APPETITE REGULATION AND OBESITY: EMPHASIS ON GHRELIN AND GHRELIN RECEPTOR

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Summary: Obesity is one of the leading preventable causes of death worldwide. In its epidemic it is of increasing interest for the pharmaceutical industry to develop drugs that reduce appetite. By reducing appetite overall energy consumption is also reduced. The idea is simple; however, the hormonal system and mechanisms regulating energy intake are extremely complex and therefore drug development is not straightforward. Throughout the central nervous system (CNS), central and peripheral hormones are involved in food intake and body weight balance. They are tightly regulated by the hypothalamus, brainstem and reward circuits, on the basis of both cognitive inputs and diverse humoral and neuronal signals of nutritional status. Several peptides and hormones such as: neuropeptide Y (NPY), melanocortins, cocaine and amphetamineregulated transcript (CART), peptide YY (PYY), pancreatic polypeptide (PP), cholecystokinin (CCK), oxyntomodulin (OXM), glucagon-like peptide 1 (GLP-1), glucose-dependant insulinotropic hormone (GIP), bombesin, leptin and one of the latest discovered and not yet well known - ghrelin, have all revealed an important role in short- and long-term regulation of food intake. This review summarizes the complexity of factors involved in the regulation of appetite and food intake in different areas of the brain, especially in the hypothalamus, and the relationships between the central and peripheral peptides and hormones involved, with emphasis on ghrelin and its receptor together with their potential role as targets for treatment of obesity. The ghrelin receptor originally received considerable attention from pharmaceutical companies because of its prominent role in the release of growth hormone. However, the discovery of the orexigenic properties of ghrelin turned the ghrelin receptor (ghrR) into a target for anti-obesity drugs.

Key words: appetite regulation; obesity; ghrelin; ghrelin receptor

Introduction

Obesity is one of the most prevalent nutritional problems worldwide; therefore it is important to understand the role of the brain, the peptides and hormones secreted by brain, as well as the periphery organs that regulate appetite and food intake. Two-thirds of North Americans are overweight or obese (1), indicated by a body mass index (BMI) exceeding 30 kg/m². Similar trends are observed in Slovenia and other parts of Europe. Being overweight, especially if the body fat is concentrated around the abdomen, is associated with higher risk of various

Received: 3 December 2010 Accepted for publication: 1 August 2011 diseases: type 2 diabetes, hypertension, ischaemic heart disease, stroke, endometrial carcinoma, osteoarthritis, gall stones and cancers (colon, kidney, endometrial and postmenopausal breast cancer) (2). Even obesity in pets, especially dogs and cats, is becoming more frequent. Proportions of overweight and obese dogs in the USA range from 23% to 41%, with about 5.1 % classed as obese. In Australia the proportion of obesity in dogs and cats is even higher (7.6 % and 6.4 %, respectively) (3). Furthermore it has been shown that the risk of obesity in dogs is related to obesity of their owners; however, no correlation between cats and their owners was observed (4).

With the expanding knowledge about the areas of the CNS and the central and peripheral hor-

mones involved in food intake and body weight balance, we are increasing our ability to influence the processes. Hormones, and more so their receptors, represent potential targets in the development of novel anti-obesity drugs. The neuroscience of obesity and the connecting knowledge about all factors influencing obesity is a target of intense interest. The emphasis of research is focused on transferring the knowledge into new treatments for obesity and its related diseases (5). Recent areas of interest are the orexigenic (appetite stimulant) properties of ghrelin and its receptor - ghrelin receptor (ghrR) (6). Discovery of both turned ghrR into a target for pharmaceutical industry to develop anti-obesity drugs.

Peripheral and central appetite regulation

1. Central nervous system (CNS)

The most important part of the brain that processes signals from periphery regarding nutritional status is the hypothalamus, especially the morphologically defined areas such as the lateral hypothalamic area (LHA), ventromedial hypothalamus (VMH), arcuate nucleus (ARC), paraventricular hypothalamus (PVH), perifornical area (PFA) and dorsomedial hypothalamus (DMH). Besides hypothalamus, nucleus tractus solitarii (NTS) also plays an important role in appetite regulation.

LHA was defined as the hunger promoting feeding centre that includes neurons that produce orexigenic peptides. VMH was defined as the satiety centre, which when stimulated suppresses the desire for food. Lesions in this area have been shown to elicit rapid excessive food ingestion (hyperphagia) and abnormal body weight (7). It secretes specific anorexigenic and orexigenic neuropeptides and expresses high levels of VMH brain-derived neurotrophic factor (BDNF). VMH lesion-type hyperphagia and obesity could also be accomplished by other defects (8).

The ARC is one of the highly important areas for receiving signals from the periphery regarding appetite control in the CNS. It is closely connected to the median eminence (ME), which is not entirely protected by the blood-brain barrier (9), therefore enabling ARC to be accessible for satiety or starvation circulating signals of energy balance, such as insulin and leptin. The main neurotrasmitters in the ARC are pro-opiomelanocortin (POMC), yielding the melanocortin MSH as a cleavage product and cocaine and amphetamine-regulated transcript (CART), which both inhibit food intake (10). Furthermore, it was shown that PVH is very important for feeding behavior. Lesions in PVH cause hyperphagia in rats (7) and if an orexigenic signaling molecule (NPY, GAL, orexins, GABA, opioids, norepinephrine and epinephrine) is microinjected into PVH, it stimulates food intake (8). In feeding, it is involved in the opposite manner to LHA. PFA is one of the most sensitive areas for NPY stimulation of feeding. DMH has a role in the modulation of food intake. Lesion studies of this region resulted in hyperphagia and obesity, although not to the same degree as lesions in the VMN (8). Reward pathways that involve complex interaction between several signaling systems such as the dopaminergic and endocannabinoid system are also important. Food intake stimulated by central and peripheral administration of endocannabinoids is believed to be mediated through the cannabinoid receptor type 1 (CB1), which co-localizes with CART, MCH and orexin peptides in the hypothalamus. Opioids are important in the reward circuitry, as mice lacking either β -endorphin or enkephalin do not respond to the reinforcing property of food regardless of palatability (10).

