

Expression of *IRT1* gene in barley seedlings under zinc deficiency at optimal and low temperatures

Natalia KAZNINA ^{1,2}, Natalia REPKINA ¹, Yulia BATOVA ¹, Alexandr TITOV ¹

Received December 17, 2020; accepted October 21, 2021.
Delo je prispelo 17. decembra 2020, sprejeto 21. oktobra 2021

Expression of *IRT1* gene in barley seedlings under zinc deficiency at optimal and low temperatures

Abstract: The deficiency or excess of zinc (Zn) cause negative effect on plant metabolism and development. Therefore, plants have established a tightly controlled system, including protein transporters to balance the uptake and utilization of metal ions. In this study, the relative expression of *HvIRT1* gene, encoding the transmembrane protein IRT1 was analyzed in shoots and roots of barley (*Hordeum vulgare* 'Nur') under zinc deficiency at optimal (22 °C) or low (4 °C) temperatures. The Zn deficiency (0 μmol) caused an increase in *HvIRT1* gene expression under both optimal temperature condition and cold. Although, the difference in mRNA content of *HvIRT1* gene in roots of barley under optimal and low temperature was not observe. However, the *HvIRT1* expression in leaves was higher at optimal temperature compare with cold condition. Moreover, long-term (7 days) of low temperature influence along with zinc deficiency leads to a significant decrease in the amount of *HvIRT1* transcripts in leaves, that corresponds to a decrease of photosynthesis rate and biomass accumulation. Overall, these findings suggest that *HvIRT1* gene play an important role in plant's response to zinc deficiency under optimal temperatures condition as well as at cold.

Key words: *IRT1*; *Hordeum vulgare*; zinc deficiency; low temperatures

Izražanje *IRT1* gena v sejankah ječmena ob pomanjkanju cinka pri optimalnih in nizkih temperaturah

Izvleček: Pomanjkanje ali prebitek cinka (Zn) povzročata negativne učinke na presnovo in razvoj rastlin. Zaradi tega so rastline razvile dobro nadzorovan sistem, vključno s proteinskimi transporterji za uravnavanje privzema in porabe kovinskih ionov. V raziskavi je bilo analizirano izražanje *HvIRT1* gena, ki kodira transmembranski protein *IRT1* v poganjkih in koreninah ječmena (*Hordeum vulgare* 'Nur') ob pomankanju cinka pri optimalni (22 °C) in nizki (4 °C) temperaturi. Pomanjkanje cinka (0 μmol) je povzročilo povečano izražanje *HvIRT1* gena pri optimalni kot pri nizki temperaturi. Razlika v vsebnosti mRNK *HvIRT1* gena v koreninah ječmena v optimalnih razmerah in pri nizki temperaturi ni bila opažena, a kljub temu je bilo izražanje gena *HvIRT1* v listih večje pri optimalni temperaturi v primerjavi s hladnimi rastnimi razmerami. Daljša izpostavitve (7 dni) nizki temperature je ob pomanjkanju cinka povzročila značilno zmanjšanje transkriptov *HvIRT1* v listih, kar ustreza upadu fotosinteze in akumulacije biomase. Ta odkritja nakazujejo, da igra *HvIRT1* gen pomembno vlogo pri odzivu rastlin na pomanjkanje cinka tako v optimalnih razmerah kot pri nizkih temperaturah.

Gljučne besede: *IRT1*; *Hordeum vulgare*; pomanjkanje cinka; nizke temperature

¹ Institute of Biology, Karelian Research Centre, Russian Academy of Sciences, Petrozavodsk, Russia

