft	al., 2009, (24)	genic tumors derived from the same patients.			
	bicin						
	The effect of hyperovic	BT/C rat diama	Stubr I FB et al	Increased pO2 levels in experimental gliamas using normalyzic and			
	treatment on BT4C rat	xenografts	2007 (31)	moderate hyperbaric oxygen therapy, caused a significant reduction in			
	glioma xenografts	8		tumor growth, a process characterized by enhanced cell death, reduced			
	5 5			vascular density and changes			
Gene Therapy	Adenoviral vector (AAV)	Glioma cell lines,	Thorsen FA et al.,	AAV4 and AAV5 serotypes may be used to transduce biologically di-			
	transduction	glioblastoma spher-	2006 (32)	verse glioma cell lines. They also penetrate and transduce solid human			
		olds, nude rat glioma		tumor tissue derived from patient diopsies.			
	Lentiviral vector transduc-	human embryonic	Huszthy PC et al.	Lymphocytic choriomeningitis virus glycoprotein (LCMV-GP) and			
	tion	kidney cell line 293T,	2009 (33)	vesicular stomatitis virus glycoprotein (VSVG) pseudotyped lentiviral			
		TE671 cell line, nude		vectors efficiently transduced human glioblastoma cells and cancer			
		rat glioma xenograft		stem-like cells. Pseudotyped gamma retroviral vectors, similar to those			
				evaluated for clinical therapy of glioblastoma, showed inefficient gene			
	Oncolvtic HSV-1 based	Nude rat glioma	Huszthy PC et al.	Favorable cellular responses to G207 treatment seen from a clinical			
	vector G207	xenograft	2010, (34)	viewpoint, such as reduced tumor cell proliferation, more frequent			
				events of tumor cell death and a strongly attenuated tumor vascular			
				compartment.			
Biomarker	The glioma-associated	Nude rat glioma	Hedberg KM et al,	Different biological roles for individual gangliosides; antibodies or			
research	gangliosides 3k-isoLM1,	xenograft	2001, (35)	ligands directed against GD3 and 3k-isoLM1 might be complementary			
L	Expression of extracel-	Glioma cell lines.	Mahesparan R et al.	Possible biological function of tenascin, vitronectin, laminin, fibronec-			
	lular matrix components	nude rat glioma	2003, (16)	tin and collagen type IV in highly invasive malignant tumors of glial			
	in a highly infiltrative	xenograft		origin.			
	glioma model						
	PDI protein expression	Nude rat glioma	Goplen D et al.,	PDI was shown to be expressed on migrating and invading glioma cells.			
	tumors	ACHUGIAN	2000 (30)				
	Tumor initiating cell	Nude rat glioma	Wang J et al.,	Cd133- glioma cells are tumorigenic and can produce cd133+ tumors,			
	markers	xenograft	2007(29)	CD133 expression coincides with the onset of angiogenesis and a			
				shorter survival			
	Global membrane pro-	Nude rat glioma	Rajcevic U et al.,	Known and novel candidate proteins were identified that characterize			
	teomics	xenogran	2009 (37)	use switch from a non-angrogenic to a nignly angrogenic pnenotype. En-			
				cells in the angiogenic compared with the non-angiogenic phenotype.			
	Neural cell adhesion mol-	Nude rat glioma	Duenisch P et al.,	The expression of NCAM-140 inversely correlated with the WHO grade			
	ecule (NCAM) as a glioma	xenograft	2010, (38)	of human gliomas			
	marker for the biological						
	aggressiveness	1	1	1			

Tumor initiation, take and reproducibility

The tumor take rates for xenografted GBM biopsies were 96% and close to 100% at subsequent passages *in vivo*. Only one of four lower grade tumors engrafted. MRI typically revealed changes related to tumor growth, several months prior to the onset of symptoms (23).