On the other hand, the NTS as the visceral sensory part of the brainstem receives sensory inputs from the larynx, intestinal and respiratory tracts, heart, large blood vessels and taste buds. The NTS and the reward pathways mainly in nucleus accumbens (NAc) are involved in the control of energy intake. The NTS is in close contact with the area postrema (AP) which has an incomplete blood brain barrier. Therefore NTS is like the ARC, able to respond to peripheral circulating signals as well as receiving vagal afferents signals from the gastrointestinal (GI) tract and afferent signals from the glossopharyngeal nerves. Several connections between the hypothalamus and the NTS exist, including the glucagon-like peptide 1 (GLP-1) neuronal circuit, which is believed to be of major importance in the signaling circuit of the brainstem (11).

2. Neuropeptides in CNS

Various neuropeptides are involved in mediating signals regarding energy balance, such as neuropeptide Y (NPY), melanocortins, as well as cocaine and amphetamine-regulated transcript (CART). A review of examples is presented in Table 1.

	Location	Neuropeptide	Effect (neuropeptide)	Receptor	Effect (receptor)	Refe- rence
NPY	- ARC - autonomic nervous system	- 36 amino acids with many tyrosine residues - structurally related to PP and PYY	 regulation of energy balance increases food intake and proportion of energy stored as fat decreases physical activity role in memory and learning epilepsy anorexia nervosa 	 7TM receptors: NPY₁₋₅ (brain), NPY₆ (orexigenic) Gα_i coupled combined signaling through these receptors and other unknown receptors 	- role in eating disorders (obesity)	(10, 12- 17)
Melano- cortins	- ARC - NTS	 peptide hormones cleavage products of proopiomelanocortin (POMC) in the pituitary gland different forms of melanocyte- stimulating hormone (MSH) adrenocorticotropic hormone (ACTH) 	- suppresses food intake - POMC gene or gene product mutation or abnormally processing - early-onset obesity	- melanocortin receptors MC1R - MC5R in hypothalamus - very important MC3R and MC4R - Gα _s coupled	- MC4R role in regulation of food intake and body weight - 3-5 % of the cases of high obesity in the humans are caused by MC4R mutations - early-onset obesity in humans with deletion or blockade of MC4R	(10, 18, 19)
CART	 hypothalamus pituitary endocrine cells adrenomedullary cells somatostatin cells rat antral gastrin cells 	- anorexigenic peptide - neurotransmitter	 roles in reward, feeding, stress endogenous psychostimulant regulates energy homeostasis and interacts with several central appetite circuits 	- not yet identified - <i>in vitro</i> studies show that CART binds to a specific 7TM receptor coupled to $G\alpha_{e}/G_{o}$	/	(8, 11, 20- 23)

Table 1: Overview of neuropeptides in the CNS involved in mediating s	signals regarding the energy balance

3. Peripheral Signals

Signals from the periphery importantly influence the energy status of the body, as well as the amount of fat and glucose in the blood. These signals are hormones secreted from various organs in connection with meal initiation satiety and long-term energy changes. The most important are peptide YY (PYY), pancreatic polypeptide (PP), cholecystokinin (CCK), oxyntomodulin (OXM), GLP-1, gastric inhibitory polypeptide (GIP), bombesin, leptin, adiponectin, resistin, visfatin and ghrelin. Review of examples is presented in Table 2. Ghrelin and its receptor are described in more detail below Table 2.

	Location and secretion	Neuropeptide	Effect (neuropeptide)	Receptor	Effect (receptor)	Refe- rence
РҮҮ	 esophagus stomach duodenum jejunum L cells in ileum and colon (with GLP-1) neurons brainstem concentration in the circulation increases after food ingestion and decreases on fasting obese people secrete less PYY than non-obese people released in response to feeding (presence of carbohydrates, lipids and proteins in the GI tract) 	 - 36 amino acids two forms: - PYY₁₃6, PYY₃₃₆ structurally related to NPY and PP - it crosses blood brain barrier 	 slows gastric emptying increases efficiency of digestion and nutrient absorption after meal - reduces appetite weight loss was observed after chronic peripheral administration of the peptide to mice iv administration of PYY3-36 strongly decreased food intake and weight loss in humans 	- 7TM receptors: NPY (especially for PYY ₃₃₆) - PYY ₃₃₆ preferentially binds to NPY ₂ and NPY ₅ - $G\alpha_1$ coupled	- role in the food intake and immnune response	(24-29)
РР	 - PP cells in endocrine pancreas secretion in humans is increased after a protein meal, fasting, exercise, acute hypoglycemia - secretion is decreased by somatostatin and iv application of glucose - the amount of peptide released depends upon calorie intake and the composition of a meal 	 - 36 amino acids - structurally related to PYY and NPY - does not cross the blood brain barrier 	 regulates the endocrine and exocrine pancreas secretion effects hepatic glycogen levels and GI secretions peripheral infusion of PP reduces food intake, while central administration increases food intake 	- 7TM receptors: NPY1, NPY ₂ , NPY ₄ , NPY ₅ - Gα ₁ coupled	- could mediate the orexigenic effect	(9, 30, 31)
ССК	 GI tract enterocrine I cells (jejunum, duodenum) secreted in response to the presence of nutrients in the lumen of GI tract peripheral CCK crosses the blood-brain barrier and acts directly in the CNS 	- different post- translational modification of the CCK gene product, preprocholecystokinin - e.g. CCK8, CCK33 and CCK58	 satiety hormones stimulate digestion of fat and protein and act as a hunger suppressant able to detect the presence of fat in the chime and inhibit gastric emptying and gastric acid secretion, together with mediating digestion in the duodenum stimulatory effects oppose those of ghrelin effect is dependant on the vagus nerve 	 cholecystokinin B receptor (CCK2) widely distributed in brain areas such as the NTS, AP and DMH adrenal cortex sensory fibers of the vagus nerve in the pyloric sphincter Gα_s and Gα_q coupled 	 - inhibit gastric emptying, which might be involved in the satiety effect of CCK 	(10, 29, 32, 33)

Table 2: Overview of neuropeptides in PNS that influence the energy status of the body, amount of fat and amount of blood glucose