² Corresponding author, e-mail: kaznina@krc.karelia.ru

1 INTRODUCTION

Zn deficiency has been recognized as an important factor affecting crop production. Cell transmembrane proteins from ZIP family (*zinc-iron-regulated transporter*) play an important role in providing plants of the necessary amount of zinc (Pedas et al., 2008; Lee and An, 2009; Yamunarani et al., 2013). The IRT1 (*iron-regulated transporter1*) proteins, belonging to the ZIP family, were firstly discovered in cereals. ZIP proteins are able to transport various divalent cations, such as Fe²⁺, Zn²⁺, Cu²⁺, Mn²⁺ from the rhizosphere through the plasma membrane into the cytoplasm of root cells, as well as from xylem vessels in leaf mesophyll cells (Palmer and Guerinot, 2009). It has been reported that zinc deficiency leads to increase in the activity of IRT1 protein and *IRT1* gene expression in parallel with high accumulation of Zn in roots and shoots of rice, maize and *Arabidopsis* (Ishimaru, 2006; Pedas et al., 2008; Yamunarani et al., 2013; Kabir et al., 2017 etc.). Therefore, IRT1 protein play essential role in Zn uptake, translocation and storage of Zn in plant cells especially under Zn deficiency. Although, most of the evidence from these studies was performed on plants under optimal temperature conditions, however, in nature plants are often exposed to low temperatures during the growing season, that caused in decrease in supply of nutrients to root cells, that result in their deficiency in plants (Hacisalihoglu et al., 2001). Perhaps this effect may be associated with a decrease in the activity of transport proteins (Guerinot, 2000; Hacisalihoglu et al., 2001). Despite this data, the effect of cold on *IRT1* gene expression and IRT1 protein activity is still unclear. Some reports demonstrated that the IRT1 protein activity regulated at the both translation and transcription level (Shin et al., 2013; Brumbarova et al., 2015). According these findings, we studied the expression of the *IRT1* gene in the roots and leaves of barley under zinc deficiency at optimal and low temperatures.

2 MATERIALS AND METHODS

2.1 PLANT MATERIAL AND GROWTH CONDITIONS

Seeds of barley (*Hordeum vulgare* 'Nur') were purchased from the Tula Research Institute of Agriculture, Tula, Russia. Seedlings were cultivated in a growth chamber with 14 h photoperiod, a photo-synthetic photon flux density of 180 $\mu\text{mol m}^{-2} \text{s}^{-1}$, a temperature of 22 °C and a relative humidity of 60 - 70 % on Hoagland-Arnon nutrient solution (pH 6.2 to 6.4) with optimal (variant Zn 2 $\mu\text{mol} + 22$ °C) zinc content or its deficiency (variant Zn

0 $\mu\text{mol} + 22$ °C). Seven-day-old seedlings (initial level) were separated. One part of the plants of both variants was exposed to low temperature (4 °C) (variants Zn 2 $\mu\text{mol} + 4$ °C and Zn 0 $\mu\text{mol} + 4$ °C), and the other part was left under the optimal temperature during 7 days. All parameters were measured at day 0 (initial level) and 1, 3, 7 days after treatments.

2.2 BIOMASS AND NET PHOTOSYNTHETIC RATE DETERMINATION

For biomass determination plants were collected, their shoots and roots separated and dried in an oven at 85 °C for 24 h. The net photosynthetic rate (P_N) was measured during a day using portable photosynthesis system HCM-1000 (Walz, Effeltrich, Germany).

2.3 GENE EXPRESSION

The expression pattern of *HvIRT1* gene in leaves and roots was monitored by real-time PCR. Frozen roots and leaf tissues were homogenized with liquid nitrogen. Total RNA was extracted using a TRizol reagent (*Evrogen*, Moscow, Russia) as instructed by them manufacturer. The total RNA was treated with RNase free DNase (*Syntol*, Moscow, Russia) to remove genomic DNA. The purity of RNA samples and their concentrations were determined spectrophotometrically (*SmartSpecPlus*, *Bio-Rad*, Hercules, USA): samples with A260/A280 ratios within 1.8 - 2.0 were used for further analysis. The total RNA (1 μg) was reverse-transcribed using a *MMLV* RT kit (*Evrogen*) following the supplier's recommendations. Real-time quantitative PCR was performed using the *iCycler iQ* detection system (*Bio-Rad*). Analyzes were performed using a *SYBR Green* PCR kit (*Evrogen*). The PCR conditions consisted of denaturation at 95 °C for 5 min followed by 45 cycles of denaturation at 95 °C for 15 s, annealing at 56 °C for 30s, and extension at 72 °C for 45 s. A dissociation curve was generated at the end of each PCR cycle to verify that a single product was amplified using *iCycler iQ*. To minimize sample variations, mRNA expression of a target gene was normalized relative to the expression of a housekeeping gene *actin*. The mRNA content of target gene (*HvIRT1*) were quantified in comparison to the *actin* by the $\Delta\Delta\text{Ct}$ method (Livak and Schmittgen, 2001). Primers were designed (using the *Primer Design* program): *HvActin* (U21907) ATGTTTTTTTTCCAGACG (direct) and ATCCAAGCCAACCCAAGT (reverse), *HvIRT1* (EU54802) GTGCTTCCACCAGATGTTTGGAG (direct) and GGATGCCGACGACGATGA (reverse).

