SUPERVISED AUTOMATIC HISTOGRAM CLUSTERING AND WATERSHED SEGMENTATION. APPLICATION TO MICROSCOPIC MEDICAL COLOR IMAGES

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ABSTRACT

In this paper, an approach to the segmentation of microscopic color images is addressed, and applied to medical images. The approach combines a clustering method and a region growing method. Each color plane is segmented independently relying on a watershed based clustering of the plane histogram. The marginal segmentation maps intersect in a label concordance map. The latter map is simplified based on the assumption that the color planes are correlated. This produces a simplified label concordance map containing labeled and unlabeled pixels. The formers are used as an image of seeds for a color watershed. This fast and robust segmentation scheme is applied to several types of medical images.

Keywords: clustering, color, histogram, segmentation, watershed.

INTRODUCTION

Color image segmentation refers to the partitioning of a multi-channel image into meaningful objects. The various approaches to color image segmentation reported in the literature may be roughly classified into a few categories: clustering methods (Park et al., 1998), edge-based methods (Carron and Lambert, 1994), region growing methods (Trémeau and Borel, 1998), and variational methods (Kichenassamy et al., 1995). In the context of segmenting medical color images, we propose to combine two types of methods: clustering- and region growing- methods. Both methods are based on the watershed principle (Soille, 1999). Because clustering methods relying on the 3D information of a color histogram may be timeconsuming (Géraud et al., 2001), methods based on the multi-thresholding of the color planes may be preferred (Soille, 2000; Albiol et al., 2001). Herein, we intend to use the merging of segmentation maps obtained by treating the color planes independently (marginal approach to color segmentation) as the starting point for a color watershed. Thus, our strategy consists in the following three steps: 1) clustering of each color plane, 2) merging of the resulting segmentation maps, and 3) color watershed growing. The paper is structured consistently with the abovementioned segmentation scheme.

GRAY-SCALE IMAGE PLANES CLUSTERING

Each color plane is here clustered independently by splitting its (gray-scale) histogram in several sections corresponding to representative classes of pixels in the image. We assume that the number N_c of classes of pixels is a priori known. Furthermore, we suppose that the representative sections (or clusters) of each color plane histogram are located around its main modes. It is thus reduced to find the threshold values dissecting the histograms in a finite number of clusters. This is here achieved by computing the 1D watershed of the complement (H^c) of each color plane histogram (H) as reported in Soille (1999).

Using all the minima of H^c as seeds for the *1D* watershed would result in a severe over-clustering. In this respect, only the so-called *h*-minima constrain the watershed growing. In the process of extracting the h-minima, *h* is experimentally defined as:

$$h = \left| \sigma(H) - \sqrt{max(H)} \right|, \qquad (1)$$

where σ and *max* denote the standard deviation and the maximum of the histogram H, respectively. Once the *h*-minima are extracted (by a morphological reconstruction (Serra, 1982)), the watershed operation divides the histogram in a finite number of sections. The class of every pixel *p* in the corresponding grayscale image is defined by the label value of the section *p* belongs to in the clustered histogram.

The essential drawback of the method is the number of resulting sections, generally larger than the desired number of classes N_c . Adjacent sections of the histogram are accordingly merged according to the following strategy. A 1D adjacency graph of the clustered histogram is first built, and the section showing the smallest number of pixels in the histogram is identified. The current smallest section is then merged with that one of its two adjacent sections (S_i) minimizing the following quantity:

$$\begin{array}{c}
\overset{u(S_i)}{\longleftarrow} & H(k) \\
\overset{k=l(S_i)}{\underbrace{u(S_i) - l(S_i)}}, \\
\end{array}$$
(2)

where a histogram section is denoted by S_i , H(k) is the number of pixels of gray-level k, and $l(S_i)$ (resp. $u(S_i)$) is the lowest (resp. highest) gray level in section S_i . The adjacency graph is updated each time two sections merge and the algorithm iterates till the number of sections in the histogram is equal to N_c . Fig. 1 illustrates various steps of the clustering in 3 sections of a color plane histogram (the original color image associated to Fig. 1 is displayed in Fig. 5a). We can note in Fig. 1 that the three final sections correspond to the main three peaks of the histogram and that the zone of influence of each peak is well defined. Of course, the merging step is not performed when the number of the sections resulting from the watershed is lower than N_c .

MERGING OF THE SEGMENTED COLOR PLANES

Concordance of the marginal labels

In color images, the three color planes express a priori different types of information. In this respect, the spatial regions exhibited with a marginal segmentation (every color plane clustered independently) may differ depending on the color plane which is considered. This is illustrated by Figs. 2a-c showing for each color plane (R, G, B) the segmentation map obtained through the clustering approach of the previous section with $N_c=3$. The main objective of color segmentation is to take advantage of the information available with several color channels. Along that line, one approach to color image segmentation consists in blending the information carried by the segmentation maps of the different channels (Figs. 2a-c). The resulting map is often referred to as the so-called label concordance map (Richards and Jia, 1999; Kurugollu et al., 2001). The label concordance map (Fig. 2d), denoted by J, is here derived as the *intersection* of the three marginal segmentation maps.