	Location and secretion	Neuropeptide	Effect (neuropeptide)	Receptor	Effect (receptor)	Refe- rence
OXM	- intestinal L cells of ileum and other parts of intestines - co-secreted with GLP-1 and GLP-2	- 37-amino acids - group of numerous tissue-specific cleavage products of proglucagon	 satiety hormone appetite reducing effects when administered centrally or peripherally - could be used as a weight loss treatment reduces circulating ghrelin levels in humans and rodents by 44 % and 20 % 	- family B 7TM GLP-1 receptor glucagon receptor - Gα _s coupled - cAMP accumulation with almost the same potency as glucagons	 L cells in ileum and colon (with GLP-1) very important in appetite regulation and energy homeostasis one of the most interesting new targets in the management of type 2 diabetes and obesity 	(34- 38)
GLP-1	 - intestinal L cells - ileum and other parts of intestine (together with PYY and OXM) - secretion is dependent on the presence of nutrients in the lumen of the small intestine by carbohydrates, proteins and lipids - an inhibition of NPY signaling by GLP-1 and an increase by exendin is observed, indicating that the GLP-1 signal is mediated via NPY neurons 	 tissue-specific cleavage product of proglucagon biologically active forms of GLP-1 are: GLP-1-(7-37) and GLP-1-(7-36) 	 satiety hormone antihyperglycemic hormone inhibits pancreatic β-cell apoptosis - stimulates the proliferation and differentiation of insulin-secreting β-cells inhibits gastric secretion and motility decreases acute food intake when administered centrally or peripherally to rats 	 family B 7TM GLP-1, GLP-2, GLP-3 receptors central administration of exendin, a GLP-1 receptor antagonist abolishes anorectic effect of GLP-1 in OXM OXM and glucagon are biased ligands on the GLP-1 receptor G_αs coupled 	 very important in appetite regulation and energy homeostasis one of the most interesting new targets in the management of type 2 diabetes and obesity 	(34, 39- 42)
GIP	 synthesized and secreted after ingestion of fat from the K cells in the intestines mucosa of duodenum and jejunum of the GI tract transported by blood 	- secretin family of hormones	glucose-dependant insulinotropic hormone increases secretion of insulin before rise in blood glucose is observed effect on adipocytes, enhancing fatty acid synthesis and their incorporation into triglycerides in ruminants role in nutrient partitioning in milk production (lipid metabolism)	- 7TM receptors - GIP receptors - on β-cells in the pancreas - Gα _s coupled	- effect on glucocorticoid metabolism	(43, 44)
bombe- sin	- most likely secreted from the intestines	 - 14-amino acids - closest homologs: neuromedin B (NMB) and gastrin releasing peptide (GRP) 	 stimulates gastrin release from G cells causes satiety with markedly increased plasma levels after feeding together with CCK, is the source of negative feedback signals that stops eating behavior peripheral and central administration of bombesin reduces food intake 	 - 7TM receptors - bombesin receptors: BB1, BB2 and BB3 - BB3 is of great interest for pharmacological industry as a drug target - Gα_q coupled 	- regulation of endocrine processes and metabolism responsible for energy balance and adiposity	(12, 41, 45, 46)

	Location and secretion	Neuropeptide	Effect (neuropeptide)	Receptor	Effect (receptor)	Refe- rence
leptin	- fat tissue - secretion is proportional to the amount of white adipose tissue in the body	 - 16 kDa - a product of the <i>ob</i> (obesity) gene, located on chromosome 7 in humans 	 key role in regulating energy intake and its influence on appetite and metabolism signal for status of energy stores reduces appetite in response to feeding obese people develop resistance to leptin suppresses NPY expression in the brain chronic administration - reduction in food intake and body weight deficiency of the hormone has shown the opposite effect 	 leptin receptor - LEP-R; CD295 (cluster of differentiation 295) a protein in humans encoded by the <i>LEPR</i> gene 	- the absence of a leptin or LEP-R - uncontrolled food intake - severe obese phenotype - variations in the leptin receptor have been associated with obesity	(47- 52)
adipo- nectin	 fat tissue releases into blood and abundant in plasma levels are inversely correlated with body fat percentage in adults plasma concentration of adiponectin is suppressed in obesity and when insulin levels rises 	- 244-amino-acids	 modulates metabolic processes, including glucose regulation and fatty acid catabolism role in the suppression of the metabolic disorders that may result in type 2 diabetes, obesity, atherosclerosis non-alcoholic fatty liver disease (NAFLD) and is an independent risk factor for metabolic syndrome contributes to increased energy expenditure by activating AMP-activated protein kinase in liver and muscle, leading to an increase in glucose utilization and fatty-acid oxidation in these tissues 	- adiponectin receptors in the skeletal muscle (AdipoR1) and liver (AdipoR2)	- reductions in adiponectin receptors may play roles in the development of insulin resistance, type 2 diabetes, metabolic syndrome, and cardiovascular diseases that are linked to obesity	(53- 57)
resistin	 - adipose tissue of mice and rats, as well as macrophages of primates, pigs and dogs - activated by specific cytokines - a linkage to a certain type of inflammation connected with development of insulin resistance 	- cysteine-rich protein - cytokine	 - involvement with obesity and type 2 diabetes - may contribute to the mechanism of obesity-induced insulin resistance 	- resistin receptor	- role in insulin resistance and type 2 diabetes	(58)
visfatin	- adipose tissue	- adipokine - cytokine	 promotes B cell maturation inhibits neutrophil apoptosis acts as an insulin mimetic by binding t insulin receptors and thereby stimulating glucose uptake 	- might bind to insulin receptor	/	(59)

	Location and secretion	Neuropeptide	Effect (neuropeptide)	Receptor	Effect (receptor)	Refe- rence
ghrelin	 - X/A like cells in the oxyntic glands of the gastric fundus mucosa in the pre-meal situation (plasma levels decrease 80% after removal of stomach) - small intestine - kidney - immune system - placenta - gonads - pituitary - adrenal cortex - lung - hypothalamus - pancreas - collocalize with glucagons in X cells of pancreas or beta cells of pancreas or in E cells of pancreas 	- 28 amino acids - chromoseome 3p - 3 intrones - encodes 511 base pair cDNA	 only peripheral hormone that stimulates food intake promotes adiposity if administered peripherally effect on secretion of growth hormone 	- 7TM receptor: ghrelin receptor (ghrR) - NPY/AGRP neuron in ARC - Gα _q coupled	- mRNA found in hypothalamus, pituitary gland, pancreas, adrenal gland, spleen, myocardium, vagal nerve	(60- 69)

Ghrelin and ghrelin receptor (ghrR)

The gastric hormone ghrelin was identified as the endogenous ligand for the former orphan receptor, the growth hormone secretagoue receptor (GHS- R_{ia}), later named ghrelin receptor (ghrR) (6). Pairing of the hormone to the receptor 20 years after its cloning made the discovery of ghrelin an example of reverse pharmacology. Both names for the receptor are still in use; however, in November 2005, the name "ghrelin receptor (ghrR)" was officially established by the International Union of Pharmacology Committee on Receptor Nomenclature and Drug Classification (NC-IUPHAR) (70).