2.4 STATISTICAL ANALYSIS

All data are presented as means \pm standard errors (SEs) from at least three independent replicates. Significant differences between variants and relative to the initial level were calculated by two-way analysis of variance (ANOVA) using Microsoft Excel 2010. Student's *t*-test was applied to compare statistical significance at level of $p < 0.05$.

3 RESULTS AND DISCUSSION

Table 1 shows the effect of Zn deficiency under optimal and low temperatures on plant dry mass (DM) accumulation and net photosynthetic rate (P_N). Under optimal temperature conditions Zn deficiency did not significantly affect the DM and P_N parameters compared with plants grown with optimal Zn concentration. However, after 7 days Zn deficiency (Zn 0 $\mu\text{mol} + 22^\circ\text{C}$) caused a slight reduction in root DM compared with variant Zn 2 $\mu\text{mol} + 22^\circ\text{C}$.

Despite the Zn concentration the low temperature leads to reduce DM accumulation and P_N parameter. Although, after 7 days Zn deficiency in combination with low temperature (Zn 0 $\mu\text{mol} + 4^\circ\text{C}$) leads to a significant decrease in root DM and photosynthesis activity compared with variant Zn 2 $\mu\text{mol} + 4^\circ\text{C}$.

Under optimal growth conditions (variant Zn 2 $\mu\text{mol} + 22^\circ\text{C}$) the transcript level of *HvIRT1* gene gradually increases in roots and leaves of barley during 7 days (Fig.). While, at the initial level, the *HvIRT1* gene expression was 3-fold higher in roots of seedlings grown with Zn deficiency (Zn 0 $\mu\text{mol} + 22^\circ\text{C}$). Further *HvIRT1* gene mRNA content in variant Zn 0 $\mu\text{mol} + 22^\circ\text{C}$ slightly increased compared with variant Zn 2 $\mu\text{mol} + 22^\circ\text{C}$. At the initial level there was no significant difference in mRNA content of *HvIRT1* gene in leaves between Zn 2 $\mu\text{mol} + 22^\circ\text{C}$ and variant Zn 0 $\mu\text{mol} + 22^\circ\text{C}$ variants (Fig.). However, leaves of barley exposed to Zn deficiency (Zn 0 $\mu\text{mol} + 22^\circ\text{C}$) showed higher *HvIRT1* mRNA content within 1 day and a slight decrease on the seventh day compared with Zn 2 $\mu\text{mol} + 22^\circ\text{C}$ variant.

Low temperature caused an increase in *HvIRT1* gene expression in roots along with time of exposure in both variants (Zn 2 $\mu\text{mol} + 4^\circ\text{C}$ and Zn 0 $\mu\text{mol} + 4^\circ\text{C}$) (Fig.). However, on 7th day of experiment the amount of *HvIRT1* gene transcripts in roots of barley variant Zn 0 $\mu\text{mol} + 4^\circ\text{C}$ were greater than in variant Zn 2 $\mu\text{mol} + 4^\circ\text{C}$. Low temperature resulted in *HvIRT1* gene transcript accumulation in leaves of barley variant Zn 2 $\mu\text{mol} + 4^\circ\text{C}$. After 1 day of cold impact the *HvIRT1* gene expression was 4-fold higher in variant Zn 2 $\mu\text{mol} + 4^\circ\text{C}$ com-

pared with initial level and 10-fold higher on the seventh day of experiment. In variant Zn 0 $\mu\text{mol} + 4^\circ\text{C}$ the *HvIRT1* gene mRNA content increased after 3 days of exposure to low temperature and significantly decreased on the seventh day of experiment.

In general, our results demonstrated a high tolerance of barley 'Nur' to Zn deficiency. It was shown that under conditions of zinc deficiency, the accumulation of biomass and photosynthetic activity remained stable until the end of the experiment. Similar data were described previously (Hajiboland and Beiramzadeh, 2008; Kabir et al., 2017). The capability of plants to grow under Zn deficiency mostly depends on metal transporters activity, including protein IRT1 (Suzuki et al., 2012; Yamunarani et al., 2013; Kabir et al., 2017). Zn deficiency under optimal temperature leads to an increase in *HvIRT1* transcript amount in roots and leaves of barley that resulted in activation of transport metal ions in cells and kept growth and photosynthetic activity. This also supports the fact that after 7 days the gene expression of *HvIRT1* decreased along with photosynthetic activity. The negative effect of Zn deficiency on the photosynthesis process was described previously in *Oryza sativa* L. (Hajiboland and Beiramzadeh, 2008), *Zea mays* L. (Liu et al., 2016), *Sorghum bicolor* (L.) Moench (Li et al., 2013), however the transporter protein activity was not studied.

