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Cover photo: *Sestoj subasociacije Pseudostellario-Carpinetum betuli leucojetosum aestivi*, Lijak. (See paper in 57/1, page 25)
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- 27 Nomenclature correction of the name and rank of the association *Chamaecytiso hirsuti-Quercetum petraeae* Dakskobler 2014 nom. illeg.
- 27 Nomenklturni popravek imena in ranga asociacije *Chamaecytiso hirsuti-Quercetum petraeae* Dakskobler 2014 nom. illeg.

COMMUNICATING THE VALUE OF NATURE'S SERVICES TODAY FOR TOMORROW

KOMUNICIRANJE VREDNOSTI STORITEV NARAVNEGA OKOLJA DANES ZA JUTRI

Tjaša BALOH^{1*}, Boris RANTAŠA¹

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ABSTRACT

Communicating the value of nature services today for tomorrow

To ensure appreciation of forest related nature's services, they must be presented to the public in an appropriate way. Humans are usually not willing to pay for something they consider as 'free goods'. The challenge is changing the mind-set through awareness-raising. A good communication strategy should convey the message as a tangible concern that contributes to the quality of our lives. Communicating nature's services will help understand the benefits we gain and joy and pleasure they can give us if we treat them with knowledge, respect and wisdom.

The actions taken in the past are not giving the results we need. A multi-stakeholder approach can harness the creativity from stakeholders through solution driven dialogue.

In the LIFE GENMON project (Life for Forest Genetic Monitoring System) coordinated by Slovenian Forestry Institute, the multi-stakeholder approach is used to communicate the forest related nature's services and importance of genetic resources. A clear visual and verbal communication brings the key project messages to life. To forge meaningful connections with core stakeholders it is essential to use compelling and targeted messages. Ultimately, the results will demonstrate the value of engaging supporters in co-creating communications to extend reach and impact.

Keywords: nature's services, communicating, forests, society, forest values, multi-stakeholder approach

IZVLEČEK

Komuniciranje vrednosti storitev naravnega okolja danes za jutri

Za uspešno ozaveščanje javnosti o vrednosti storitev, ki jih zagotavlja naravno okolje, morajo te biti predstavljene na ustrezen način. Ljudje običajno niso pripravljeni plačati za storitve, ki so do zdaj veljale za proste (brezplačne). Spreminjanje načina razmišljanja s pomočjo ozaveščanja je torej velik izziv. Dobra komunikacijska strategija mora posredovati sporočilo o prispevku naravnega okolja h kakovosti človeškega življenja in bivanja. Ozaveščanje in širjenje znanja o storitvah, ki jih nudi naravno okolje, nam pomaga razumeti vrednost teh storitev in veselje ter zadovoljstvo, ki nam jih ponuja okolje danes in nam jih bo tudi v prihodnosti, če ga bomo obravnavali z znanjem, spoštovanjem in modrostjo.

Ukrepi, sprejeti v preteklosti, niso dali zadovoljivih rezultatov. Z aktivnim vključevanjem deležnikov preko usmerjenega dialoga lahko pridobimo ključne informacije in kreativne rešitve, ki jih nato vključimo v komunikacijsko strategijo.

Projekt LIFE GENMON, ki ga vodi Gozdarski inštitut Slovenije, v svojo komunikacijsko in diseminacijsko strategijo vključuje splošne in ciljne skupine deležnikov, s katerimi komunicira o vrednosti storitev, ki jih nudijo gozdovi. S pomočjo jasne vizualne in verbalne komunikacije posreduje ključna sporočila, ki so prepričljiva in ciljno usmerjena (prilagojena posamezni ciljni skupini oz. posamezniku). Tako oblikuje smiselne povezave z deležniki in s ciljnimi skupinami. Rezultati prizadevanj v okviru projekta bodo pokazali dodano vrednost, ki jo prinaša vključevanje deležnikov v soustvarjanje njegovih učinkov in večjega dosega njegovih sporočil in posledično komunikacijske strategije.

Ključne besede: storitve naravnega okolja, komuniciranje, gozdovi, družba, vrednost gozda, vključevanje ključnih deležnikov

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INTRODUCTION

To ensure that the real value of nature's services is adequately appreciated, they must be presented to the public in an appropriate way. Humans are usually not willing to pay for something they take for granted today and is considered as 'free goods' (COULL & VALANTIN 2008). The challenge is changing the mind-set through awareness-raising. A good communication strategy should convey the message as a tangible concern that contributes to the quality of our lives and the lives of our children (IUCN REGIONAL OFFICE FOR PAN-EUROPE 2011). Communicating nature's services will help the younger generation to understand the benefits we gain and joy and pleasure they can give us if we treat them with knowledge, respect and wisdom (WIBORN 2013).

The actions that have been taken in the past are not giving the results we need. HOHNEN (2001) suggests that we should work together to find the right path forward. A multi-stakeholder approach can engage and harness the creativity from wide and balanced cross-section of stakeholders through solution driven dialogue (HOHNEN 2001).

"Learning to engage in dialogue means to move from hearing to listening. It means taking one step beyond fighting, beyond adversarial, conflict-based interaction... Dialogue is the foundation for finding

consensus solutions which integrate diverse views and generate the necessary commitment to implementation. It can form the basis to take us one step beyond talking towards common action" (HEMMATI et al. 2002).

HEMMATI et al. (2002) view communication as a tool to exchange views (opinions) among stakeholders in a Multi Stakeholder Platform (MSP). It includes the expression of views in combination with the understanding of views to the point of mutual understanding (HEMMATI et al. 2002).

In the LIFEGENMON (2016) project, coordinated by Slovenian Forestry Institute, the multi-stakeholder approach is used to communicate the forest related nature's services in connection to the 360-degree brand communication. A clear visual and verbal communication creates maximum awareness and brings the key project messages to life. 360-degree brand communication means showcasing the use of compelling and targeted messages, both online and offline, in an effort to forge meaningful connections with core stakeholders. Ultimately, the results will demonstrate the value of engaging supporters and champions in co-creating communications to extend reach and impact (DARI-GAN MERENDA 2013).

METHOD OF IMPLEMENTATION

The communication strategy of the LIFEGENMON project is carried out in two focused tasks. General dissemination and communication addresses the following stakeholders:

- Kindergarten children
- Pupils in primary and secondary schools
- Students at Universities
- Pedagogues
- Socially active groups (different for each country)
- Media

In task Targeted dissemination and communication the most important groups of stakeholders have been identified through dialogue between project partners:

Policy makers – key actors (the ones to convince) in the legislative process on a local, regional and European scale.

Municipalities within the region – local authorities can give necessary promotion and credibility

needed for the maximisation of dissemination impact. They are also very appropriate partners for spreading information to target areas.

Active groups (ASP – Women organisation; third university for everlasting education, recreational sports associations and others, different per project country) – through "spreading the good word", socially active groups can increase the dissemination impact if addressed appropriately.

Academic audience – academics are often opinion makers, e.g. what they say usually resonates in public and professional space.

Chamber of Commerce – the Chamber of Commerce is an important player in the legislation department. It is very important to include big business in our dissemination strategy, because no legislation or regulation get passed without their influence. We need to raise their awareness of the importance of our main dissemination strategy topics.

Professional Chambers – smaller, but more specific legislation and regulation impact. Also a base of

professional experience. We also need to raise their awareness of the importance of our main dissemination strategy topics.

NGOs (such as professional associations, mountaineering organisations, scouts, hunters associations, climbers association, UMANOTERA – SI, WWF, Birdlife and others) – NGOs are well heard opinion makers among the urban population and are traditionally inclined to support the topics of our dissemination strategy. Very important players in any legislation process if activated.

Forest owners – forest owners are the ones that do (or at least approve) the majority of the actual work done in the focus area. It is aim to reach them with the conservational aspects of our dissemination strategy.

Unions of forest workers – they need to be addressed to spread the information on genetic forest protection and the future of European forests based on genetic diversity.

Civil protection service and volunteers – they are first to be activated at any major disaster events in the areas, due to extreme weather conditions or un-appropriate forest managing. Their activities are well visible

in the public and can spread the word on the need for sustainable forest management and an early warning system for changes in forest populations.

Tourist organisations – sustainable and eco-tourism is becoming very popular. They are also traditionally inclined to support the topics of our dissemination strategy and reach a wide audience with their marketing so it makes sense to include them in our dissemination activities.

Carrying out communication activities goes beyond the narrow scope of the project to reach the highest impact through:

Connecting (in Slovenia cooperation with Slovenia Forest Service, University of Ljubljana – Biotechnical Faculty, Slovenian Academy of sciences and arts, etc.; participation at and organisation of events and conferences, trade fairs, media events etc.)

Dialogue (organisation of Workshops, receiving and evaluating feedback)

All activities are supported by media presence, web updates (Facebook, Twitter, LinkedIn, project web-site) and publications (newsletters, leaflets, teaching materials for teachers etc.).

MONITORING METHODS AND RESULTS

The impact of the activities will be monitored through lists of participants and an advanced monitoring matrix, based on questionnaires and interviews of participants at project's events. The monitoring is carried out by an internal project monitoring team, project partner Centre for information service, co-operation and development of NGOs.

The LIFE GENMON team has by end of 2016 reached over 4000 people at over 90 different events and the LIFE GENMON website had over 8300 users who viewed over 30000 pages (Source: Google analytics).

Results and progress of implementation will be presented in LIFE GENMON progress reports and scientific and professional articles.