Different natural isoforms of ghrelin exist. One of them is isoform des-Gln 14-ghrelin, which has the same activity as ghrelin; however, it is present in much lower amounts in the serum (71). Interestingly, it was shown that the entire sequence of ghrelin is not necessary to exert its activity, therefore this isoform and other short peptides are very important. Short peptides encompassing the first four or five residues of ghrelin were capable of activating ghrR at about the same efficiency as the full length ghrelin in calcium mobilization assays *in vitro*. 80-90% of circulating ghrelin is in the non-acylated form which is not able to bind to and activate ghrR (6).

Ghrelin is able to reach the brain through the areas where the blood brain barrier is incomplete. A minor quantity of ghrelin is produced within the hypothalamus in neurons adjacent to the third ventricle and between the VMH, DMH, PVH and ARC, therefore ghrelin modulates the neurotransmission and interacts pre- and post-synaptically with NPY/ AGRP, POMC and CRH circuits (72). Ghrelin transduces signals to hypothalamic regulatory nuclei that control energy homeostasis. NPY/AGRP neurons of the ARC are major targets for ghrelin. In the hypothalamus, ghrelin increases the firing rate and induces increased expression and release of NPY and AGRP. Ghrelin also seems to exert important actions on energy expenditure, both through ghrR on adipocytes and through modulation of thermogenesis. Therefore it has been suggested that the high constitutive signaling activity of the ghrR could serve as a signaling set-point in the control of appetite and energy expenditure, where it would counteract a large number of inhibitory hormones and neurotransmitters such as leptin and insulin (73, 74). Besides its actions through the hormonal route, ghrelin is also believed to exert its actions through the vagus nerve. Ghrelin produced in the stomach could conduct some of its orexigenic signals via the vagus nerve which signals to areas of the brain very much involved in appetite regulation. It is interesting to note that the stomach may play an important role not only in digestion, but also in pituitary growth hormone release and central feeding regulation (75).

The main effect of ghrelin is observed regarding food intake and metabolism. Ghrelin secretion is

regulated by many factors. In humans, ghrelin level changes during the day. High level is detected during fasting, with a peak before meal initiation, and then it drops after food intake (74). In humans the rise in ghrelin is also associated with feeling hungry (76). Ghrelin levels decline to basal within an hour after food intake (74), proportional to the load of ingested calories (77). In rats, ghrelin is released in a pulsatile manner with a regularity of about 2 episodes per hour. Ghrelin levels change in response to acute changes in nutritional status but chronic changes also have an influence on the plasma ghrelin level. Fasting plasma ghrelin levels are lower in obese research participants compared to normal weight participants, while those with low BMI (patients suffering from anorexia nervosa or carchexia) have an increased ghrelin secretion (76). Ghrelin level is decreased by oral glucose load, after secretion and/or expression of OXM, PYY³⁻³⁶, administration of somatostatin and its natural analogue cortistatin and agonist/antagonist binding of cholinergic muscarinic receptors (67). Levels of ghrelin are increased by energy restriction, thyroid hormones, testosterone, parasympathetic activity, leptin, and low BMI. The importance of ghrelin in the metabolism has not been entirely clarified. Despite all important actions, deletions of ghrelin or ghrR do not have any major effect on food intake and body weight (8).

As reported in several publications on ghrelin and ghrR, ghrelin plays an important role in appetite stimulation and increase of food intake. Chronic administration of ghrelin to lean rats is followed by an increase in food intake and bodyweight (78); however, the increase in fat mass following ghrelin administration is not caused only by increased food intake. When administering ghrelin twice a day to rodents in a period of four days, it was observed that body weight increases due to reduced fat utilization and fat deposition becomes independent of food intake, suggesting a role for ghrelin in lipid metabolism (79). Therefore, ghrelin may stimulate food intake and at the same time induce adiposity by reducing the use of fat as an energy source (79), which was also supported by calorimetric studies. In humans it was shown, that iv administration of ghrelin increases appetite and food intake in normal weight volunteers (80) and increases food intake in patients with cancer-related anorexia by more than 30% (81, 82).

Besides the effect of ghrelin administered either orally or iv, growth hormone (GH) -releasing effect of iv administered ghrelin in pharmacological doses is also described in humans and animals. Co-administration of ghrelin and GHRH has a significant synergistic effect on GH secretion, indicating that they act via different mechanisms (83). The GH-releasing effect of ghrelin varies with age. It increases at puberty. The rise in estrogens at this time leads to an increase in the expression of the ghrR, which is probably responsible for the GH increment. When reaching adulthood, the level reaches a plateau and declines during further aging (67). In both humans and animals, ghrelin was reported to act as a functional antagonist to the GHRH hormone somatostatin (67). In families with naturally occurring ghrR mutation (Ala204Glu), it leads to selective loss of constitutive activity of the ghrR, but does not affect ghrelin affinity, potency, or efficacy to ghrR. Furthermore, a tendency for developing obesity around the time of puberty was observed (84).

Besides appetite stimulating effect, some other effects of ghrelin have also been observed. It was shown that ghrelin stimulates the release of ACTH and prolactin and consequently increases cortisol levels in humans (85). It has also been reported to cause anxiogenic behavior in humans, creating a possible link between the main place of ghrelin production- the stomach and the brain (stress/anxiety). Furthermore, sleeping patterns have been reported to be affected by ghrelin treatment. In rats it improved memory retention (76). Chronic administration of a ghrelin mimic to old mice restored IGF-I levels and stimulated growth and differentiation of the thymus with an increase in the production of T-cells (86). The GI functions of ghrelin in rats are reported to be a slight increase in acid secretion, ileal peristalsis and modulation of gastric motility (76). Ghrelin also has an influence on the cardiovascular system; improving cardiac contractility and performance in humans following ghrelin injection and counter inflammation in these tissues (67). Decreased ghrelin levels are independently associated with type 2 diabetes mellitus, insulin sensitivity and secretion in humans (87), except in lean humans with type 2 diabetes mellitus.