There are fragmentary data about the influence of low temperature on metal transporters activity. The negative effect of 0°C temperature on gene expression, encoding protein transporters ZIP1 and ZIP3 was described previously (Grotz et al., 1998). We have also shown the increase in *HvIRT1* gene expression in barley under chilling (Kaznina et al., 2019). It is supposed that due to the negative influence of chilling on nutrient transport into cells the increase in *HvIRT1* gene expression in roots that we reported can be a result of the requirement of mesophyll cells of leaves in nutrients that are necessary for the photosynthesis process. There are no data about the activity of transporter proteins under chilling and Zn deficiency conditions. Thus, considering the results described above for barley under low temperature and Zn deficiency during 7 days in leaves the *HvIRT1* mRNA content significantly decreased. It seems to be a result of a decrease in the requirement for nutrients caused by a slowdown of photosynthetic activity and growth of seedlings under stress conditions. Additionally, it can be a result of interruption in signal transduction from leaves to roots that was demonstrated previously in plants under chilling and optimal nutrient level (Giehl et al., 2009; Romera et al., 2011).

Table 1: The effect of zinc deficiency on the root and shoot dry biomass and photosynthesis rate of barley plants 'Nur' at optimum (22 °C) and low (4 °C) temperatures

Variant	Time, days			
	0 (initial point)	1	3	7
Dry root biomass, mg				
Zn 2+22°C	5.51 ± 0.55 aA	6.72 ± 0.48 aA	7.29 ± 0.42 bA	8.21 ± 0.60 bA
Zn 0 +22°C	5.10 ± 0.45 aA	6.51 ± 0.41 aA	6.92 ± 0.38 bA	7.96 ± 0.46 bA
Zn 2+4°C	5.51 ± 0.55 aA	6.20 ± 0.47 aA	6.26 ± 0.48 aB	7.36 ± 0.54 bAB
Zn 0+4°C	5.10 ± 0.45 aA	6.13 ± 0.29 aA	6.23 ± 0.35 aB	6.36 ± 0.46 aB
Dry shoot biomass, mg				
Zn 2+ 22°C	20.13 ± 1.88 aA	20.89 ± 1.21 aA	27.21 ± 1.48 bA	31.32 ± 2.41 bA
Zn 0 +22°C	18.99 ± 1.89 aA	19.51 ± 1.09 aA	24.06 ± 1.62 bA	32.13 ± 1.32 bA
Zn2+4°C	20.13 ± 1.88 aA	20.00 ± 1.51 aA	20.87 ± 1.50 aB	24.77 ± 1.39 bB
Zn 0 +4°C	18.99 ± 1.89 aA	19.63 ± 1.16 aB	18.42 ± 0.95 aC	19.28 ± 1.21 aC
Net photosynthetic rate, $\mu\text{mol CO}_2 \text{ m}^{-2} \cdot \text{s}^{-1}$				
Zn 2+ 22°C	6.82 ± 0.16 aA	7.77 ± 0.17 bA	6.54 ± 0.50 aA	6.53 ± 0.10 aA
Zn 0 +22°C	6.95 ± 0.14 aA	7.69 ± 0.14 bA	6.46 ± 0.49 aA	6.27 ± 0.12 aA
Zn2+4°C	6.82 ± 0.16 aA	4.65 ± 0.20 bB	4.86 ± 0.12 bB	4.16 ± 0.09 cB
Zn 0 +4°C	6.95 ± 0.14 aA	4.83 ± 0.12 bB	4.35 ± 0.15 bB	3.82 ± 0.17 cC

Different lowercase letters indicate significant differences in columns (between variants), uppercase letters - in rows (relative to the initial level) ($p < 0.05$). Values perform mean \pm SE (n = 10)