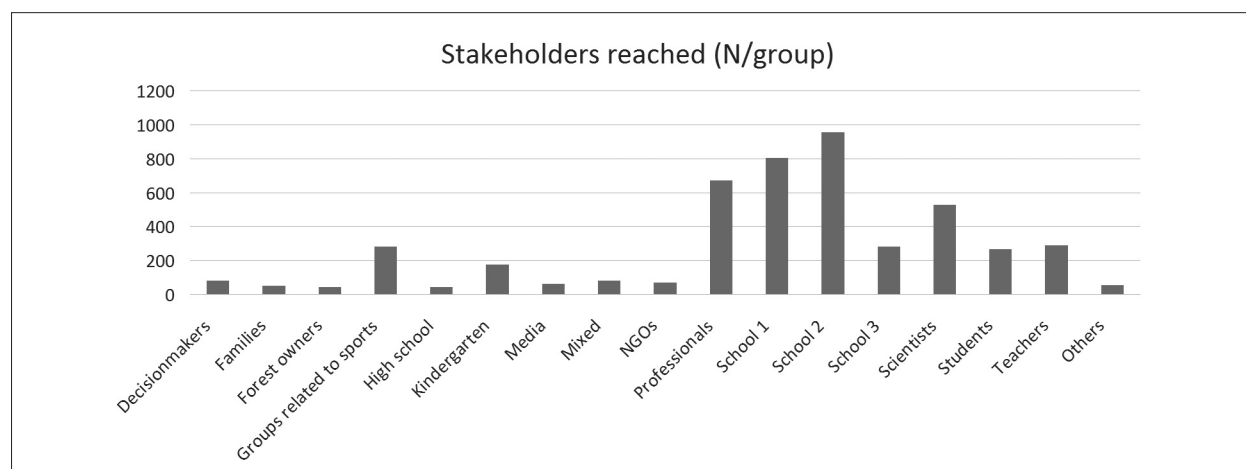


Figure 1: Participants at the LIFE GENMON events per target group from July 2014 to June 2016
Slika 1: Udeleženci na LIFE GENMON dogodkih za ciljne skupine, od julija 2014 do- junija 2016

POVZETEK

Za uspešno ozaveščanje javnosti o vrednosti storitev, ki jih zagotavlja naravno okolje, morajo te biti predstavljene na ustrezen način. Ljudje običajno niso pripravljeni plačati za storitve, ki so do zdaj veljale za proste (brezplačne). Spreminjanje načina razmišljanja s pomočjo ozaveščanja je torej velik izziv. Dobra komunikacijska strategija mora posredovati sporočilo o prispevku naravnega okolja h kakovosti človeškega življenja in bivanja. Ozaveščanje in širjenje znanja o storitvah, ki jih nudi naravno okolje, nam pomaga razumeti vrednost teh storitev in veselje ter zadovoljstvo, ki nam jih ponuja okolje danes in nam jih bo tudi v prihodnosti, če ga bomo obravnavali z znanjem, spoštovanjem in modrostjo.

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kov v soustvarjanje njegovih učinkov in večjega dosega njegovih sporočil in posledično komunikacijske strategije.

Komunikacijska strategija projekta LIFEGENMON se izvaja v dveh projektih aktivnostih preko različnih deležnikov in ciljnih skupin. Splošna diseminacija naslavlja otroke, študente, pedagoge, razne interesne skupine in medije. Usmerjena diseminacija pa aktivno vključuje v svoje aktivnosti odločevalce, regionalne oblasti, interesne skupine, akademsko sfero, gospodarske in obrtne zbornice, lastnike gozdov in njihova združenja, sindikate gozdnih delavcev, službe za civilno zaščito in prostovoljce ter turistične organizacije.

Povezovanje deležnikov in dialog s ključnimi organizacijami na področju (v sodelovanju z Zavodom za gozdove Slovenije, Univerzo v Ljubljani - Biotehniška fakulteta, Slovensko akademijo znanosti in umetnosti) preko (so-)organizacije konferenc, dogodkov in sejmov je ključno pri izvajanju projektih aktivnosti. Ozaveščanje in prisotnost na socialnih omrežjih pa dopolnjujeta in skrbita za širšo prepoznavnost ciljev projekta.

Učinek aktivnosti se spremlja preko seznamov udeležencev na dogodkih in preko monitoringa napredka projekta na podlagi vprašalnikov in intervjujev udeležencev.

LIFEGENMON ekipa je konec leta 2016 dosegla več kot 4000 ljudi, na več kot 90 različnih dogodkih, spletno stran LIFEGENMON je obiskalo več kot 8300 uporabnikov, ki so si ogledali več kot 30.000 strani.

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To the LIFEGENMON project coordinating beneficiary, Slovenian Forestry Institute.

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CONCENTRATION OF PROTEINS, BETA-GLUCANS, TOTAL PHENOLS AND ANTIOXIDANT CAPACITY OF SLOVENIAN SAMPLES OF BARLEY

VSEBNOST PROTEINOV, BETA-GLUKANOV, SKUPNIH FENOLOV IN ANTIOKSIDATIVNA VREDNOST SLOVENSКИH VZORCEV JEČMENA

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ABSTRACT

Concentration of proteins, beta-glucans, total phenols and antioxidant capacity of Slovenian samples of barley

Four Slovenian barley samples were analyzed for the content of the ash, proteins, β -glucans and total polyphenols; and for the antioxidant capacity by two methods. Antioxidant capacity was determined by the ferric reducing antioxidant power (FRAP) activity and by the oxygen radical absorbance capacity (ORAC) assay. For the comparison, four foreign barley commercial samples were analyzed, three of German origin and one from Italy. Slovenian samples had equal or lower content of ash, proteins and β -glucans in comparison to foreign samples. A Slovenian barley sample from Šalovci, Prekmurje had the highest content of β -glucans, namely 5.73% in dry weight (DW). The results obtained by both methods of the determination of antioxidant capacity showed significant correlation with the total polyphenols content.

Key words: *Hordeum*, barley, beta-glucans, phenols

IZVLEČEK

Vsebnost proteinov, beta-glukanov, skupnih fenolov in antioksidativna vrednost slovenskih vzorcev ječmena

Štiri vzorce ječmena iz Slovenije smo analizirali na vsebnost pepela, beljakovin, β -glukanov in skupnih fenolov. S pomočjo dveh metod smo tudi ugotavljali antioksidativno zmogljivost. Antioksidativna zmogljivost je bila določena z zmanjšanjem antioksidativne sposobnosti železa (metoda FRAP) in s sposobnostjo absorbance kisikovega radikala (metoda ORAC). Za primerjavo smo analizirali štiri komercialne vzorce tujega ječmena, tri nemške in enega iz Italije. Slovenski vzorci so imeli enako ali nižjo vsebnost pepela, beljakovin in β -glukanov v primerjavi s tujimi. Vendar je slovenski vzorec ječmena iz Šalovcev (Prekmurje) vseboval najvišjo vsebnost β -glukanov, in sicer 5,73% suhe teže (DW). Rezultati obeh metod določanja antioksidativne kapacitete so bili primerljivi in so pokazali pomembno povezavo z vsebnostjo celokupnih fenolov.

Ključne besede: *Hordeum*, ječmen, beta-glukani, fenoli

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1 INTRODUCTION

Interest in growing barley, a crop cultivated by human-kind for a long time, has become more intense in recent years in Slovenia, Europe and elsewhere, due to the nutritional value of this ancient crop. Barley was cultivated for centuries for human food, but the areas of growing of barley declined in the middle of the 20th century. However high fibre foods are recently preferred in human nutrition. It should be noted that the cultivation of barley was for many years particularly focused on achieving high dry matter content, high starch, and low content of dietary fibre. This is because barley has been used mostly for either animal feed or for production of barley malt. Cultivars with higher levels of dietary fibre, particularly of β -glucans, are therefore now gaining importance for use in human nutrition and some of these are being introduced to Southern and Central Europe, including Slovenia.

Identification of suitable cultivars high in β -glucan content, evolution of milling and dry separation techniques and using the β -glucan-enriched fractions as an ingredient in bakery products are subjects for further needed research. The methods developed will allow obtaining new, nutritionally valuable barley products with high potential for use in foods.

In recent months it was a quick development of research and published papers on barley β -glucans. SHAIK et al. (2016) reported that in amylose-only starch chemotypes it is showed significant reduction in

starch accumulation with the re-direction to protein and β -glucan accumulation. Ho et al. (2016) newly reported a systematic review of the effects of different high molecular weight β -glucans on cholesterol and cardiovascular disease risk reduction.

MOZA & GUJRAL (2016) reported that in India high altitude barley cultivars (between 1200 m and 3500 m above sea level) contain higher levels of total β -glucans in comparison to the barley cultivars grown in plains (97-126 m altitude). Cultivars grown in higher altitudes may find better utilization in nutraceutical foods. HAN et al. (2016), ZHU et al. (2016) and DIMITROFF et al. (2016) suggested how, understanding genetic background, barley β -glucans based functional food may be developed for the management of obesity. BELOBRAJDIC et al. (2016) suggest a wholegrain barley high molecular weight β -glucan to lower food intake.

After the separation in barley after milling to ten milling fractions, the highest β -glucan content was determined in the bran fraction (WIEGE et al. 2016). It was a strong positive correlation between β -glucan and protein content. Fractions containing high level of β -glucan have as well high concentration of protein and ash (WIEGE et al. 2016).

Croatian barley samples were screened for total high molecular weight β -glucans content (KRSTANOVIĆ et al. 2016), but such kind of research was not yet performed in Slovenia.

2 MATERIALS AND METHODS

2.1 Samples

Barley samples analysed are presented in Table 1.

Table 1: Description of studied barley samples
Preglednica 1: Opis preučevanih vzorcev ječmena

Sample	Description
A	Winter barley sample from Šentjernej, Dolenjska, Slovenia
B	Winter barley sample from Trbonje, Dravograd, Slovenia
C	Winter barley sample from Šalovci, Prekmurje, Slovenia
D	Winter barley sample from Murska Sobota, Prekmurje, Slovenia
E	Winter barley sample from Nienstad, Germany
F	Winter barley sample from Germany, cultivated in Jable, Slovenia
G	Naked barley sample from Asheberg, Germany
H	Winter barley sample from Lazio, Italy

Samples A-D are samples of barley grain, each of barley domestic cultivars was grown for years by farmers in different parts of Slovenia. The origin of barley

cultivars is not known. Samples E-F are commercial barley samples available in Slovenia, originating, as far as it is known, in Germany or Italy.

2.2 Methods

2.2.1 Determination of moisture, ash, proteins and β -glucans

Constituents of barley samples were determined using the standard methods.

For the determination of moisture: Method ICC No. 109/1, was used.

In short: This method can be taken as the standard for the development of methods which are specifically suited to the practical determination of the moisture content of wheat, rice (hulled paddy), barley, maize or whole maize meal, millet, rye and oats, as grains, ground grains, semolina and flour. It is not to be used for the settlement of commercial disputes. Measurement of moisture loss when the material, ground if necessary without change of moisture content, is equilibrated in an anhydrous atmosphere at a temperature between 45 and 50 °C and at a pressure of 1.3 ... 2.7 KPa (10 ... 20 mm Hg).

For the determination of ash: Method ICC No. 104, was used.