It is interesting to note that normally, a reciprocal relationship exists between leptin and ghrelin levels. It is suggested that leptin plays a regulatory role in the secretion of ghrelin (88), by having a role in the circadian and ultradian rhythmic fluctuation of ghrelin secretion. This makes it tempting to believe that there might be a feedback mechanism between ghrelin and leptin such that ghrelin also has an effect on leptin secretion. However, according to experiments performed by Sun et al., which included ghrelin knockout and ghrR knockout mice, this is not the case (89). No difference in the postprandial leptin levels was observed, indicating that ghrelin does not effect leptin secretion. Considerable evidence points to leptin being a regulator of ghrelin levels and an important part of the hypothalamic appetite regulating circuit. In March 2010, researchers reported that mice with type 1 diabetes treated with leptin alone or in conjunction with insulin, had better values of blood sugar and cholesterol than mice with type 1diabetes treated with insulin alone, raising the prospect of a new treatment for diabetes (52).

Ghrelin transmits its signal through the ghrelin receptor (ghrR) (40). GhrR has features characteristic of family A 7TM receptors, including conserved cysteine residues in the top of TM-III and in extracellular loop 2, conserved prolines, E/DRY motif, polar transmembrane residues and several potential sites for posttranslational modifications (N-linked glycosylation and phosphorylation) (90). GhrR signals through $G\alpha_{\alpha/11}$, which results in accumulation of inositol (1,4,5)-triphosphate (IP) and Ca²⁺ release. Ghrelin has also been shown to activate the MAP kinase cascade and the PI3-K/AKT pathway (91). GhrR pharmacology started with the synthesis of analogs long before the discovery of its natural ligand (6, 92). It belongs to a subfamily of receptors for peptide hormones and neuropeptides. Besides ghrelin, the family includes receptors for motilin (previously orphan receptor GPR38), neurotensin, neuromedin U (NMU) and orphan receptor GPR39 (40). GhrR is encoded by a single gene found at chromosomal location 3q26.2. As a result of alternate processing of pre-mRNA, two different variants of the ghrR exist (93). The full length ghrR contains 366 amino acids (ghR R1a). The other splice variant, designated ghR R1b, consists of 289 amino acids and has 5 TM regions. Unlike ghR R1a, GhR R1b is not activated by ghrelin or a synthetic analogue such as the ghrR agonist hexarelin or non-peptidyl GHS such as MK-0677 (94).

Importantly it was discovered (84) that the ghrR signals with ~ 50% of its maximal activity in the absence of its ligand. GhrR has constitutive activity by demonstrating a gene dose dependent but ligand independent increase in IP accumulation (85). It was suggested that the constitutive activity of the ghrR could function as an appetite set-point against the signals from the many anorexigenic hormones such as leptin and insulin. The level of the recep-

tor could then be regulated by ghrelin and perhaps by an endogenous inverse agonist, which would act by decreasing the constitutive activity of the receptor. Biological effects different from those seen after ghrelin treatment have been observed following administration with non-acylated ghrelin and synthetic homologs, suggesting the existence of one or more receptors in the ghrelin receptor family that have not yet been identified (60).

Conclusions

The discovery that ghrelin is one of the most powerful orexigenic and adipogenic agents known in mammalian physiology, triggered the exploitation of ghrR antagonists and/or inverse agonists that can be used to treat obesity (60). GhrR antagonist could be used to block the stimulation generated by ghrelin and thereby reduce meal size. A specific inverse agonist of ghrR would lower constitutive activity of the ghrR and thereby lower the set point of signaling from the receptor between meals. This could increase the sensitivity to the multiple inhibitory signals, e.g. leptin, insulin and PYY₃₋₃₆ and consequently eliminate between-meal food intake. The advantages of combining ghrR antagonist and a ghrR inverse agonist in an anti-obesity drug would be that the antagonist could block the effect of the increase in plasma ghrelin seen before meals (40).

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References

1. Bell CG, Walley AJ, Froguel P. The genetics of human obesity. Nat Rev Genet 2005; 6: 221-34.

2. Mathieu P, Lemieux I, Després JP. Obesity, inflammation, and cardiovascular risk. Clin Pharmacol Ther 2010; 87: 407-16.

3. Lund EM. Prevalence and risk factors for obesity in adult dogs from private US veterinary practices. Intern J Appl Res Vet Med 2006; 4: 177-86.

4. Nijland ML, Stam F, Seidell JC. Overweight in dogs, but not in cats, is related to overweight in their owners. Public Health Nutr 2010; 13: 102-6.

5. Arora S, Anubhuti. Role of neuropeptides in appetite regulation and obesity: a review. Neuropeptides 2006; 40: 375-401.

6. Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormonereleasing acylated peptide from stomach. Nature 1999; 402: 656-60.

7. Kishi T, Elmquist JK. Body weight is regulated by the brain: a link between feeding and emotion. Mol Psychiatry 2005; 10: 132-46.

8. Kalra SP, Dube MG, Pu SP, Xu B, Horvath TL, Kalra PS. Interacting appetite-regulating pathways in the hypothalamic regulation of body weight. Endocr Rev 1999; 20: 68-100.

9. Small CJ, Bloom SR. Gut hormones as peripheral anti obesity targets. Curr Drug Targets CNS Neurol Disord 2004; 3: 379-88.

10. Wynne K, Stanley S, McGowan B, Bloom S. Appetite control. J Endocrinol 2005; 184: 291-318.

11. Stanley BG, Kyrkouli SE, Lampert S, Leibowitz SF. Neuropeptide Y chronically injected into the hypothalamus: a powerful neurochemical inducer of hyperphagia and obesity. Peptides 1986; 7: 1189-92.

12. Cone RD, Cowley MA, Butler AA, Fan W, Marks DL, Low MJ. The arcuate nucleus as a conduit for diverse signals relevant to energy homeostasis. Int J Obes Relat Metab Disord 2001; 25 (Suppl 5): S63-7.

13. Colmers WF, El Bahh B. Neuropeptide Y and epilepsy. Epilepsy Curr 2003; 3: 53-8.

14. Larhammar D. Structural diversity of receptors for neuropeptide Y, peptide YY and pancreatic polypeptide. Regul Pept 1996; 65: 165-74.