In another study, CD133 expression was analyzed at various passages. CD133 is a cell surface marker identified as a putative marker of brain tumor-initiating cells (28). During passaging, the tumors gradually displayed more contrast enhancement, increased angiogenesis, shorter survival and increased CD133 expression. CD133 negative cells derived from 6 different patients were tumorogenic when implanted into the rat brains. For 3 of these patients, analysis showed that the resulting tumors contained CD133 positive cells. In this assay, the authors showed that CD133 negative glioma cells were tumorogenic in nude rats, and that CD133 positive cells can be obtained from these tumors. Upon passaging of the tumors in vivo, CD133 expression is up regulated, coinciding with the onset of angiogenesis and a shorter survival. Authors also suggested that CD133 may not be essential in tumor initiation process (29).

Anti-tumor drug testing and glioma treatment studies

In a study focused on the radiobiological effects of the Gamma knife (Gamma Knife is currently used to boost treatment of malignant gliomas) the growth and invasiveness of human glioblastoma spheroids xenografted into nude rat brains were assessed after radio surgery. A dose-dependent inhibition of tumor growth and invasion, as well as a dose-dependent increase in animal survival was observed. The results indicated that radio surgery of malignant gliomas might be effective in controlling tumor progression in selected glioblastoma patients (30).

In a study aimed at investigating how the two phenotypes responded *in vitro* to doxorubicin, a clinically potent cytotoxic drug for solid tumors, highly invasive tumors shown to be more chemo resistant than angiogenic tumors derived from the same patients. It was suggested that treatment resistance in glioblastomas could be related to PI3K/AKT activity in stemlike tumor cells, and that targeted interference with the PI3K/AKT pathway might differentiate and sensitize this subpopulation to chemotherapy (24).

Another study described the biological effects of

hyperoxic treatment on BT4C rat glioma xenografts *in vivo* with special reference to tumor growth, angiogenesis, apoptosis, general morphology and gene expression parameters. Increased pO2–levels in experimental gliomas, using normobaric and moderate hyperbaric oxygen therapy, caused a significant reduction in tumor growth. This process was characterized by enhanced cell death, reduced vascular density and changes in gene expression corresponding to these effects (31).

Gene Therapy

In one of the initial studies on the delivery vehicles for gene transfer strategies directed at the central nervous system (CNS), muscle and liver performed on the featuring model the transduction efficacy of AAV serotypes 4 and 5 were compared to AAV2, both in vitro and in intracranial GBM xenografts . While all three AAV serotypes were able to transduce the glioma cell lines when added individually or when they were administered in concert, AAV2 transduced the glioma cells most effectively compared to AAV4 or AAV5. Upon infecting glioblastoma spheroids in vitro, all three AAV serotypes efficiently transduced cells located at the surface as well as within deeper layers of the spheroids. In addition, both AAV4 and AAV5 were able to transduce human glioblastoma xenografts implanted intracranially. Authors suggested that beside AAV2 serotype, AAV4 and AAV5 serotypes may also be used to transduce biologically diverse glioma cell lines and may be used in developing treatment vehicles for human malignant gliomas (32).

The rat xenograft model was used to analyze the transduction pattern and therapeutic efficacy of lentiviral pseudotyped vectors. Both, lymphocytic choriomeningitis virus glycoprotein (LCMV-GP) and vesicular stomatitis virus glycoprotein (VSVG) pseudotyped lentiviral vectors efficiently transduced human glioblastoma cells and cancer stem-like cells. In contrast, pseudotyped gamma retroviral vectors, similar to those evaluated for clinical therapy of glioblastoma, showed inefficient gene transfer. In a therapeutic approach using the suicide gene herpes simplex virus thymidine kinase (HSV-1-tk) fused to eGFP, both lentiviral vectors mediated a complete remission. In all recurrent tumors, surviving eGFP-positive tumor cells were found, advocating prodrug application for several cycles to even enhance and prolong the therapeutic effect. The inefficient gene delivery by gamma retroviral vectors is in line with the results obtained in clinical therapy for GBM and thus confirms the high reproducibility of the invasive glioma animal model for translational research (33).