Fig. 1. Illustration of the clustering of an histogram. (a) Over-clustered histogram, (b) The histogram after the merging operation.



Fig. 2. The marginal segmentation maps of the microscopic image shown in Fig. 5a and the resulting label concordance map. In (a) to (c), colors express the different classes of pixels (a) Segmentation map of R, (b) Segmentation map of G. (c) Segmentation map of B. (d) Label concordance map; colors correspond to a specific combination of marginal labels.

SIMPLIFICATION OF THE CONCORDANCE MAP

Due to the intersection, the number of clusters in the image J may be higher than the desired number of clusters N_c . Herein, we propose to use a simplification operator in order to reduce the number of clusters associated to the concordance map. We assume that the representative clusters of a color image shall be those associated to the largest areas in the concordance map. In other words, the representative clusters shall be defined as the combinations of marginal labels showing the largest overlaps across the marginal segmentation maps. It is here the right point to emphasize that the more the three marginal segmentation maps overlap, the more the color planes are correlated (and the smaller is the number of labels in the concordance map). Fig. 2d illustrates this point on an experimental basis. The simplification of the label concordance map (Fig. 2d) consists accordingly in keeping only the most representative combinations of labels. Altogether, such simplification is based on the idea that the color planes are correlated. In fact this is often the case for microscopic images. In particular, an experimental study carried on many hematological and serous microscopic images revealed that the correlation rate between the three color planes is commonly higher than 90% in the RGB color space.

To perform the simplification, the histogram of the concordance map J is first computed. Each peak in the histogram corresponds to the number of pixels of a class in image J. The simplification operator may be defined as follows. Let M be the set of all the local maxima in the histogram H_i of J:

$$M = \{ \sup_{q} H_{j}(x) \mid q \in V(x) \}$$

$$(4)$$

where sup is the supremum and V(x) is the neighborhood of a current concordance label (denoted by x) (here, the two adjacent concordance labels in the histogram). $M(N_c)$ is the set of concordance labels associated to the N_c greatest maxima of the histogram in the set M. The simplification may so be expressed as: if, for a pixel p of J, $J(p) \notin M(N_c)$ then J(p)=0else J(p) remains unchanged. The resulting simplified image includes N_c+I classes. The labeled pixels are associated to the N_c most representative clusters of the concordance image. Pixels labeled as 0 (referred to as unlabeled hereafter) are considered as uncertain pixels since their label in the concordance map does not relate to the N_c representative clusters. Fig. 3 presents the simplified concordance map I_f of Fig. 2d. Unlabeled pixels are expressed in black.



Fig. 3. The label concordance map after simplification: black pixels correspond to unlabeled pixels. The labeled pixels correspond to the three different classes of pixels (background, cytoplasm and nucleus) and will be used as markers for the color watershed.

COLOR WATERSHED

Instead of using classical probability values (Bloch, 1996) to assess the basic assignment of unlabeled pixels (black pixels in Fig. 3) to a cluster, we propose to use a color watershed. The seeds for the watershed are provided by the labeled connected components of the simplified concordance image I_{f} . This is necessary to let the regions grow independently one of the other.

Aggregation function

The color watershed used in this paper is defined according to a specific aggregation function. The aggregation (or distance) function estimates the certainty that a pixel p belongs to an adjacent region R_i (considered as a single connected component). It is based on two pieces of information (local and global) describing the spatial information of the image (Adams and Bischof, 1994). This aggregation function can be formally defined. Let $\overline{I(R_i)}$ denote the mean color vector of the region R_i of image I in the color space $C_1C_2C_3$. Furthermore, let vector I(p) represent the color of a pixel p and $\nabla I(p)$ its color gradient (the gradient is processed using DiZenzo's (1986) definition). The aggregation function is then expressed as (Lezoray *et al.*, 2000):

$$f(p,R_i) = (I - \alpha) || I(R_i) - I(p) || + \alpha || \nabla I(p) ||.$$
(5)

This function combines local information (the modulus of the color gradient), and global information in the form of a (statistical) comparison between the color of a pixel p and the mean color of its neighbor region R_i (performed with the Euclidean distance). During the growing process, every time a pixel is added to a region R_i , the mean color of the region is updated. Both the color image and the gradient image are normalized before the watershed growing in order to have the same range of values. In Eq. (5), the blending coefficient α allows to tune the influence of the local and global criteria during the growing process. The watershed uses the value of the aggregation function f as the ordering of a classical FIFO priority queue (Vincent and Soille, 1991). Pixels are assigned to the region corresponding to the smallest value of f as reported in Salembier et al. (1996).