15. Kaye WH, Berrettini W, Gwirtsman H, George DT. Altered cerebrospinal fluid neuropeptide Y and peptide YY immunoreactivity in anorexia and bulimia nervosa. Arch Gen Psychiatr 1990; 47: 548-56.

16. Takenoya F, Kageyama H, Shiba K, Date Y, Nakazato M, Shioda S. Neuropeptide W: a key player in the homeostatic regulation of feeding and energy metabolism? Ann N Y Acad Sci 2010; 1200: 162-9.

17. Haas DA, George SR. Neuropeptide Y-induced effects on hypothalamic corticotropin-releasing factor content and release are dependent on noradrenergic/adrenergic neurotransmission. Brain Res 1989; 498: 333-8.

18. Gantz I, Fong TM. The melanocortin system. Am J Physiol Endocrinol Metab 2003; 284: E468-74.

19. Barsh GS, Farooqi IS, O'Rahilly S. Genetics of body-weight regulation. Nature 2000; 404: 644-51.

20. Douglass J, Daoud S. Characterization of the human cDNA and genomic DNA encoding CART:

a cocaine- and amphetamine-regulated transcript. Gene 1996; 169: 241-5.

21. Maletínská L, Maixnerová J, Matyšková R, et al. Synergistic effect of CART (cocaine- and amphetamine-regulated transcript) peptide and cholecystokinin on food intake regulation in lean mice. BMC Neurosci 2008; 9: 101-11.

22. Wierup N, Kuhar M, Nilsson BO, Mulder H, Ekblad E, Sundler F. Cocaine- and amphetamineregulated transcript (CART) is expressed in several islet cell types during rat development. J Histochem Cytochem 2004; 52: 169-77.

23. Lakatos A, Prinster S, Vicentic A, Hall RA, Kuhar MJ. Cocaine- and amphetamine-regulated transcript (CART) peptide activates the extracellular signal-regulated kinase (ERK) pathway in AtT20 cells via putative G-protein coupled receptors. Neurosci Lett 2005; 384: 198-202.

24. Sandström O, El-Salhy M. Ontogeny and the effect of aging on pancreatic polypeptide and peptide YY. Peptides 2002; 23: 2263-7.

25. Taylor IL. Distribution and release of peptide YY in dog measured by specific radioimmunoassay. Gastroenterology 1985; 88: 731-7.

26. Glavas MM, Grayson BE, Allen SE, et al. Characterization of brainstem peptide YY (PYY) neurons. J Comp Neurol 2008; 506: 194-210.

27. Liu CD, Aloia T, Adrian TE, et al. Peptide YY: a potential proabsorptive hormone for the treatment of malabsorptive disorders. Am Surg 1996; 62: 232-6.

28. Bednarek MA, Feighner SD, Pong SS, et al. Structure-function studies on the new growth hormone-releasing peptide, ghrelin: minimal sequence of ghrelin necessary for activation of growth hormone secretagogue receptor 1a. J Med Chem 2000; 43: 4370-6.

29. Barazzoni R, Bosutti A, Stebel M, et al. Ghrelin regulates mitochondrial-lipid metabolism gene expression and tissue fat distribution favoring triglyceride deposition in liver but not skeletal muscle. Am J Physiol Endocrinol Metab 2005; 288: E228-35.

30. Nonaka N, Shioda S, Niehoff ML, Banks WA. Characterization of blood-brain barrier permeability to PYY3-36 in the mouse. J Pharmacol Exp Ther 2003; 306: 948-53.

31. Lundell I, Blomqvist AG, Berglund MM, et al. Cloning of a human receptor of the NPY receptor family with high affinity for pancreatic polypeptide and peptide YY. J Biol Chem 1995; 270: 29123-8.

32. Kissin I, Bright CA, Bradley EL. Acute tolerance to continuously infused alfentanil: the role of cholecystokinin and N-methyl-d-aspartate-nitric oxide systems. Anesth Analg 2000; 91: 110-6.

33. Harikumar KG, Clain J, Pinon DI, Dong M, Miller LJ. Distinct molecular mechanisms for agonist peptide binding to types A and B cholecystokinin receptors demonstrated using fluorescence spectroscopy. J Biol Chem 2005; 280: 1044-50.

34. Mayo KE, Miller LJ, Bataille D, et al. International Union of Pharmacology. XXXV. The glucagon receptor family. Pharmacol Rev 2003; 55: 167-94.

35. Wynne K, Park AJ, Small CJ, et al. Oxyntomodulin increases energy expenditure in addition to decreasing energy intake in overweight and obese humans: a randomised controlled trial oxyntomodulin and energy balance. Int J Obes 2006; 30: 1729-36.

36. Cohen MA, Ellis SM, Le Roux CW, et al. Bloom oxyntomodulin suppresses appetite and reduces food intake in humans. J Clin Endocrinol Metabol 2003; 88: 4696-701.

37. Dakin CL, Gunn I, Small CJ, et al. Oxyntomodulin inhibits food intake in the rat. Endocrinology 2001; 142: 4244-50.

38. Bataille D, Coudray AM, Carlqvist M, Rosselin G, Mutt V. Isolation of glucagon-37 (bioactive enteroglucagon/oxyntomodulin) from porcine jejunoileum: isolation of the peptide. FEBS Lett 1982; 146: 73-8.

39. Sinclair EM, Drucker DJ. Proglucagon-derived peptides: mechanisms of action and therapeutic potential. Physiology 2005; 20: 357-65.

40. Holst B, Schwartz TW. Ghrelin receptor mutations - too little height and too much hunger. J Clin Invest 2006; 116: 637-41.

41. Holst JJ. Treatment of type 2 diabetes mellitus with agonists of the GLP-1 receptor or DPP-IV inhibitors. Expert Opin Emerg Drugs 2004; 9: 155-66.

42. Jorgensen R, Kubale V, Vrecl M, Schwartz TW, Elling CE. Oxyntomodulin differentially affects glucagon-like peptide-1 receptor beta-arrestin recruitment and signaling through Galpha(s). J Pharmacol Exp Ther 2007; 322: 148-54.

43. Meier JJ, Nauck MA. Glucagon-like peptide 1(GLP-1) in biology and pathology. Diabetes Metab Res Rev 2005; 21: 91-117.