In another assay the cellular effects of the oncolytic HSV-1 based vector, G207, on the tumor microenvironment were evaluated. The xenografted tumors were quantitatively evaluated 10-30 days after G207 injection for virus-induced changes in proliferation, apoptosis and vascularity. Vector spread and the infiltration pattern of CD68-positive inflammatory cells were assessed. Proliferation indices were lower, whereas apoptotic counts were elevated in plaques as compared with that in non-infected areas of the same lesions, as well as in corresponding control xenografts. A decline in the number of blood vessels was noticed in the plaques and the vascular area fractions were reduced. CD68-positive inflammatory cells accumulated in the plaques. The study highlighted the favorable cellular responses to G207 treatment seen from a clinical viewpoint, such as reduced tumor cell proliferation, more frequent events of tumor cell death and a strongly attenuated tumor vascular compartment (34).

Biomarker research

A substantial number of biomarker research studies were thus far performed on the model. Initially roles for glioma-associated gangliosides (3kisoLM1, GD3 and GM2) were assessed in the model, pointing out that different biological roles for individual gangliosides may exist and that the antibodies or ligands directed against GD3 and 3k-isoLM1 might be complementary when applied in the treatment of human glioblastomas (35).

In a different study expression of extracellular matrix (ECM) components in this highly infiltrative *in vivo* glioma model was analyzed. The cellular origin for several of these ECM components was identified using human-specific monoclonal antibodies and polyclonal antibodies detecting epitopes from both species (human and rat). Observed staining patterns clarified the cellular origin and indicated the possible biological function of tenascin, vitronectin, laminin, fibronectin and collagen type IV in these highly invasive malignant tumors of glial origin (16).

In the initial proteomic assays on biomarkers on the featuring model the researchers showed that PDI, one of the most prominently up-regulated proteins in invasive tumors is strongly expressed on invasive glioma cells, in both xenografts and at the invasive front of human glioblastomas. Using *in* *vitro* assays, PDI was shown to be expressed on migrating glioma cells. Functional significance of PDI in cell migration and invasion was tested *in vitro*, showing an important role of PDI in glioma cell invasion (36).

In a global proteomics comparison of the two xenograft phenotypes we were able to identify several thousand proteins in membrane-enriched fractions of which 1460 were extracted as quantifiable proteins (isoform- and species specific and present in more than one sample). Known and novel candidate proteins were identified that characterize the switch from a non-angiogenic to a highly angiogenic phenotype. The data pointed to enhanced intercellular cross-talk and metabolic activity adopted by tumor cells in the angiogenic compared with the non-angiogenic phenotype. The identified proteins could be further exploited as biomarkers or therapeutic targets for malignant gliomas (37).

In a different assay the relevance of neural cell adhesion molecule (NCAM) as a glioma tissue marker for the biological aggressiveness of these tumors was assessed. The expression of NCAM-140 inversely correlated with the WHO grade of human gliomas. The lost expression of NCAM-140 in human glioblastomas and in brain metastases enabled the investigation of the brain– tumor interface and the definition of glioblastoma invasion patterns and showed that brain metastases are more invasive than ever thought (38).

Conclusions

In vivo passaging of patient GBM biopsies in rats or mice produced tumors representative of the patient tumors, with high take rates and a reproducible disease course. The main advantage of the model provides combinations of angiogenic and invasive phenotypes and represents a good alternative to in vitro propagated cell lines for dissecting mechanisms of brain tumor progression (Wang J, BMC Cancer 2009) as well as to other in vivo orthotopic rodent GBM models. Thus far, the model has been extensively used in a variety of applications, ranging from basic research of cancer biology and biomarkers including genomics, transcriptomics, proteomics and metabolomics, through translational research of therapeutics and therapeutic modalities to preclinical research of drugs approved for use in other types of cancer. The results of this research provided important novel insights into the mechanisms of tumor initiation, promotion and progression

through transcriptomic markers, through novel protein biomarkers, validated on clinical and pre-clinical material. Results also indicated the metabolic changes linked to phenotype switch in the model. A possibility of separating the tumor-host compartments in the model based on cellular, fluorescence based- or biochemical, protein sequence-based level enables an unprecedented potential in tumor-host interaction studies, the complex of events crucial for the understanding of tumor initiation and function. The research of novel therapeutics and therapeutic modalities has shown a great potential of this model as it recapitulates the main features of the GBM, for *in vivo* studies including radio-, chemo- and gene therapies.