In short: Samples were ground and placed into crucibles, and crucibles placed into a muffle furnace. The ashing was carried out at 900 °C, and is completed when the cool residue was white or nearly white. As the ash quantity has to be related to dry matter, the moisture content of the test sample has to be determined separately.

For the determination of proteins: Method ICC 105/2. Kjeldhal Method was used.

In short: The organic matter of the sample is oxidized with concentrated sulfuric acid in the presence of a catalyst: the product of the reaction $(\text{NH}_4)_2\text{SO}_4$ is treated by alkali; free ammonia is distilled and titrated.

For the determination of β -glucans: Method ICC No.166 was used.

In short: β -D-glucan is determined using highly purified lichenase and β -D-glucosidase. β -D-glucan is specifically hydrolyzed by lichenase to oligosaccharides, which are quantitatively cleaved to glucose by β -glucosidase. Glucose is measured using glucose oxidase - peroxides - buffer mixture. Method is rapid procedure for direct, quantitative measurement of (1->3) (1->4)- β -D-glucan (β -D-using highly purified lichenase and β -glucosidase). β -D-glucan is specifically hydrolyzed by lichenase to oligosaccharides, which are then quantitatively cleaved to glucose by β -glucosidase. Glucose is measured using glucose oxidase-peroxidase-buffer mixture.

2.2.2 Extraction method

In this study, 80% methanol extracts from barley were used for the determination of total phenolic content and antioxidant property. Barley samples (1 g) were extracted with 6 ml acidified methanol (HCl/methanol/water, 1:80:10, v/v/v) at room temperature (25 °C) for 2 h using orbital shaker. The mixture was centrifuged at 3000g for 10 min. The supernatant was used for determination of total phenolic content and antioxidant capacity. Several studies have shown that 80% methanol is an effective solvent in extracting phenolic and other polar substances from cereals (LAHOUAR et al. 2014).

2.2.3 Total phenolic content determination

Total phenols (TP) were determined using a modification of the Folin-Ciocalteu method method described by SINGLETON & ROSSI (1965). Briefly, the assay was conducted by mixing 4 mL of deionized water, 0.25 mL of extracts (see 'Extract preparations' in this section), 0.25 mL of Folin-Ciocalteu reagent, and 0.5 mL of Na_2CO_3 . After 30 min at room temperature, the absorbance of the mixture was measured at 725 nm. A standard curve was prepared with gallic acid. The final results were expressed as mg of gallic acid equivalents (GAE) per g of dry weight (DW). All of the analyses were conducted in triplicate and the results report the sum of bound and soluble phenolic compounds.

2.2.4 Determination of ferric reducing antioxidant power (FRAP) activity

A ferric reducing antioxidant power (FRAP) assay was performed according to the method described by BENZIE & STRAIN (1999), which was adapted for 96-well plates and an automatic reader (Infinite 2000, Tecan, Salzburg, Austria). The method is based on the reduction of the Fe^{3+} -2,4,6-tripyridyl-s-triazine (TPTZ) complex to its ferrous form at a low pH. Briefly, 160 mL of FRAP assay solution (consisting of 20 mM ferric chloride solution, 10 mM TPTZ solution, and 0.3 M acetate buffer at pH 3.6) was prepared daily, mixed with 10 mL of the sample, standard, or blank, and dispensed into each well of a 96-well plate. The absorbance was measured at 595 nm at 37 °C after 30 min of incubation. All of the analyses were conducted in triplicate. The final results are expressed as $\mu\text{mol Fe}^{2+}$ equivalents per g of the DW of the samples, and the results were obtained using a standard curve with different concentrations of ferrous sulfate heptahydrate ($\text{FeSO}_4 \times 7\text{H}_2\text{O}$).

2.2.5 The oxygen radical absorbance capacity (ORAC) assay

The oxygen radical absorbance capacity (ORAC) was determined using the hydroxyl radical antioxidant capacity (HORAC) assay kit (Oxford Biomedical Re-

search, Oxford USA) according to the manufacturer's instructions. The final ORAC values were expressed as gallic acid equivalents and determined according to the standard curve. All of the analyses were conducted in triplicate.

3. RESULTS AND DISCUSSION

Slovenian barley samples were analysed and compared with the foreign ones. Results of the ash (i.e. total con-

tent of mineral elements), protein and β -glucans concentration are presented in Table 2.

Table 2: Concentration of ash, protein and β -glucans in barley samples, mean values and standard deviations (DW=dry weight).

Preglednica 2: Koncentracija pepela, beljakovin in β -glukanov v vzorcih ječmena, povprečne vrednosti in standardni odkloni (DW=sušina).

Sample	Moisture %	Ash % DW	Protein % DW	β -glucans % DW
A	10.9 \pm 0.3	1.83 \pm 0.08	8.7 \pm 0.1	4.56 \pm 0.09
B	11.0 \pm 0.5	1.67 \pm 0.10	8.8 \pm 0.2	4.06 \pm 0.07
C	11.0 \pm 0.3	1.30 \pm 0.11	8.2 \pm 0.6	5.73 \pm 0.11
D	11.3 \pm 0.2	1.66 \pm 0.04	8.2 \pm 0.5	4.19 \pm 0.06
E	10.9 \pm 0.4	1.72 \pm 0.07	9.7 \pm 0.5	5.36 \pm 0.08
F	10.7 \pm 0.3	1.95 \pm 0.09	9.8 \pm 0.2	4.76 \pm 0.04
G	10.3 \pm 0.5	2.25 \pm 0.06	11.1 \pm 0.7	5.19 \pm 0.06
H	11.4 \pm 0.3	2.42 \pm 0.03	9.6 \pm 0.4	3.73 \pm 0.10

All analyzed samples show a high protein content (average 9.3% DW), with values ranging from 8.2% DW to 11.1% DW. The sample G had a highest value. From the point of view of β -glucans the sample D (Šalovci, Slovenia) is the most interesting because it had highest β -glucans concentration.

Correlation coefficient between ash and proteins in grain among all samples analysed was 0.74; correlation coefficient between ash and β -glucans was -0.46; correlation coefficient between proteins and β -glucans was 0.15. We used Pearson correlation coefficient. Thus the correlation between ash and proteins is significant ($p < 0.5$) and positive, correlation between ash and β -glucans was significant ($p < 0.5$) and negative. There was no significant correlation between the amount of proteins and β -glucans in studied barley samples. Barley has a distinct advantage over some other grains in that β -glucan soluble fiber is found throughout the entire barley kernel. In some other grains, the fiber is only found in the outer bran layer. If these grains are processed, the fiber can be easily lost (WIEGE et al. 2016).

The all samples analysed showed average values of β -glucans 4.69% DW, with a maximum value of 5.73% DW and a minimum value of 3.73% DW.

The ash values were with a minimum of 1.3% DW, and a maximum of 2.42% DW.

To better understand the relationship between the antioxidant activity and total phenolic content, the TPC of the sample extracts were determined using the Folin-Ciocalteu phenol reagent. The results are expressed as mg of gallic acid equivalents per gram of dry mater, and are presented in Table 3. Barley samples with high amount of phenolics also showed high antioxidant activity (FRAP and ORAC assay).

The total antioxidant capacity was in linear regression with the total phenolic content in barley samples ($R^2 = 0.9948$ and 0.8192 for the results of FRAP and ORAC assays, respectively) (Fig. 1).

There were differences in the antioxidant activities by ORAC and FRAP, and in content of total polyphenols of investigated barley samples.

Table 3: The total phenolic contents (TP) and antioxidant capacity (FRAP and ORAC method) of barley samples. Data are reported to mean (n=3) ± standard error. Values in the same column not sharing a common letter differ significantly at P≤0.05. Test ANOVA. X mg GAE/g dried weight (DW). y μmol Fe²⁺ equivalent/g DW. z GAE mM/g DW. GAE: gallic acid equivalent; DW: dry weight

Preglednica 3: Vsebnost skupnih fenolov (TP) in antioksidativna zmogljivost (metodi FRAP in ORAC) pri različnih vzorcih ječmena. Podatki so povprečje (n = 3) ± standardna napaka. Vrednosti v istem stolpcu, ki nimajo iste črke se med seboj značilno razlikujejo P ≤ 0.05. Test ANOVA. X mg GAE/g suhe teže (DW). y μmol Fe²⁺ ekvivalent/g DW. z GAE mM/g DW. GAE: ekvivalent galne kisline; DW: suha teža

Sample	TPCx	FRAPy	ORACz
A	2.8 ± 0.2b	227.0 ± 1.5b	21.0 ± 0.1b
B	2.0 ± 0.3b	180.2 ± 2.9c	21.0 ± 0.3b
C	1.5 ± 0.2c	149.2 ± 1.6d	17.1 ± 0.2c
D	2.4 ± 0.3b	202.7 ± 0.2b	20.6 ± 0.4b
E	3.5 ± 0.5a	273.0 ± 0.9a	23.2 ± 0.9a
F	2.1 ± 0.2b	184.2 ± 1.3c	21.1 ± 0.2b
G	3.8 ± 0.4a	286.6 ± 2.1a	25.7 ± 0.2a
H	3.0 ± 0.1 a	231.0 ± 2.2 b	24.1 ± 0.1 a