44. Yamada Y, Seino Y. Physiology of GIP - a lesson from GIP receptor knockout mice. Horm Metab Res 2004; 36: 771-4.

45. Ladenheim EE, Moore KA, Salorio CF. Characterization of bombesin binding sites in the rat stomach. Eur J Pharmacol 1997; 319: 245-51. 46. Gbahou F, Holst B, Schwartz TW. Molecular basis for agonism in the BB3 receptor: an epitope located on the interface of transmembrane-III, -VI, and -VII. J Pharmacol Exp Ther 2010; 333: 51-9.

47. Green ED, Maffei M, Braden VV, et al. The human obese (OB) gene: RNA expression pattern and mapping on the physical, cytogenetic, and genetic maps of chromosome 7. Genome Res 1995; 5: 5-12.

48. Casanueva FF, Dieguez C. Leptin and ghrelin: what is the impact on pituitary function? Rev Endocr Metab Disord 2005; 6: 39-45.

49. Majdič G. Leptin and its company (molecular mechanisms of appetite regulation, energy consumption and fat deposits) = Leptin in njegova družina (molekularni mehanizmi urejanja apetita, porabe energije in nalaganja maščob). Slov Vet Res 2000; 37: 181-9.

50. Schwartz MW, Woods SC, Porte D, Seeley RJ, Baskin DG. Central nervous system control of food intake. Nature 2000; 404: 661-71.

51. Tartaglia LA, Dembski M, Weng X, et al. Identification and expression cloning of a leptin receptor, OB-R. Cell 1995; 83; 1263-71.

52. Wang M, Chen L, Clark GO. Leptin therapy in insulin-deficient type I diabetes. PNAS 2010; 107: 4813-9.

53. Diez JJ, Iglesias P. The role of the novel adipocyte-derived hormone adiponectin in human disease. Eur J Endocrinol, 2003; 148: 293-300.

54. Ukkola O, Santaniemi M. Adiponectin: a link between excess adiposity and associated comorbidities? J Mol Med 2002; 80: 696-702.

55. Renaldi O, Pramono B, Sinorita H, Purnomo LB, Asdie RH, Asdie AH. Hypoadiponectinemia: a risk factor for metabolic syndrome. Acta Med Indones 2009; 41: 20-4.

56. Yamauchi T, Kamon J, Minokoshi Y. Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. Nat Med 2002; 8: 1288-95.

57. Meier U, Gressner AM. Endocrine regulation of energy metabolism: review of pathobiochemical and clinical chemical aspects of leptin, ghrelin, adiponectin, and resistin. Clin Chem 2004; 50: 1511-25.

58. Badman MK, Flier JS. The gut and energy balance: visceral allies in the obesity wars. Science 2005; 307: 1909-14.

59. Fukuhara A, Matsuda M, Nishizawa M, et al. Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. Science 2005; 307: 426-30. 60. van der Lely AJ, Tschop M, Heiman ML, Ghigo E. Biological, physiological, pathophysiological, and pharmacological aspects of ghrelin. Endocr Rev 2004; 25: 426-57.

61. de Ligt RA, Kourounakis AP, Ijzerman AP. Inverse agonism at G protein-coupled receptors: (patho)physiological relevance and implications for drug discovery. Br J Pharmacol 2000; 130: 1-12.

62. Wierup N, Yang S, McEvilly RJ, Mulder H, Sundler F. Ghrelin is expressed in a novel endocrine cell type in developing rat islets and inhibits insulin secretion from INS-1 (832/13) cells. J Histochem Cytochem 2004; 52: 301-10.

63. Date Y, Kojima M, Hosoda H, et al. Ghrelin, a novel growth hormone-releasing acylated peptide, is synthesized in a distinct endocrine cell type in the gastrointestinal tracts of rats and humans. Endocrinol 2000; 141: 4255-61.

64. Tsubone T, Masaki T, Katsuragi I, Tanaka K, Kakuma T, Yoshimatsu H. Ghrelin regulates adiposity in white adipose tissue and UCP1 mRNA expression in brown adipose tissue in mice. Regul Pept 2005; 130: 97-103.

65. Wang L, Saint-Pierre DH, Tache Y. Peripheral ghrelin selectively increases Fos expression in neuropeptide Y - synthesizing neurons in mouse hypothalamic arcuate nucleus. Neurosci Lett 2002; 325: 47-51.

66. Wren AM, Small CJ, Ward HL, et al. The novel hypothalamic peptide ghrelin stimulates food intake and growth hormone secretion. Endocrinology 2000; 141: 4325-8.

67. Korbonits M, Goldstone AP, Gueorguiev M, Grossman AB. Ghrelin-a hormone with multiple functions. Front Neuroendocrinol 2004; 25: 27-68.

68. Gnanapavan S, Kola B, Bustin SA, et al. The tissue distribution of the mRNA of ghrelin and subtypes of its receptor, GHS-R, in humans. J Clin Endocrinol Metab 2002; 87: 2988.

69. Sakata I, Yamazaki M, Inoue K, Hayashi Y, Kangawa K, Sakai T. Growth hormone secretagogue receptor expression in the cells of the stomach-projected afferent nerve in the rat nodose ganglion. Neurosci Lett 2003; 342: 183-6.

70. Davenport AP, Bonner TI, Foord SM, et al. International Union of Pharmacology. LVI. Ghrelin receptor nomenclature, distribution, and function. Pharmacol Rev 2005; 57: 541-6.

71. Hosoda H, Kojima M, Matsuo H, Kangawa K. Purification and characterization of rat des-Gln14-Ghrelin, a second endogenous ligand for the growth hormone secretagogue receptor. J Biol Chem 2000; 275: 21995-2000.

72. Silva Elipe MV, Bednarek MA, Gao YD. 1H NMR structural analysis of human ghrelin and its six truncated analogs. Biopolymers 2001; 59: 489-501.

73. Holst B, Cygankiewicz A, Jensen TH, Ankersen M, Schwartz TW. High constitutive signaling of the ghrelin receptor - identification of a potent inverse agonist. Mol Endocrinol 2003; 17: 2201-10.

74. Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE, Weigle DS. A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. Diabetes 2001; 50: 1714-9.