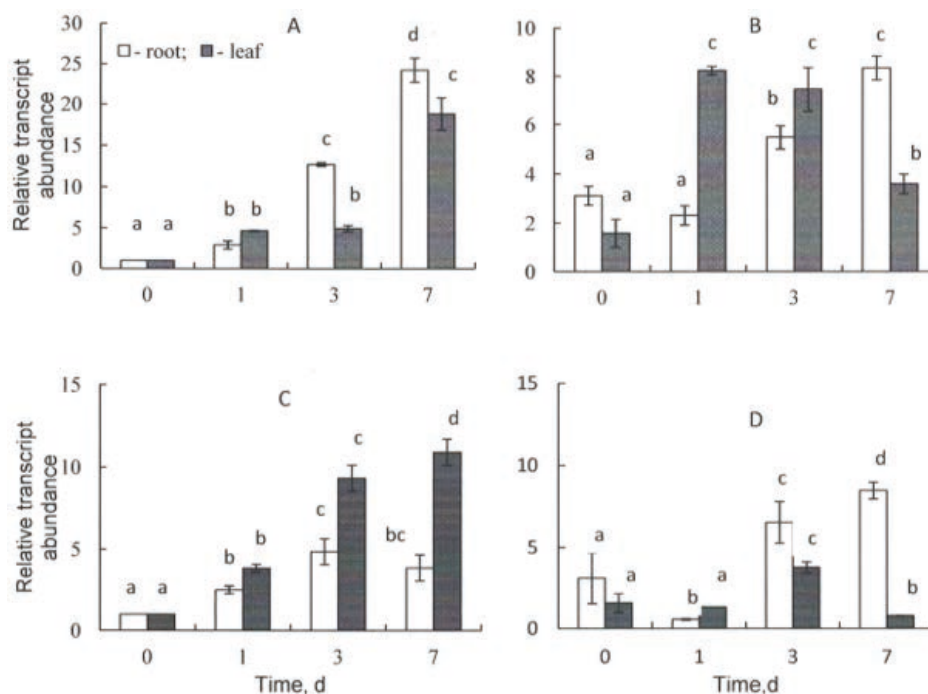


Figure 1: The effect of zinc optimum (a, c) and zinc deficiency (b, d) on *HvIRT1* gene transcription in the roots and leaves of barley plants 'Nur' at 22 °C (a, b) and 4 °C (c, d). Different lowercase letters indicate significant differences relative to the initial level ($p < 0.05$)

4 CONCLUSIONS

According to our results it was shown that Zn deficiency caused in increase in *HvIRT1* gene expression in leaves of barley not only under optimal temperature as was shown in another reports but also under cold. For the first time we demonstrated that long-term exposure (7 days) to low temperature leads to significant decrease in *HvIRT1* mRNA amount in leaves in parallel to slow down in photosynthetic activity and growth. Taken together, the results presented here illustrate the participation of the *HvIRT1* gene in adaptation to Zn deficiency under optimal and low temperatures conditions.

5 ACKNOWLEDGEMENTS

This research was carried out using the equipment of the Core Facility of the Karelian Research Centre of the Russian Academy of Sciences and under state order (No. 0218-2019-0074).