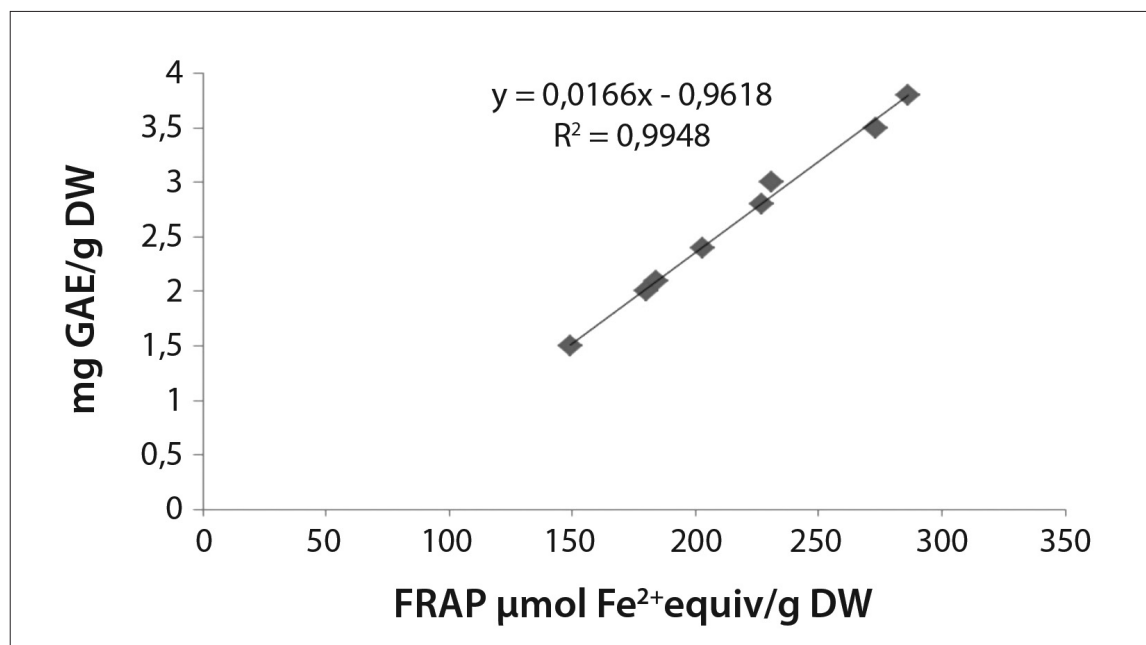


Figure 1: Linear regression between total phenolic content and the total antioxidant capacity (FRAP method - a ferric reducing antioxidant power) of barley samples. GAE: gallic acid equivalent; DW: dry weight

Slika 1: Linearna regresija med skupno vsebnostjo fenolov in skupno antioksidativno kapaciteto vzorcev ječmena (metoda FRAP - zmanjšanje antioksidativne moči železa). GAE: ekvivalent galne kisline; DW: suha teža

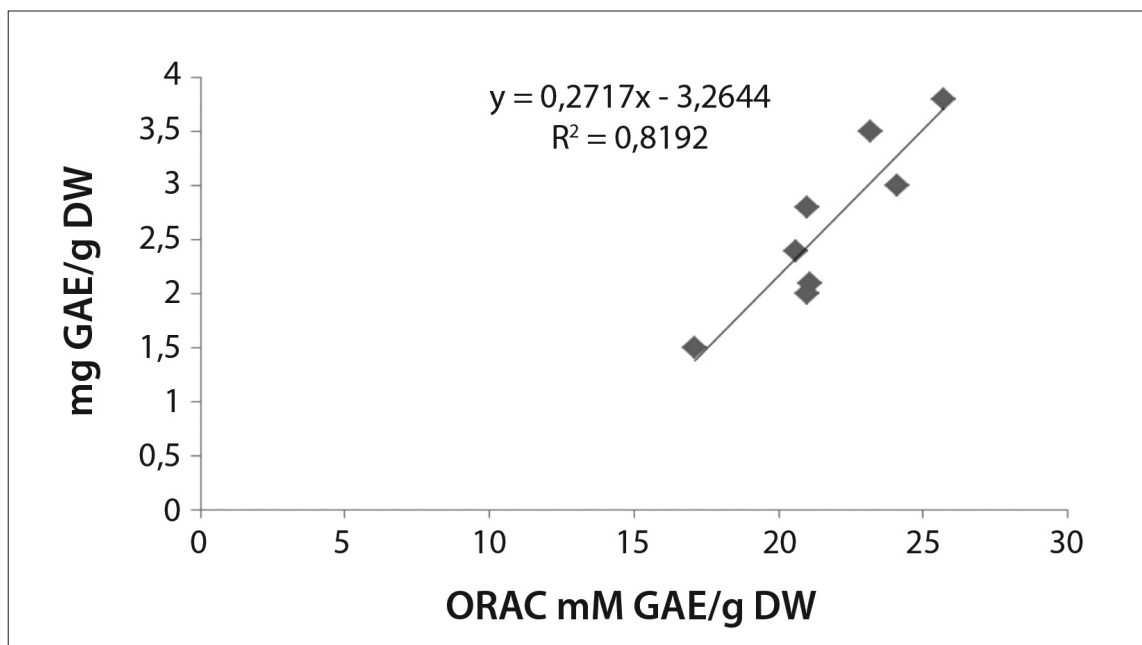


Figure 2: Linear regression between total phenolic content and the total antioxidant capacity (ORAC method - the oxygen radical absorbance capacity) of barley samples. GAE: gallic acid equivalent; DW: dry weight

Slika 2: Linearna regresija med skupno vsebnostjo fenolov in skupno antioksidativno kapaciteto vzorcev ječmena (ORAC metoda - absorbanca kisikovega radikala). GAE: ekvivalent galne kisline; DW: suha teža

4 CONCLUSIONS

The Slovenian barley sample from Šalovci, Prekmurje had the highest content of β -glucans among the studied Slovenian and foreign samples, namely 5.73%

in dry weight. The results obtained by both methods of the determination of antioxidant capacity were in linear relation with the total phenols content.

5 POVZETEK

Zanimanje za pridelovanje ječmena, stare poljščine, se je zadnje čase povečalo v Sloveniji, Evropi in drugod, predvsem zaradi njegove hranilne vrednosti. Ječmen gojijo za prehrano ljudi že stoletja, vendar so se območja pridelave zmanjšala zlasti v sredini 20. stoletja. V prehrani ljudi so zadnje čase zelo zaželena živila, kot je ječmen, z visoko vsebnostjo vlaknin. Žlahtnjenje in pridelovanje ječmena sta bili vrsto let osredotočeni predvsem na doseganje visokih pridelkov sušine, veliko škroba in nizko vsebnost prehranskih vlaknin, ker so ga večinoma uporabljali bodisi za živalsko krmo ali za proizvodnjo ječmenovega sladku. Sorte, ki vsebujejo več prehranskih vlaknin, še posebej β -glukanov, so v novejšem času vedno bolj pomembne za uporabo v prehrani ljudi.

V zadnjih mesecih je bil hiter razvoj raziskav in objavljanja člankov o ječmenovih β -glukanih. SHAIK

in sod. (2016) so poročali, da je genotip ječmena z amiloznim škrobom pokazal pomembno zmanjšanje kopičenja škroba s preusmerjanjem sinteze na beljakovine in akumulacijo β -glukanov.

Štiri vzorce ječmena iz Slovenije smo analizirali na vsebnost pepela, beljakovin, β -glukanov in skupnih fenolov ter za antioksidativno kapaciteto z dvema metodama. Antioksidativna zmogljivost je bila določena z zmanjšanjem antioksidativne moči železa (metoda FRAP) in s sposobnostjo absorbcije kisikovega radikala (metoda ORAC). Za primerjavo smo analizirali štiri tuje komercialne vzorce ječmena, tri nemškega izvora in enega iz Italije. Slovenski vzorci so imeli enako ali nižjo vsebnost pepela, beljakovin in β -glukanov v primerjavi s tujimi. Vendar je slovenski vzorec ječmena iz Šalovcev (Prekmurje) imel najvišjo vsebnost β -glukanov

in sicer 5,73% suhe teže (DW). Vzorci imajo povprečne vrednosti β -glukanov 4,69% suhe teže (DW), z najvišjo vrednostjo 5,73% suhe teže (DW) in minimalno vrednostjo 3,73% suhe teže (DW). Rezultati obeh metod določanja antioksidativne kapacitete so pokazali pomembno korelacijo s celotno vsebnostjo polifenolov.

Za boljše razumevanje razmerja med antioksidativnim delovanjem in skupno vsebnostjo fenolov so bili analizirani ekstrakti vzorcev za TPC z uporabo Folin-Ciocalteu-jevega fenolnega reagenta. Rezultati so bili izraženi kot mg ekvivalentov galne kisline na gram

suhe teže in so predstavljeni v razpredelnici 2. Raziskani vzorci ječmena E, G, H vsebujejo najvišjo vrednost TPC, medtem ko je bila najnižja vrednost v vzorcu C (razpredelnica 2, prvi stolpec). Na splošno je ječmen z veliko količino fenolov pokazal tudi visoko antioksidativno aktivnost (po metodah FRAP in ORAC). Skupna antioksidativna zmogljivost je bila tudi linearno povezana s skupno vsebnostjo fenolov v vzorcih ječmena ($R^2 = 0,9948$ in $0,8192$ za rezultate iz FRAP in ORAC metod) (sl. 1). Rezultati so pokazali, da so različni vzorci imeli različne antioksidativne sposobnosti.

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**Appendix 1 to paper in 57/1, pp. 5-61: Synoptic table of *Alnus glutinosa* dominating communities in Slovenia and N-Italy
 Priloga 1 k članku v 57/1, str. 5-61: Sintezna tabela združb črne jelše (*Alnetum glutinosae* s. lat.) v Sloveniji in severni Italiji**