Estimation of alpha

The α parameter was introduced to control the relative influence of the global and local criteria. α may be fixed once for all according to some a priori knowledge on the images (Lezoray et al., 2000). However, an adaptable segmentation modifying the value of α across the iterations seems more suitable. The initial value of α is θ , and α is so allowed to evolve during the growing. At each iteration k, the quantity $V_k = \sum f(p, R_i)$ is computed where V_k relates to the set of unlabeled pixels p currently processed (the set of processed pixels is a subset of all the unlabeled pixels and concerns only unlabeled pixels currently adjacent to a given region). Therefore, V_0 gives an initial value relating to the unlabeled pixels of the image adjacent to the markers at the first iteration. Since the initial value of α is 0, V_0 quantifies the initial global similarity between unlabeled pixels and region markers. If the global similarity V_k is small, the unlabeled pixels are close to the regions so that α shall be small to essentially take into account the global information. On the contrary, if V_k increases, α shall increase to give more weight to the local information. To fulfill these requirements α is defined by:

$\alpha = V_k / V_0.$

It is not desirable that α varies abruptly across successive iterations. In this respect, the value retained for the next iteration k+1 is the mean of all the previous values of α (including the new computed one). This enables a smoother evolution of α . Table 1 gives the mean value of α during the growing for 10 different medical images (cytology, hematology, skin, brain, histology, etc.) One shall note the quite different mean values of α listed in table 1. The proposed automatic evolution of α obviously avoids the difficulty to tune this parameter according to some a priori knowledge (which is very difficult to express numerically). Fig. 4 illustrates the variations of α across the iterations of the watershed region growing process applied to Fig. 3. The visual evaluation of the segmented images confirms that the self-adaptable segmentation gives the more accurate results.

Table 1. Mean value of α for several medical images.

Image	α	N_c
I1	0.054	2
I2	0.037	2
13	0.212	2
I4	0.191	2
15	0.207	2
I6	0.078	3
I7	0.051	3
18	0.216	4
19	0.188	4
I10	0.216	4



Fig. 4. *An example of variation of the alpha parameter* (*obtained on the watershed growing of Fig. 3*).

RESULTS

The color watershed is performed using the result of the merging as markers. Fig. 5b presents the final regions resulting from the growing process with α automatically estimated (Fig. 4). Fig. 5a presents the original image (RGB color space). One will note that the segmentation is very close to the expected solution. All the unlabeled pixels (Fig. 3) have been assigned to a region of the image; the spatial information provided by the color watershed algorithm allows to complete the segmentation.



a)



b)

Fig. 5. (a) The original microscopic image: cells from serous cytology stained by the Papanicolaou protocol (b) Final regions obtained after the watershed growing initialized on the labeled regions of the simplified concordance map (Fig. 3).

The influence of the choice of the color space onto the current color image segmentation method, has been studied. Indeed, the correlation between the color planes is an important property in our approach to color segmentation. Because the strength of the correlation may vary with the color space, its choice may influence the output of the method. To address the later point, we used a database of 63 hematological color images available on the internet network for *e-learning* and *e-diagnosis* purposes. The point of the segmentation exercise is here to extract the cells in the images, and moreover to provide a classification of the cells into $N_c=3$ classes.

To compare the different color spaces for the image database (mention *RGB*, *XYZ*, *HSL*, *111213*, *YIQ*, *YUV*, *Ych1Ch2*, *YcbCr*, $L^*u^*v^*$, $L^*a^*b^*$), a Mean Squared Error (MSE) is computed between the original image and a *colorized* image where each pixel carries the mean color value of the connected component it belongs after the segmentation; the lower the *MSE*, the better the segmentation. Fig. 6 shows that some color spaces seem more adapted than others. In this respect, the color space relevance shall be studied and quantified with an appropriate measure for each considered application.



Fig. 6. The MSE obtained for several color spaces.

Once the appropriate color space has been identified, the segmentation scheme provides a set of connected and labeled regions. Fig. 7b presents the simplified concordance map obtained for the color hematological image displayed in Fig. 7a. Fig. 7c presents the final regions resulting from the growing process. The class label of each independent region can be obtained as the majority one encountered within the area of its final segmentation.







c)

Fig. 7. The segmentation of an hematological image: (a) the original image (b) the label concordance map after simplification into three different classes of pixels (c) Final regions obtained after the watershed growing.

CONCLUSION

A supervised method for segmenting microscopic color images has been discussed and illustrated. In a first step, for each color plane, distinct segmentation maps are extracted by a marginal approach. Their intersection, the so-called label concordance map, is then simplified. The simplification relies on the assumption that the color planes are correlated and leaves some pixels unlabeled. A self-adaptable region growing method is last performed to obtain the final regions. The proposed method is reliable and fast and provides a classification of each region among a predefined number of classes. Further research concerns the automatic determination of the number of classes to be used in the segmentation for indexing color medical images.

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