6 REFERENCES

- Brumbarova, T, Bauer, P, Ivanov, R (2015). Molecular mechanisms governing *Arabidopsis* iron uptake. *Trends in Plant Science*, 20(2), 124–133. <https://doi.org/10.1016/j.tplants.2014.11.004>
- Giehl, R. F. H., Meda, A.R., von Wirén, N. (2009). Moving up, down, and everywhere: signaling of micronutrients in plants. *Current Opinion in Plant Biology*, 12, 320–327. <https://doi.org/10.1016/j.pbi.2009.04.006>
- Grotz,N., Fox, T., Connolly, E., Park, W., Guerinot, M.L., Eide, D. (1998). Identification of a family of zinc transporter genes from *Arabidopsis* that respond to zinc deficiency. *Proceedings of the National. Academy of Sciences USA*, 95, 7220–7224. <https://doi.org/10.1073/pnas.95.12.7220>
- Guerinot,M.L.(2000). The ZIP family of metal transporters. *Biochimica et Biophysica Acta*, 1465, 190–198. [https://doi.org/10.1016/S0005-2736\(00\)00138-3](https://doi.org/10.1016/S0005-2736(00)00138-3)
- Hacisalihoglu,G., Hart, J.J., Kochian, L.V. (2001). High- and low- affinity zinc transport systems and their possible role in zinc efficiency in bread wheat. *Plant Physiology*, 125, 456–463. <https://doi.org/10.1104/pp.125.1.456>
- Hajiboland, R., Beiramzadeh, N. (2008). Growth, gas exchange and function of antioxidant defense system in two contrasting rice genotypes under Zn and Fe deficiency and hypoxia. *Acta Bioogica. Szegediensis*, 52(2), 283–294. <http://www.sci.u-szeged.hu/ABS>
- Ishimaru, Y., Suzuki, M., Tsukamoto, T., et al. (2006). Rice plants take up iron as an Fe³⁺-phytosiderophore and as Fe²⁺. *The Plant Journal*, 45, 335–346. <https://doi.org/10.1111/j.1365-313X.2005.02624.x>
- Kabir, A. H., Hossain, M.M., Khatun, M.A., Sarkar, M.R., Haid-er, S.A. (2017). Biochemical and molecular mechanisms associated with Zn deficiency tolerance and signaling in rice (*Oryza sativa* L.). *Journal of Plant Interactions*, 12(1), 447–456. <https://doi.org/10.1080/17429145.2017.1392626>
- Kaznina, N.M., Titov, A.F., Repkina, N.S., Batova, Yu.V. (2019). Effect of zinc excess and low temperature on the *IRT1* gene expression in the roots and leaves of barley. *Doklady Biochemistry and Biophysics*, 48, 264–268. <https://doi.org/10.1134/S1607672919040057>
- Lee, S., A., G. (2009). Over-expression of OsIRT1 leads to increased iron and zinc accumulations in rice. *Plant, Cell and Environment*, 32, 408–416. <https://doi.org/10.1111/j.1365-3040.2009.01935.x>
- Li, Y., Zhang, Y., Shi, D., Kiu, X., Qin, J, Ge, Q. ... Xu, J. (2013). Spatial-temporal analysis of zinc homeostasis reveals the response mechanisms to acute zinc deficiency in *Sorghum bicolor*. *New Phytologist*, 200, 1102–1115. <https://doi.org/10.1111/nph.12434>
- Liu, H., Gan, W., Renge, Z., Zhao, P. (2016). Effects of zinc fertilizer rate and application method on photosynthetic characteristics and grain yield of summer maize. *Journal of Soil Science and Plant Nutrition*, 16(2), 550–662. <https://doi.org/10.4067/S0718-95162016005000045>
- Livak, K.J., Schmittgen, T.D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2^{-ΔΔCt} method. *Methods*, 25, 402–408. <https://doi.org/10.1006/meth.2001.1262>
- Palmer, G.M., Guerinot, M.L. (2009). Facing the challenges of Cu, Fe and Zn homeostasis in plants. *Nature Chemical Biology*, 5, 333–340. <https://doi.org/10.1038/nchembio.166>
- Pedas, P., Ytting, C. K., Fuglsang, A. T., Jahn, T. P., Schioerring, J. K., Hasted, S. (2008). Manganese efficiency in barley: identification and characterization of the metal ion transporter HvIRT1^[OIA]. *Plant Physiology*, 148, 455–466. <https://doi.org/10.1104/pp.108.118851>
- Romera, F. J., García, M. J., Alcántara, E., Pérez-Vicente, R. (2011). Latest findings about the interplay or auxin, ethylene and nitric oxide in the regulation of Fe deficiency responses by strategy I plants. *Plant Signaling and Behavior*, 6, 167–170. <https://doi.org/10.4161/psb.6.1.14111>
- Shin, L.-J., Lo, J.-C., Chen, G. H., Callis, J., Fu, H., Yeh, K.-C. (2013). IRT1 degradation factor1, a ring E3 ubiquitin ligase, regulates the degradation of iron-regulated transporter1 in *Arabidopsis*. *The Plant Cell*, 25, 3039–3051. <https://doi.org/10.1105/tpc.113.115212>
- Suzuki, M., Bashir, K., Inoue, H., Takahashi, M., Nakanishi, H., Nishizawa, N.K. (2012). Accumulation of starch in Zn-deficient rice. *Rice*, 59, 1–8. <https://doi.org/10.1186/1939-8433-5-9>
- Yamunarani, R., Ramegowda, V., Pavithra, J., Geetha, G., Rajashekar-Reddy, H., Udayakumar, M., Shankar, A. G. (2013). Expression of a rice Zn transporter, OsZIP1, increases Zn concentration in tobacco and finger millet transgenic plants. *Plant Biotechnology Reports*, 7, 309–319. <https://doi.org/10.1007/s11816-012-0264-x>