Successive number (Zaporedna številka)		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	
Number of relevés (Število popisov)		18	6	4	10	5	4	10	9	7	4	8	11	7	52	32	11	6	
Sign for syntaxa (Oznaka sintaksona)		OrClo	LoAg-Vd	LoAgsb-R	LoAgcb-R	LoAgsbl-R	LoAg1-Vd	LoAg-ZP	PsCbla-Vd	Fra-Is	CacAg-R	CraA-Md	CelAaggr-A	SnAg-A	ChAg-It	CelAg-It	CelAgla-A	CelAgcr-A	
AG	<i>Alnetea glutinosae</i>																		
	<i>Alnus glutinosa</i>	E3b	50	100	100	100	100	100	89	14	100	100	100	100	100	100	100	100	
	<i>Alnus glutinosa</i>	E3a	11	17	.	30	40	25	70	11	.	13	
	<i>Alnus glutinosa</i>	E2b	.	.	.	10	0	25	40	22	.	75	.	27	29	.	64	33	
	<i>Alnus glutinosa</i>	E2a	.	.	.	20	20	25	10	11	.	38	
	<i>Salix cinerea</i>	E3a	20	25	
	<i>Salix cinerea</i>	E2b	20	11	14	25	38	.	.	42	34	27	.	
	<i>Salix aurita</i>	E2b	25	
	<i>Thelypteris palustris</i>	E1	20	21	47	.	.	
	<i>Ribes nigrum</i>	E2a	25	27	33	
	<i>Calamagrostis canescens</i>	E1	6	.	.	
AI	<i>Alnion incanae</i>																		
	<i>Rubus caesius</i>	E1	22	50	50	70	100	25	80	22	100	.	50	18	14	52	22	9	33
	<i>Carex pendula</i>	E1	22	50	.	.	.	25	10	78	29	.	.	.	6	.	.	.	
	<i>Hemerocallis fulva</i>	E1	11	.	.	.	20	
	<i>Cardamine impatiens</i>	E1	6	.	.	10	60	25	10	
	<i>Ribes rubrum</i>	E2a	6	33	30	3	.	.	
	<i>Alnus incana</i>	E3b	20	
	<i>Alnus incana</i>	E3a	6	10	14	
	<i>Alnus incana</i>	E2a	20	
	<i>Festuca gigantea</i>	E1	5	.	.	10	40	13	.	57	4	.	.	.	
	<i>Humulus lupulus</i>	E2b	.	33	.	10	60	10	.	.	.	25	.	0	
	<i>Humulus lupulus</i>	E2a	.	.	.	30	.	50	20	.	.	75	25	36	14	33	29	9	50
	<i>Humulus lupulus</i>	E1	.	17	.	.	.	10	
	<i>Populus alba</i>	E3b	.	33	14	2	.	.	.	
	<i>Aesculus hippocastanum</i>	E3a	.	17	25	
	<i>Aesculus hippocastanum</i>	E2b	.	0	25	10	20	
	<i>Aesculus hippocastanum</i>	E2a	.	17	25	10	20	
	<i>Equisetum arvense</i>	E1	.	17	75	70	20	50	80	11	29	50	50	36	100	10	9	.	
	<i>Equisetum telmateia</i>	E1	.	17	.	10	20	25	20	.	71	.	.	.	15	9	.	.	
	<i>Populus alba</i>	E3a	.	17	
	<i>Populus alba</i>	E2a	.	17	
	<i>Populus alba</i>	E1	.	17	
	<i>Agropyron caninum</i>	E1	25	
	<i>Carex remota</i>	E1	.	.	.	30	20	50	20	100	.	13	9	.	23	25	18	.	
	<i>Frangula alnus</i>	E3a	10	
	<i>Frangula alnus</i>	E2	.	.	.	10	.	25	20	14	50	13	18	.	36	53	74	.	
	<i>Frangula alnus</i>	E1	13	
	<i>Knautia drymeia</i> subsp. <i>intermedia</i>	E1	20	.	10	.	25	
	<i>Dryopteris carthusiana</i>	E1	50	30	11	.	13	18	.	40	16	81	50	
	<i>Chrysosplenium alternifolium</i>	E1	30	.	.	.	13	9	.	
	<i>Impatiens noli-tangere</i>	E1	30	2	.	9	67	
	<i>Arum italicum</i>	E1	29	
AqR	<i>Alno-Quercion roboris</i>																		
	<i>Quercus robur</i>	E3b	17	33	.	.	.	25	.	78	27	9	36	.	
	<i>Quercus robur</i>	E3a	6	17	.	.	.	10	
	<i>Quercus robur</i>	E2b	10	18	.	
	<i>Quercus robur</i>	E2a	.	17	.	.	.	10	
	<i>Quercus robur</i>	E1	11	17	25	.	.	25	.	22	.	25	13	.	.	.	9	.	
	<i>Fraxinus angustifolia</i>	E3b	.	17	78	100	.	13	.	.	.	18	.	
	<i>Fraxinus angustifolia</i>	E3a	.	17	11	86	.	13	.	.	4	.	50	
	<i>Fraxinus angustifolia</i>	E2b	.	17	13	
	<i>Fraxinus angustifolia</i>	E2a	.	33	56	43	25	13	
	<i>Fraxinus angustifolia</i>	E1	.	17	33	57	
	<i>Ulmus laevis</i>	E3b	.	.	.	10	
	<i>Ulmus laevis</i>	E3a	.	.	.	10	29	
	<i>Ulmus laevis</i>	E2b	14	
	<i>Ulmus laevis</i>	E2a	.	.	.	10	.	.	.	14	
	<i>Ulmus laevis</i>	E1	13	
	<i>Pseudostellaria europaea</i>	E1	25	.	.	44	
	<i>Leucopodium aestivum</i>	E1	100	.	100	13	18	.	2	91	.	
	<i>Clematis viticella</i>	E1	11	
	<i>Carex brizoides</i>	E1	13	27	71	27	3	18	17	
	<i>Prunus padus</i>	E2	82	29	.	.	55	33	
	<i>Prunus padus</i>	E1	18	14	
	<i>Rumex sanguineus</i>	E1	14	
	<i>Pulmonaria dacica</i>	E1	27	.	
	<i>Glechoma hirsuta</i>	E1	17	

Successive number (Zaporedna številka)		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	
SP	<i>Salicetea purpureae, Salicion albae</i>																		
	<i>Populus nigra</i>	E3b	28	67	.	.	25	.	.	43	8	3	.	.	
	<i>Populus nigra</i>	E3a	
	<i>Populus nigra</i>	E2b	14	
	<i>Populus nigra</i>	E2a	
	<i>Salix alba</i>	E3b	6	17	25	30	60	25	20	33	71	25	38	.	.	27	9	.	
	<i>Salix alba</i>	E3a	
	<i>Salix alba</i>	E2b	10	14	
	<i>Vitis sylvestris</i>	E3a	6	29	
	<i>Vitis sylvestris</i>	E2a	
	<i>Salix fragilis</i>	E3b	.	17	25	10	.	.	.	11	
	<i>Salix fragilis</i>	E3a	.	.	.	10	
	<i>Salix eleagnos</i>	E3b	.	17	2	.	.	
	<i>Salix eleagnos</i>	E3a	
	<i>Acer negundo</i>	E3b	
	<i>Acer negundo</i>	E3a	.	17	
	<i>Acer negundo</i>	E2b	
	<i>Acer negundo</i>	E2a	
	<i>Acer negundo</i>	E1	
	<i>Populus x canescens</i>	E3b	.	.	.	10	60	38	
	<i>Salix purpurea</i>	E2a	10	.	.	.	25	
	<i>Salix purpurea</i>	E3a	
	<i>Salix purpurea</i>	E2b	.	.	.	20	.	20	
	<i>Salix triandra</i>	E2b	13	.	29	.	.	.	
	TA	<i>Tilio-Acerion</i>																	
		<i>Acer platanoides</i>	E3b	.	.	.	20
		<i>Acer platanoides</i>	E2a	.	.	.	40
		<i>Acer platanoides</i>	E1	20
		<i>Acer pseudoplatanus</i>	E3b	33	33	25	20	40	.	30	.	.	13
		<i>Acer pseudoplatanus</i>	E3a	28	25	20
		<i>Acer pseudoplatanus</i>	E2b	28	17	25	50	40	25	50	.	.	50	.	9	29	2	.	.
		<i>Acer pseudoplatanus</i>	E2a	28	17	.	.	.	25	50
		<i>Acer pseudoplatanus</i>	E1	39	17	50	40	.	50	60	.	.	25	25
<i>Juglans regia</i>		E3b	28	
<i>Juglans regia</i>		E3a	25	
<i>Juglans regia</i>		E2b	.	.	.	20	
<i>Juglans regia</i>		E2a	.	.	10	20	14	
<i>Juglans regia</i>		E1	11	17	29	
<i>Lunaria rediviva</i>		E1	17	17	20	.	.	.	13	.	71	.	.	.	
<i>Tilia platyphyllos</i>		E3b	17	
<i>Tilia platyphyllos</i>		E3a	6	
<i>Tilia platyphyllos</i>		E2b	6	.	25	9	14	.	.	.	
<i>Tilia platyphyllos</i>		E2a	11	25	10	
<i>Tilia platyphyllos</i>		E1	
<i>Ulmus glabra</i>		E3b	11	17	
<i>Ulmus glabra</i>		E3a	17	
<i>Ulmus glabra</i>		E2b	22	10	
<i>Ulmus glabra</i>		E2a	11	.	.	20	.	20	6	.	
<i>Ulmus glabra</i>		E1	6	10	
<i>Thalictrum aquilegifolium</i>		E1	11	.	25	10	.	.	30	.	.	.	25	.	57	4	3	.	
<i>Arum maculatum</i>		E1	6	.	25	70	40	.	20	56	86	25	25	.	14	2	.	.	
<i>Aruncus dioicus</i>		E1	6	17	.	10	57	.	.	.	
<i>Cardamine flexuosa</i>		E1	10	
<i>Geranium robertianum</i>		E1	.	17	.	.	20	.	20	.	.	.	13	
<i>Hesperis candida</i>		E1	.	.	25	
<i>Adoxa moschatellina</i>		E1	.	.	.	20	.	10	14	.	.	.	
<i>Dryopteris affinis</i>		E1	10	14	.	.	.	
<i>Tephrosia pseudocrispa</i>		E1	10	
<i>Polystichum aculeatum</i>		E1	10	
<i>Staphylea pinnata</i>		E2b	11	
<i>Staphylea pinnata</i>		E2a	11	
EC		<i>Erythronio-Carpinion</i>																	
		<i>Primula vulgaris</i>	E1	83	33	.	50	40	50	60	22	43	.	.	.	29	2	.	.
		<i>Ornithogalum pyrenaicum</i>	E1	78	67	100	100	60	.	10	33	29
		<i>Galanthus nivalis</i>	E1	67	100	100	10	40	.	40	22
		<i>Helleborus odoratus</i>	E1	56	40	11
	<i>Lonicera caprifolium</i>	E2a	33	50	.	.	.	50	10	11	86	.	.	.	12	.	9	.	
	<i>Crocus vernus subsp. vernus</i>	E1	28	.	.	90	.	50	60	44	
	<i>Erythronium dens-canis</i>	E1	28	25	.	56	
	<i>Ranunculus aesculinus</i>	E1	6	50	30	100	
	AF	<i>Aremonio-Fagion</i>																	
		<i>Lamium orvala</i>	E1	100	100	100	100	80	50	90	78	14	.	.	.	14	2	.	.
		<i>Hacquetia epipactis</i>	E1	78	25	40	14
<i>Isopyrum thalictroides</i>		E1	50	17	10	
<i>Cardamine enneaphyllos</i>		E1	39	20	
<i>Cyclamen purpurascens</i>		E1	39	17	25	
<i>Geranium nodosum</i>		E1	11	.	50	60	.	.	10	
<i>Knautia drymeia subsp. drymeia</i>		E1	.	.	.	40	20	50	10	.	14	.	25	.	100	.	.	.	
<i>Euphorbia carniolica</i>		E1	.	.	.	20	.	25	25	.	14	.	.	.	
<i>Anemone trifolia</i>		E1	60	