Acknowledgements

Dr Simone P. Niclou, NorLux Neuro-Oncology Laboratory, CRP-Santé, Luxembourg is kindly acknowledged for the critical review of the manuscript and for providing photos of the animal model. The work was funded by Fonds National de la Recherche (FNR), Luxembourg (contract FNR-AFR # PDR-08-007) to U.R. and by Slovene Research Agency (Program Grant # P1–0245).

References

1. Louis DN, Ohgaki H, Wiestler OD, et al. The 2007 WHO classification of tumors of the central nervous system. Acta Neuropathol 2007;114:97-109.

2. Castro MG, Cowen R, Williamson IK, et al. Current and future strategies for the treatment of malignant brain tumors. Pharmacol Ther 2003;98:71-108.

3. Chiocca EA. Oncolytic viruses. Nat Rev Cancer 2002;2:938-50.

4. Curtin JF, King GD, Candolfi M, et al. Combining cytotoxic and immune-mediated gene therapy to treat brain tumors. Curr Top Med Chem 2005;5:1151-70.

5. Gomez-Manzano C, Yung WK, Alemany R, Fueyo J. Genetically modified adenoviruses against gliomas: from bench to bedside. Neurology 2004;63:418-26.

6. King GD, Curtin JF, Candolfi M, Kroeger K, Lowenstein PR, Castro MG. Gene therapy and targeted toxins for glioma. Curr Gene Ther 2005;5:535-57.

7. Prados MD. Future directions in the treatment of malignant gliomas with temozolomide. Semin On-col 2000;27:41-6.

8. Candolfi M, Curtin JF, Nichols WS, et al. Intracranial glioblastoma models in preclinical neurooncology: neuropathological characterization and tumor progression. J Neurooncol 2007;85:133-48.

9. Fomchenko EI, Holland EC. Mouse models of brain tumors and their applications in preclinical trials. Clin Cancer Res 2006;12:5288-97.

10. Fecci PE, Mitchell DA, Archer GE, et al. The history, evolution, and clinical use of dendritic cell-based immunization strategies in the therapy of brain tumors. J Neurooncol 2003;64:161-76.

11. Rainov NG, Ren H. Gene therapy for human malignant brain tumors. Cancer J 2003;9:180-8.

12. Seller WR. Cancer drug development in the modern era. In: EC H, ed. Mouse models of human cancers. Hoboken (New Jersey): John Wiley and Sons; 2004.

13. Lassmann S, Opitz OG. The new look of colorectal cancer stem cells. Gastroenterology 2008;134:1262-4.

14. Shapiro WR, Basler GA, Chernik NL, Posner JB. Human brain tumor transplantation into nude mice. J Natl Cancer Inst 1979;62:447-53.

15. Brabletz T, Jung A, Spaderna S, Hlubek F, Kirchner T. Opinion: migrating cancer stem cells an integrated concept of malignant tumor progression. Nat Rev Cancer 2005;5:744-9.

16. Mahesparan R, Read TA, Lund-Johansen M, Skaftnesmo KO, Bjerkvig R, Engebraaten O. Expression of extracellular matrix components in a highly infiltrative in vivo glioma model. Acta Neuropathol 2003;105:49-57.

17. Finkelstein SD, Black P, Nowak TP, Hand CM, Christensen S, Finch PW. Histological characteristics and expression of acidic and basic fibroblast growth factor genes in intracerebral xenogeneic transplants of human glioma cells. Neurosurgery 1994;34:136-43.