75. Hosoda H, Kojima M, Kangawa K. Ghrelin and the regulation of food intake and energy balance. Mol Interv 2002; 2: 494-503.

76. Ghigo E, Broglio F, Arvat E, Maccario M, Papotti M, Muccioli G. Ghrelin: more than a natural GH secretagogue and/or an orexigenic factor. Clin Endocrinol 2005; 62: 1-17.

77. Callahan HS, Cummings DE, Pepe MS, Breen PA, Matthys CC, Weigle DS. Postprandial suppression of plasma ghrelin level is proportional to ingested caloric load but does not predict intermeal interval in humans. J Clin Endocrinol Metab 2004; 89: 1319-24.

78. Nakazato M, Murakami N, Date Y, et al. A role for ghrelin in the central regulation of feeding. Nature 2001; 409: 194-8.

79. Barsh GS, Farooqi IS, O'Rahilly S. Genetics of body-weight regulation. Nature 2000; 404: 644-51.

80. Wren AM, Small CJ, Abbott CR, et al. Ghrelin causes hyperphagia and obesity in rats. Diabetes 2001; 50: 2540-7.

81. Wren AM, Seal LJ, Cohen MA, et al. Ghrelin enhances appetite and increases food intake in humans. J Clin Endocrinol Metab 2001; 86: 5992.

82. Nijenhuis WA, Oosterom J, Adan RA. AgRP(83-132) acts as an inverse agonist on the human-melanocortin-4 receptor. Mol Endocrinol 2001; 15: 164-71.

83. Holst B, Holliday ND, Bach A, Elling CE, Cox HM, Schwartz TW. Common structural basis for constitutive activity of the ghrelin receptor family. J Biol Chem 2004; 279: 53806-17.

84. Hataya Y, Akamizu T, Takaya K, et al. A low dose of ghrelin stimulates growth hormone (GH) release synergistically with GH-releasing hormone in humans. J Clin Endocrinol Metab 2001; 86: 4552.

85. Arvat E, Maccario M, Di VL, et al. Endocrine activities of ghrelin, a natural growth hormone

secretagogue (GHS), in humans: comparison and interactions with hexarelin, a nonnatural peptidyl GHS, and GH-releasing hormone. J Clin Endocrinol Metab 2001; 86: 1169-74.

86. Smith RG, Jiang H, Sun Y. Developments in ghrelin biology and potential clinical relevance. Trends Endocrinol Metab 2005; 16: 436-42.

87. Poykko SM, Kellokoski E, Horkko S, Kauma H, Kesaniemi YA, Ukkola O. Low plasma ghrelin is associated with insulin resistance, hypertension, and the prevalence of type 2 diabetes. Diabetes 2003; 52: 2546-53.

88. Kalra SP, Ueno N, Kalra PS. Stimulation of appetite by ghrelin is regulated by leptin restraint: peripheral and central sites of action. J Nutr 2005; 135: 1331-5.

89. SunY, Ahmed S, Smith RG. Deletion of ghrelin impairs neither growth nor appetite. Mol Cell Biol 2003; 23: 7973-81.

90. Kojima M, Kangawa K. Ghrelin: structure and function. Physiol Rev 2005; 85: 495-522.

91. Camina JP. Cell biology of the ghrelin receptor. J Neuroendocrinol 2006: 18: 65-76.

92. Shiiya T, Nakazato M, Mizuta M, et al. Plasma ghrelin levels in lean and obese humans and the effect of glucose on ghrelin secretion. J Clin Endocrinol Metab 2002; 87:240-4.

93. Howard AD, Feighner SD, Cully DF, et al. A receptor in pituitary and hypothalamus that functions in growth hormone release. Science 1996; 273: 974-7.

94. Kojima M, Hosoda H, Kangawa K. Purification and distribution of ghrelin: the natural endogenous ligand for the growth hormone secretagogue receptor. Horm Res 2001; 56 (Suppl 1): 93-7.

UREJANJE APETITA IN DEBELOST: POUDAREK NA GRELINU IN GRELINSKEM RECEPTORJU

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Povzetek: Debelost je eden od prevladujočih vzrokov smrtnosti po svetu, kljub temu da bi jo lahko preventivno zmanjšali. Epidemska razsežnost debelosti je postala podlaga za veliko zanimanje farmacevtske industrije za razvoj zdravila, ki bi vplivalo na zmanjšanje apetita. Z zmanjšanjem apetita bi se zmanjšal tudi skupen vnos energije. Ideja je enostavna, vendar so hormonski sistem in mehanizmi, ki urejajo vnos energij, e zelo kompleksni in ne omogočajo enostavnega razvoaj takšnega zdravila. V centralnem živčnem sistemu so peptidi in hormoni, ki se izločajo na periferiji in v centralnem živčnem sistemu, vključeni v vnos hrane in urejanje telesne teže. Vsi so tesno povezani. Uravnavajo jih hipotalamus, deli možganskega debla ter sistem nagrajevanja na osnovi kognitivnih zaznavanj in različnih sporočil telesnih tekočin ter živčevja glede prehrambenega položaja. Veliko nevropeptidov, kot so nevropeptid Y (NPY), melanokortini, prepis, uravnavan s kokainom in amfetaminom (CART), peptid YY (PYY), pankreasni polipeptid (PP), holecistokinin (CCK), oksintomodulin (OXM), glukagonu podobni peptid 1 (GLP-1), od glukoze odvisni inzulinotropni hormon (GIP), bombezin, leptin ter eden zadnjih odkritih in zato manj poznanih - grelin, imajo pomembno vlogo pri kratkoročnem in dolgoročnem urejanju vnosa hrane. Pregledni članek povzema zapletene dejavnike, vključene v urejanje apetita in prehranjevanja, od različnih področij možganov, še posebej hipotalamusa, do povezave med ključnimi centralnimi in perifernimi peptidi in hormoni, s poudarkom na grelinu in receptorju za grelin, skupaj z njihovimi potencialnimi pomeni kot tarče za zdravljenje debelosti. Receptor za grelin je sprva pritegnil pozornost farmacevtske industrije zaradi svoje pomembne vloge pri sproščanju rastnega hormona. Njegova apetit spodbujajoča funkcija je spremenila grelinski receptor (ghrR) v tarčo farmacevtske industrije za razvoj zdravila proti debelosti.

Ključne besede: urejanje apetita; debelost; grelin; receptor za grelin