Successive number (Zaporedna številka)	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
<i>Omphalodes verna</i>	E1	20
<i>Cardamine trifolia</i>	E1	10	11
<i>Helleborus niger</i>	E1	10
<i>Potentilla carniolica</i>	E1	10
<i>Scopolia carniolica</i>	E1	29
FS <i>Fagetalia sylvaticae</i>																	
<i>Symphytum tuberosum</i>	E1	94	100	75	100	.	100	30	67	14	.	.	71	4	.	.	.
<i>Lilium martagon</i>	E1	83	17	33
<i>Polygonatum multiflorum</i>	E1	83	17	.	.	.	75	40	56	.	25	.	14	12	.	.	.
<i>Pulmonaria officinalis</i>	E1	83	.	100	100	40	50	40	33	.	13	.	14
<i>Cardamine bulbifera</i>	E1	78	33	.	80	20	.	10	11
<i>Carpinus betulus</i>	E3b	78	17	50	30	.	.	.	56
<i>Carpinus betulus</i>	E3a	67	33	100	70	40	50	20	100
<i>Carpinus betulus</i>	E2b	56	.	.	20	.	25	10	22	.	25	.	.	2	.	.	.
<i>Carpinus betulus</i>	E2a	28	.	25	40	20	.	.	22	14
<i>Carpinus betulus</i>	E1	25	10	.	33	.	13
<i>Corydalis cava</i>	E1	67	17	.	.	40	.	10
<i>Asarum europaeum subsp. caucasicum</i>	E1	50	17	100	30	.	25	60
<i>Brachypodium sylvaticum</i>	E1	50	50	75	100	100	75	70	.	86	25	13	.	100	10	.	.
<i>Viola reichenbachiana</i>	E1	50	50	.	.	.	75	30	67	57	.	.	14	4	3	.	.
<i>Sambucus nigra</i>	E3a	0	33	.	.	40
<i>Sambucus nigra</i>	E2b	28	100	50	40	80	50	70	.	.	25	50	45	57	48	9	33
<i>Sambucus nigra</i>	E2a	44	33	50	60	60	25	40	11	.	.	75
<i>Sambucus nigra</i>	E1	6	17	10
<i>Heracleum sphondylium</i>	E1	39	33	25	40	20	25	30	14
<i>Allium ursinum</i>	E1	33	.	75	10	60	.	.	100	.	.	13	.	.	2	.	.
<i>Galeobdolon flavidum</i>	E1	28	17	80	2	.	.
<i>Galeobdolon montanum</i>	E1	28	.	25	90	60	25	.	67	.	25	.	18	57	2	.	17
<i>Mercurialis perennis</i>	E1	28	.	.	20	.	.	10	33	14	.	.	.
<i>Euphorbia dulcis</i>	E1	22	33	100	30	.	50	20
<i>Fagus sylvatica</i>	E3b	11
<i>Fagus sylvatica</i>	E3a	6	.	.	.	20
<i>Fagus sylvatica</i>	E2b	17
<i>Fagus sylvatica</i>	E2a	11	25	10
<i>Fagus sylvatica</i>	E1	.	.	25	0	20	0
<i>Fraxinus excelsior</i>	E3b	17	.	25	70	100	50	50	71	.	.	.
<i>Fraxinus excelsior</i>	E3a	.	17	25	40	.	50	80	.	.	.	13	27	.	12	31	.
<i>Fraxinus excelsior</i>	E2b	6	17	.	70	.	50	90	.	.	50	50	9	71	.	.	.
<i>Fraxinus excelsior</i>	E2a	.	.	.	90	40	50	90	.	.	75	75
<i>Fraxinus excelsior</i>	E1	6	.	25	60	40	25	60	.	.	75	13
<i>Tilia cordata</i>	E3b	11	10
<i>Tilia cordata</i>	E3a	6	.	.	.	20
<i>Tilia cordata</i>	E2b	6	.	25	.	20	.	30
<i>Tilia cordata</i>	E2a	6	.	25	.	20	.	20	.	.	25	13
<i>Tilia cordata</i>	E1	10
<i>Carex sylvatica</i>	E1	11	.	.	60	.	75	20	33	86	.	13	.	.	8	.	.
<i>Lathyrus vernus</i>	E1	11
<i>Prunus avium</i>	E3b	11	17	25	10	14
<i>Prunus avium</i>	E3a	.	33	.	.	20	25	.	.	43	.	13	.	.	8	6	.
<i>Prunus avium</i>	E2b	.	17	10	.	14	.	25
<i>Prunus avium</i>	E2a	.	33	.	20	.	25
<i>Prunus avium</i>	E1	6	17	.	10	20	50	10	.	57	50	13
<i>Salvia glutinosa</i>	E1	11	17	25	100	40	50	40	.	71	3	.
<i>Circaea lutetiana</i>	E1	6	17	.	30	.	50	30	.	.	.	13	18	14	17	.	27
<i>Daphne mezereum</i>	E2a	6	30	.	.	.	13	18
<i>Dryopteris filix-mas</i>	E1	6	20	.	.	.	13	.	.	33	3	.
<i>Euphorbia amygdaloides</i>	E1	6	10	.	14	.	.	.	43	.	.	.
<i>Galium laevigatum</i>	E1	6	.	.	.	20	25
<i>Melica nutans</i>	E1	6	10	.	.	25
<i>Neottia nidus-avis</i>	E1	6	.	.	10
<i>Ranunculus lanuginosus</i>	E1	6	50	100	100	60	.	30	.	29	.	13	.	86	.	.	.
<i>Actaea spicata</i>	E1	.	.	.	10	.	25
<i>Campanula trachelium</i>	E1	.	.	.	10	.	25	30
<i>Scrophularia nodosa</i>	E1	.	.	.	10	60	13	.	14	2	.	18
<i>Phyteuma spicatum subsp. coeruleum</i>	E1	.	.	.	10	.	.	10	33
<i>Sanicula europaea</i>	E1	50	.	.	29
<i>Mycelis muralis</i>	E1	25
<i>Paris quadrifolia</i>	E1	25	80	67	3	9
<i>Myosotis sylvatica</i>	E1	30	.	.	.	9	14
<i>Leucjum vernum</i>	E1	10	.	.	.	13	9	29	2	.	.
<i>Asarum europaeum subsp. europaeum</i>	E1	25	.	14	.	.	.
<i>Petasites albus</i>	E1	9	14
QP <i>Quercetalia pubescenti-petraeae</i>																	
<i>Helleborus odoratus subsp. istriacus</i>	E1	61	.	25	.	20	25
<i>Ruscus aculeatus</i>	E1	44	33	.	.	.	50	.	56	86
<i>Lathyrus venetus</i>	E1	28	17
<i>Cornus mas</i>	E2b	22
<i>Ostrya carpinifolia</i>	E3b	17	25
<i>Ostrya carpinifolia</i>	E3a	11	10
<i>Acer obtusatum</i>	E2a	6

Successive number (Zaporedna številka)	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
<i>Acer obtusatum</i>	E1	11
<i>Fraxinus ornus</i>	E3a	11	.	.	.	25
<i>Fraxinus ornus</i>	E2b	25
<i>Fraxinus ornus</i>	E2a	.	17	.	.	25	10	.	.	.	13	.	.	2	.	.	.
<i>Fraxinus ornus</i>	E1	50
<i>Cornus mas</i>	E2a	11	10
<i>Sesleria autumnalis</i>	E1	11	.	.	.	25
<i>Aristolochia lutea</i>	E1	6	.	.	.	25
<i>Asparagus tenuifolius</i>	E1	6	10
<i>Convallaria majalis</i>	E1	6	.	.	20	25
<i>Mercurialis ovata</i>	E1	6
<i>Tamus communis</i>	E1	6	71	.	.	14	17
<i>Carex flacca</i>	E1	57	25	38
<i>Quercus cerris</i>	E2a	.	.	10
<i>Quercus cerris</i>	E3b	.	.	75
<i>Quercus cerris</i>	E1	.	.	10	25
<i>Aristolochia pallida</i>	E1	43
<i>Buglossoides purpureoacerulea</i>	E1	29
<i>Quercus pubescens</i>	E3a	14
<i>Quercus pubescens</i>	E1	29
<i>Carpinus orientalis</i>	E21	14
<i>Orchis purpurea</i>	E1	14
<i>Sorbus aria</i>	E2b	13
QR <i>Quercetalia roboris</i>																	
<i>Betonica officinalis</i>	E1	.	.	10	.	75	10	.	.	50	38
<i>Rubus hirtus</i>	E2a	25	13
<i>Castanea sativa</i>	E1	14
<i>Serratula tinctoria</i>	E1	14	50	.	29
<i>Hieracium racemosum</i>	E1	14
<i>Potentilla erecta</i>	E1	13	.	.	4	.	.	.
<i>Populus tremula</i>	E3	9	.	.
<i>Spiraea salicifolia</i>	E1	36	.
QF <i>Quercus-Fagetea</i>																	
<i>Anemone nemorosa</i>	E1	100	83	100	90	60	75	50	89	.	25	13	.	57	2	.	9
<i>Hedera helix</i>	E3a	61	100	75	10	60	50	30	44	.	25
<i>Hedera helix</i>	E1	94	100	75	20	80	100	100	67	100	25	88	9	43	17	3	.
<i>Ranunculus ficaria</i>	E1	83	100	100	100	60	25	60	89	71	50	63	9	86	4	.	18
<i>Corylus avellana</i>	E3a	78	50	75	60	60	.	.	11
<i>Corylus avellana</i>	E2b	28	83	75	70	100	50	30	56	14	25	25	.	.	6	9	18
<i>Corylus avellana</i>	E2a	6	17	25	40	.	40	22	.	25
<i>Corylus avellana</i>	E1	6
<i>Gagea lutea</i>	E1	78	17	100	80	40	.	10
<i>Hepatica nobilis</i>	E1	78	.	25	.	.	.	10
<i>Anemone ranunculoides</i>	E1	72	33	.	60	20	.	20	33	.	13
<i>Lathraea squamaria</i>	E1	67	17	50	20
<i>Acer campestre</i>	E3b	61	50	50	50	.	.	.	11	14
<i>Acer campestre</i>	E3a	61	100	75	40	60	25	.	22	29
<i>Acer campestre</i>	E2b	61	50	100	50	60	25	10	44	86	25	13	18	29	4	6	.
<i>Acer campestre</i>	E2a	50	67	75	80	80	50	70	33	71	100	13
<i>Acer campestre</i>	E1	33	50	75	30	40	.	30	56	71	25	13
<i>Veratrum nigrum</i>	E1	39	10
<i>Orobancha hederace</i>	E1	33
<i>Carex digitata</i>	E1	28	17	25	.	.	25
<i>Scilla bifolia</i>	E1	22	17	100	.	60	25	.	11
<i>Ulmus minor</i>	E3b	11	33	.	10	.	.	.	44
<i>Ulmus minor</i>	E3a	17	33	.	10	.	.	.	67	29	.	.	.	10	6	.	.
<i>Ulmus minor</i>	E2b	.	.	.	20	.	.	.	67	29	17
<i>Ulmus minor</i>	E2a	6	50	.	10	.	.	.	100	43
<i>Ulmus minor</i>	E1	6
<i>Clematis vitalba</i>	E3a	11	.	.	40
<i>Clematis vitalba</i>	E2	6	.	.	10	.	25	10	.	14	.	13	.	2	.	.	.
<i>Clematis vitalba</i>	E1	25	20
<i>Malus sylvestris</i>	E3b	11	.	.	10
<i>Malus sylvestris</i>	E3a	6	.	.	40	.	.	10	11	14
<i>Malus sylvestris</i>	E2b	6	.	.	.	50	25
<i>Malus sylvestris</i>	E2a	6	.	.	10	.	0	.	14	25
<i>Pyrus pyraster</i>	E3b	11	.	.	20	.	25	.	14
<i>Pyrus pyraster</i>	E3a	.	.	.	10	25
<i>Pyrus pyraster</i>	E2b	.	.	.	20	9	.
<i>Pyrus pyraster</i>	E2a	.	.	25	10	.	25
<i>Quercus petraea</i>	E3b	11
<i>Quercus petraea</i>	E3a	6
<i>Quercus petraea</i>	E1	6	17	.	.	25	.	.	14	2	.	.	.
<i>Vinca minor</i>	E1	11	40	44
<i>Rosa arvensis</i>	E2a	6	17	50	30	.	25	2	.	.	.
<i>Listera ovata</i>	E1	6	.	75	10	.	25	70	22	57	.	38	9	43	.	.	.
<i>Cerastium sylvaticum</i>	E1	6	.	50	50	60	50	30	.	.	.	13	.	.	2	3	17
<i>Carex pilosa</i>	E1	6	22
<i>Crataegus curvisepala</i>	E2b	6
<i>Moehringia trinervia</i>	E1	6