18. AlbrightL,MadiganJC,GastonMR,Houchens DP. Therapy in an intracerebral murine glioma model, using Bacillus Calmette-Guerin, neuraminidasetreated tumor cells, and 1-(2-chloroethyl)-3-cyclohexyl-1-nitrosourea. Cancer Res 1975;35:658-65.

19. Ali S, Curtin JF, Zirger JM, et al. Inflammatory and anti-glioma effects of an adenovirus expressing human soluble Fms-like tyrosine kinase 3 ligand (hsFlt3L): treatment with hsFlt3L inhibits intracranial glioma progression. Mol Ther 2004;10:1071-84.

20. Ali S, King GD, Curtin JF, et al. Combined immunostimulation and conditional cytotoxic gene therapy provide long-term survival in a large glioma model. Cancer Res 2005;65:7194-204.

21. Sakariassen PO, Prestegarden L, Wang J, et al. Angiogenesis-independent tumor growth mediated by stem-like cancer cells. Proc Natl Acad Sci U S A 2006;103:16466-71.

22. Bjerkvig R, Tonnesen A, Laerum OD, Backlund EO. Multicellular tumor spheroids from human gliomas maintained in organ culture. J Neurosurg 1990;72:463-75.

23. Wang J, Miletic H, Sakariassen PO, et al. A reproducible brain tumor model established from human glioblastoma biopsies. BMC Cancer 2009;9:465.

24. Johannessen TC, Wang J, Skaftnesmo KO, et al. Highly infiltrative brain tumors show reduced chemosensitivity associated with a stem cell-like phenotype. Neuropathol Appl Neurobiol 2009;35:380-93.

25. Niclou SP, Danzeisen C, Eikesdal HP, et al. A novel eGFP-expressing immunodeficient mouse model to study tumor-host interactions. Faseb J 2008;22:3120-8.

26. Bjerkvig R, Tysnes BB, Aboody KS, Najbauer J, Terzis AJ. Opinion: the origin of the cancer stem cell: current controversies and new insights. Nat Rev Cancer 2005;5:899-904.

27. Thorsen F, Jirak D, Wang J, et al. Two distinct tumor phenotypes isolated from glioblastomas show different MRS characteristics. NMR Biomed 2008;21:830-8.

28. Singh SK, Hawkins C, Clarke ID, et al. Identification of human brain tumor initiating cells. Nature 2004;432:396-401.

29. Wang J, Sakariassen PO, Tsinkalovsky O, et al. CD133 negative glioma cells form tumors in nude rats and give rise to CD133 positive cells. Int J Cancer 2008;122:761-8.

30. Thorsen F, Enger PO, Wang J, Bjerkvig R, Pedersen PH. Human glioblastoma biopsy spheroids xenografted into the nude rat brain show growth inhibition after stereotactic radio surgery. J Neurooncol 2007;82:1-10.

31. Stuhr LE, Raa A, Oyan AM, et al. Hyperoxia retards growth and induces apoptosis, changes in vascular density and gene expression in transplanted gliomas in nude rats. J Neurooncol 2007;85:191-202.

32. Thorsen F, Afione S, Huszthy PC, et al. Adenoassociated virus (AAV) serotypes 2, 4 and 5 display similar transduction profiles and penetrate solid tumor tissue in models of human glioma. J Gene Med 2006;8:1131-40.

33. Huszthy PC, Giroglou T, Tsinkalovsky O, et al. Remission of invasive, cancer stem-like glioblastoma xenografts using lentiviral vector-mediated suicide gene therapy. PLoS One 2009;4:e6314.

34. Huszthy PC, Immervoll H, Wang J, et al. Cellular effects of oncolytic viral therapy on the glioblastoma microenvironment. Gene Ther 2010;17:202-16.

35. Hedberg KM, Mahesparan R, Read TA, et al. The glioma-associated gangliosides 3'-isoLM1, GD3 and GM2 show selective area expression in human glioblastoma xenografts in nude rat brains. Neuropathol Appl Neurobiol 2001;27:451-64.

36. Goplen D, Wang J, Enger PO, et al. Protein disulfide isomerase expression is related to the invasive properties of malignant glioma. Cancer Res 2006;66:9895-902.

37. Rajcevic U, Petersen K, Knol JC, et al. iTRAQ based proteomic profiling reveals increased metabolic activity and cellular crosstalk in angiogenic compared to invasive Glioblastoma phenotype. Mol Cell Proteomics 2009;8:2595-612.

38. Duenisch P, Reichart R, Mueller U, et al. Neural cell adhesion molecule isoform 140 declines with rise of WHO grade in human gliomas and serves as indicator for the invasion zone of multiform glioblastomas and brain metastases. J Cancer Res Clin Oncol 2011;137:399-414.

GLODAVSKI MOŽGANSKI ORTOTOPIČNI MODEL ZA ŠTUDIJ ČLOVEŠKEGA MALIGNEGA GLIOMA

U. Rajčević

Povzetek: Maligni gliomi so neozdravljivi možganski tumorji. Se relativno redko pojavljajo, vendar ne glede na izboljšave pri terapiji večjega napredka pri preživelosti bolnikov ni bilo že več desetletij. Zaradi tega je pomembno raziskovanje novih diagnostičnih orodij, ugotavljanje novih tarč v tumorskem tkivu in razvijanje novih načinov zdravljenja. Zdi se, da naša nezmožnost uspešno pozdraviti raka na možganih izvira iz nepoznavanja bioloških in biokemičnih lastnosti tumorjev. Z leti je nastalo več glodavskih ortotopičnih modelov, ki so lahko uporabni pri razvoju novih zdravil in pri proučevanju bioloških osnov malignih gliomov. Razvoj klinično pomembnih živalskih modelov za multiformni glioblastom (GBM) ostaja izziv in številni, pogosto uporabljeni modeli na osnovi celičnih linij, ne odražajo vzorcev invazivne rasti GBM pri bolnikih. Nedavno je bil razvit nov ortotopični podganji model glioblastoma - najbolj maligne oblike glioma pri človeku, ki kaže nekatere značilnosti, podobne matičnim celicam. Model je nastal na osnovi medvrstne presaditve biopsijskih sferoidov iz vzorcev človeških tumorjev v možgane podgan z oslabljenim imunskim sistemom, kjer so sferoidi povzročili rast primarnih in večinoma invazivnih, od angiogeneze neodvisnih glioblastomov. Po zaporednem prenosu teh tumorjev preko sferoidov v naslednje generacije živali se je fenotip tumorja spremenil. Najbolj dramatična sprememba je bila ugotovljena pri približno tretjini sprva invazivnih tumorjev, ki so se spremenili v visoko angiogene in zelo agresivne. Nekaj tumorjev je ostalo invazivnih kljub zaporednemu prenosu v naslednje generacije podgan, medtem ko so bili nekateri angiogeni že od začetka. Model torej prinaša kombinacijo angiogenih in invazivnih fenotipov in predstavlja dobro alternativo celičnim linijam, gojenim v pogojih in vitro, za ugotavljanje mehanizmov napredovanja možganskih tumorjev. Zaporedni prenos bolnikovih biopsij GBM in vivo je povzročil nastanek tumorjev, podobnih pacientovim, z visoko ravnijo tumorigeneze in ponovljivim potekom bolezni. Model je bil prilagojen tudi mišim z oslabljenim imunskim odgovorom, ki izražajo zeleno fluorescenčno beljakovino in kjer lahko vzpodbudimo nastanek fluorescenčno označenih tumorjev pri živih živalih.

Ključne besede: maligni možganski tumorji; glioblastom, gole podgane; miši e-GFP NOD/SCID; ksenograft; ortotopična transplantacija; biopsijski sferoidis; translacijska medicina; molekularna nevroonkologija