Successive number (Zaporedna številka)	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
<i>Viola alba subsp. alba</i>	E1
<i>Stellaria holostea</i>	E1	.	.	.	10
<i>Carex montana</i>	E1	25	10
<i>Dactylorhiza fuchsii</i>	E1	25	20	9	.
<i>Ilex aquifolium</i>	E2a	25
<i>Lonicera xylosteum</i>	E2a	25	10	9	14	2	.	.	.
<i>Platanthera chlorantha</i>	E1	25
<i>Carex umbrosa</i>	E1	20	4	.	.	.
<i>Ranunculus casubicus</i>	E1	10
<i>Viola riviniana</i>	E1	10
<i>Cruciata glabra</i>	E1	13	9
<i>Ranunculus auricomus</i>	E1	18	.	.	.	9	.
QI Quercetea ilicis																	
<i>Rosa sempervirens</i>	E2	29
<i>Asparagus acutifolius</i>	E2a	29
<i>Laurus nobilis</i>	E2b	14
<i>Rubia peregrina</i>	E1	14
EP Erico-Pinetea																	
<i>Carex ornithopoda</i>	E1	.	.	25
<i>Molinia caerulea subsp. arundinacea</i>	E1	25	.	.	14	.	13	.	.	6	9	.	.
<i>Pinus sylvestris</i>	E3b	10	.	.	.	13	.	.	2	.	.	.
<i>Pinus sylvestris</i>	E2b	13
<i>Carex alba</i>	E1	10
VP Vaccinio-Piceetea																	
<i>Aposeris foetida</i>	E1	56	.	75	60	.	25	20	14
<i>Picea abies</i>	E3a	6
<i>Picea abies</i>	E2a	38	.	29	2	.	.	.
<i>Picea abies</i>	E1	.	.	.	10	13
<i>Oxalis acetosella</i>	E1	.	.	50	.	.	25	10	6	.	.
<i>Abies alba</i>	E2a	.	.	25	14
<i>Abies alba</i>	E3b	.	.	25
<i>Abies alba</i>	E1	.	.	25
<i>Gentiana asclepiadea</i>	E1	.	.	.	30	.	25
<i>Calamagrostis arundinacea</i>	E1	25	20
<i>Dryopteris dilatata</i>	E1	25	10	2	.	.	.
<i>Dryopteris assimilis</i>	E1	10	18	.
<i>Valeriana tripteris</i>	E1	10
<i>Veronica urticifolia</i>	E1	10
RP Rhamno-Prunetea																	
<i>Ligustrum vulgare</i>	E2b	11	.	25	20	20	.	10	11	14	25	.	.	14	.	.	.
<i>Ligustrum vulgare</i>	E2a	78	50	50	60	40	100	60	22	71	50	13	9	.	21	.	.
<i>Crataegus laevigata</i>	E3a	61
<i>Crataegus laevigata</i>	E2b	33	50	.	89	29	55	.
<i>Crataegus laevigata</i>	E2a	6	17	.	.	.	50	.	33
<i>Euonymus europaea</i>	E3a	10
<i>Euonymus europaea</i>	E2b	33	.	25	20	80	25	10	.	29	.	38	.	57	.	.	.
<i>Euonymus europaea</i>	E2a	44	67	100	60	80	50	60	56	43	25	75	36	.	17	9	18
<i>Euonymus europaea</i>	E1	22	17	25	10	.	25	10	.	.	25
<i>Cornus sanguinea</i>	E3a	6	17	.	.	20	.	10	.	57
<i>Cornus sanguinea</i>	E2b	17	17	50	60	80	.	60	.	100	.	25	18	57	60	22	17
<i>Cornus sanguinea</i>	E2a	28	50	75	90	80	50	90	11	71	75	75
<i>Cornus sanguinea</i>	E1	.	17
<i>Crataegus monogyna</i>	E3a	6	17	25	30	.	75	.	.	57
<i>Crataegus monogyna</i>	E2b	22	50	75	70	40	50	.	11	86	75	25	9	.	10	3	.
<i>Crataegus monogyna</i>	E2a	17	17	50	90	40	50	10	22	43	50	13
<i>Rhamnus catharticus</i>	E2b	6	13
<i>Rhamnus catharticus</i>	E2a	.	17	10	.	29	.	38	64	.	12	3	9
<i>Rhamnus catharticus</i>	E1	25
<i>Berberis vulgaris</i>	E2a	6
<i>Viburnum opulus</i>	E2b	60	25	20	11	.	25	88
<i>Viburnum opulus</i>	E2a	.	50	75	80	40	100	60	56	.	75	100	82	86	38	16	82
<i>Viburnum opulus</i>	E1	30	14
<i>Prunus spinosa</i>	E2b	.	17	.	30	20	50	.	.	100	25	13
<i>Prunus spinosa</i>	E2a	.	17	50	80	20	25	10	.	86	75	9	.
<i>Rubus fruticosus agg.</i>	E2b	.	.	.	10	.	.	10	.	29	3	.	.
<i>Rubus ulmifolius</i>	E2b	43	.	.	.	35	.	.	.
<i>Sorbus aucuparia</i>	E2	14
<i>Viburnum lantana</i>	E2b	25	10	4	.	.	.
<i>Rosa canina agg.</i>	E2b	14
MuA Mulgedio-Aconitetea																	
<i>Aconitum lycoctonum</i>	E1	17	17	.	80	20	0	10	2	.	.	.
<i>Athyrium filix-femina</i>	E1	11	.	.	10	20	50	50	11	.	.	25	18	14	17	6	55
<i>Senecio nemorensis</i>	E1	11	.	.	50	.	50	10
<i>Senecio ovatus</i>	E1	6	.	.	30	.	.	10	.	.	25
<i>Veratrum album s. lat.</i>	E1	.	.	75	90	20	.	10	.	.	.	38	18	43	.	18	33
<i>Aconitum variegatum</i>	E1	.	.	25
<i>Doronicum austriacum</i>	E1	.	.	.	10	9	.
<i>Silene dioica</i>	E1	20	.	10
<i>Chaerophyllum hirsutum</i>	E1	50	.	.	.	50	27	100	.	.	.
<i>Stellaria nemorum s.str.</i>	E1	10	.	.	.	13	.	71	.	.	.

Successive number (Zaporedna številka)	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
<i>Aconitum degenii</i> subsp. <i>paniculatum</i>	E1	10
<i>Geum rivale</i>	E1	38	36	29
<i>Carduus personata</i>	E1	13
<i>Ribes uva-crispa</i>	E1	14	.	.	.	17
EA <i>Epilobietea angustifolii</i>																	
<i>Stachys sylvatica</i>	E1	6	17	.	60	60	25	30	.	.	25	13	.	86	.	.	.
<i>Arctium nemorosum</i>	E1	.	.	25	70
<i>Fragaria vesca</i>	E1	.	.	25
<i>Arctium minus</i>	E1	.	.	.	30	20	50	10	.	.	.	13
<i>Eupatorium cannabinum</i>	E1	.	.	.	10	20	.	30	.	14	25	63	45	43	10	3	.
<i>Solanum dulcamara</i>	E1	20	25	30	.	.	25	50	9	.	56	25	55
<i>Galeopsis speciosa</i>	E1	20	25	30	.	.	.	38
<i>Bromus ramosus</i> subsp. <i>benekenii</i>	E1	10
<i>Galeopsis pubescens</i>	E1	10
<i>Rubus idaeus</i>	E2a	25	3	.
<i>Tussilago farfara</i>	E1	13
TG <i>Trifolio-Geranietea</i>																	
<i>Campanula rapunculoides</i>	E1	17	.	25
<i>Lilium bulbiferum</i>	E1	.	.	25
<i>Vincetoxicum hirundinaria</i>	E1	25	2	.	.
<i>Viola hirta</i>	E1	10	.	14
FB <i>Festuco-Brometea</i>																	
<i>Carlina acaulis</i>	E1	10
<i>Muscari neglectum</i>	E1	29
<i>Filipendula vulgaris</i>	E1	14
CD <i>Caricetalia davallianae</i>																	
<i>Carex lepidocarpa</i>	E1	10
<i>Carex davalliana</i>	E1	13
<i>Menyanthes trifoliata</i>	E1	13
<i>Carex stellulata</i>	E1	2	.	.
<i>Cladium mariscus</i>	E1	2	3	.
<i>Ranunculus flammula</i>	E1	18
MC <i>Montio-Cardaminetea</i>																	
<i>Carex flava</i> s. str.	E1	10
<i>Cardamine amara</i>	E1	22	.	.	38	27	71	13	.	9
CU <i>Calluno-Ulicetea</i>																	
<i>Carex pallescens</i>	E1	25	.	.	.	25	.	.	.	2	.	.	.
Ca <i>Calthion</i>																	
<i>Caltha palustris</i>	E1	.	.	25	.	25	60	56	.	.	100	82	86	19	6	64	83
<i>Angelica sylvestris</i>	E1	.	.	.	30	.	50	60	11	14	75	63	73	29	10	13	9
<i>Scirpus sylvaticus</i>	E1	.	.	.	10	.	60	.	.	.	0	50	27	.	10	9	9
<i>Myosotis scorpioides</i>	E1	10	11	.	.	25	25	.	.	27	3	82
Mo <i>Molinietalia caeruleae</i>																	
<i>Colchicum autumnale</i>	E1	44	.	75	50	.	75	60	56	14	25	13	.	86	.	.	.
<i>Cirsium oleraceum</i>	E1	.	.	.	70	40	.	80	.	.	75	63	55	43	6	6	9
<i>Equisetum palustre</i>	E1	.	.	.	20	.	.	20	.	.	75	63	.	.	15	9	.
<i>Valeriana dioica</i>	E1	.	.	.	10	.	25	60	33	.	75	88	73	71	33	22	91
<i>Cirsium palustre</i>	E1	.	.	.	10	25	.	.	.	6	3	.
<i>Cardamine pratensis</i>	E1	20	.	.	11	.	.	88	9	43	.	.	55
<i>Ophioglossum vulgatum</i>	E1	25	20	.	14	25	.	9
<i>Selinum carvifolia</i>	E1	30
<i>Juncus effusus</i>	E1	20	.	.	.	50	27	.	.	.	45
<i>Juncus inflexus</i>	E1	20	.	14	25
<i>Oenanthe lachenalii</i>	E1	29
<i>Carex tomentosa</i>	E1	14	25	25
<i>Crepis paludosa</i>	E1	75	64	57	6	.	.	.
<i>Dactylorhiza majalis</i>	E1	13
<i>Cirsium canum</i>	E1	13
<i>Molinia caerulea</i> subsp. <i>caerulea</i>	E1	4	13	.
<i>Sanguisorba officinalis</i>	E1	4	.	.
<i>Thalictrum lucidum</i>	E1	2	6	.
<i>Gratiola officinalis</i>	E1	3	.
<i>Juncus conglomeratus</i>	E1	18
<i>Thalictrum flavum</i>	E1	17
FP <i>Filipendulo-Petasion</i>																	
<i>Filipendula ulmaria</i>	E1	.	.	.	30	20	.	30	22	.	75	75	82	100	33	25	64
<i>Valeriana officinalis</i>	E1	.	.	.	20	20	25	.	.	.	50	.	43	10	9	.	.
<i>Lysimachia vulgaris</i>	E1	25	30	.	14	25	75	9	14	25	66	81
<i>Lythrum salicaria</i>	E1	25	20	45	.	31	44	18	67
<i>Mentha aquatica</i>	E1	30	11	.	.	50	63	64	71	8	9	9
<i>Myosoton aquaticum</i>	E1	10	.	.	.	13	.	.	.	8	.	.
<i>Symphytum officinale</i>	E1	40	9	.
<i>Stachys palustris</i>	E1	4	6	33
PP <i>Potentillo-Polygonetalia</i>																	
<i>Ranunculus repens</i>	E1	.	.	.	40	40	25	50	11	.	75	75	73	57	10	9	45
<i>Barbarea vulgaris</i>	E1	.	.	.	20	.	.	10	.	.	.	13	.	14	.	.	.
<i>Plantago major</i> subsp. <i>intermedia</i>	E1	25	.	.	14
<i>Rumex conglomeratus</i>	E1	11	18
<i>Rumex crispus</i>	E1	43	.	13	.	.	.	3	.
<i>Duchesnea indica</i>	E1	4	3	.

Successive number (Zaporedna številka)		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
<i>Carex hirta</i>	E1	4	.	.	.
<i>Agrostis stolonifera</i>	E1	2	.	.	.
MA Molinio-Arrhenatheretea																		
<i>Allium scorodoprasum</i>	E1	6	.	25	.	20	.	30
<i>Deschampsia cespitosa</i>	E1	.	.	50	70	80	50	70	22	.	100	50	18	86	8	3	.	17
<i>Crocus albiflorus</i>	E1	.	.	25	20	.	25	20
<i>Anthriscus sylvestris</i>	E1	.	.	25	30	40	.	60	.	.	.	13
<i>Astrantia major</i>	E1	.	.	25
<i>Muscari botryoides</i>	E1	.	.	25	40	25
<i>Veronica chamaedrys</i>	E1	.	.	.	50	20	.	20	.	.	.	13
<i>Ajuga reptans</i>	E1	.	.	.	40	20	50	20	56	86	50	38	64	43
<i>Dactylis glomerata s.str.</i>	E1	.	.	.	10	20	25	10	.	14	25	13
<i>Galium mollugo agg.</i>	E1	.	.	.	10	50	.	.	4	.	.	.
<i>Poa trivialis</i>	E1	.	.	.	10	60	.	30	.	43	.	38	.	.	4	3	.	.
<i>Prunella vulgaris</i>	E1	.	.	.	10	.	25	.	.	.	25
<i>Taraxacum officinale</i>	E1	.	.	.	10	40	.	20	.	29	25
<i>Vicia sepium</i>	E1	40	.	10
<i>Lysimachia nummularia</i>	E1	20	25	10	11	71	.	.	18	29	8	.	.	.
<i>Veronica serpyllifolia</i>	E1	20
<i>Ranunculus acris</i>	E1	10	.	29	50	13
<i>Lychnis flos-cuculi</i>	E1	13	.	.	2	.	9	17
PM Phragmiti-Magnocaricetea																		
<i>Galium palustre</i>	E1	.	.	50	10	.	.	10	11	.	75	25	.	.	21	41	82	33
<i>Lycopus europaeus</i>	E1	.	.	.	50	20	50	20	11	29	100	25	82	29	29	59	82	33
<i>Glyceria notata (maxima agg.)</i>	E1	.	.	.	10	4	.	9	50
<i>Phalaris arundinacea</i>	E1	20	.	10	9	.	25	.	18	50
<i>Iris pseudacorus</i>	E1	25	.	11	29	0	38	.	.	50	47	.	50
<i>Carex elata</i>	E1	25	20	22	27	100	64	50
<i>Carex randalpina</i>	E1	10	.	.	25	50
<i>Allisma plantago-aquatica</i>	E1	10	2	.	.	17
<i>Carex vesicaria</i>	E1	10	2	.	73	33
<i>Equisetum fluviatile</i>	E1	10	9
<i>Carex vulpina</i>	E1	11
<i>Carex elongata</i>	E1	14	.	.	55	.	13	.	100	50
<i>Carex acuta</i>	E1	75	63	.	.	.	3	.	.
<i>Carex riparia</i>	E1	50	.	.	.	6	3	.	100
<i>Carex rostrata</i>	E1	25	13
<i>Poa palustris</i>	E1	25	.	.	.	10	3	.	.
<i>Carex acutiformis</i>	E1	38	.	.	100	19	36	.
<i>Carex paniculata</i>	E1	25	.	.	2	.	.	.
<i>Phragmites australis</i>	E1	13	.	.	19	41	.	50
<i>Peucedanum palustre</i>	E1	8	9	73	83
<i>Scutellaria gelericulata</i>	E1	8	16	.	.
<i>Berrula erecta</i>	E1	6	.	.	.
<i>Nasturtium officinale</i>	E1	6	6	.	.
<i>Typha latifolia</i>	E1	6	6	.	.
<i>Carex otrubae</i>	E1	2	3	.	.
<i>Scoenoplectus lacustris</i>	E1	9	.	.
<i>Sparganium erectum</i>	E1	33
FC Filipendulo-Convolutea																		
<i>Calystegia sepium</i>	E1	10	.	.	25	.	.	.	19	28	.	.
<i>Epilobium parviflorum</i>	E1	10
<i>Fallopia japonica</i>	E1	10
<i>Epilobium hirsutum</i>	E1	13
<i>Cucubalus baccifer</i>	E1	4	.	.	.
BT Bidentetea tripartitetae																		
<i>Polygonum hydropiper</i>	E1	25	18	14	12	.	9	33
<i>Bidens frondosa</i>	E1	25	2	13	.	.
<i>Bidens tripartita</i>	E1	10	6	.	.
AV Artemisietea vulgaris																		
<i>Artemisia vulgaris</i>	E1	20	3	.	.
<i>Echinops sphaerocephalus</i>	E1	20
<i>Silene latifolia subsp. alba</i>	E1	20
<i>Rumex obtusifolius</i>	E1	10	.	14
<i>Asplenium viride</i>	E1	10
<i>Artemisia verlotiorum</i>	E1	3	.	.
<i>Cirsium vulgare</i>	E1	9	.
GU Galio-Urticetea																		
<i>Aegopodium podagraria</i>	E1	94	100	.	.	.	50	90	33	.	.	.	82	100
<i>Glechoma hederacea</i>	E1	39	83	75	40	60	50	30	22	14	.	13	.	.	4	6	9	17
<i>Geum urbanum</i>	E1	28	33	50	80	80	75	80	.	43	50	.	.	.	10	6	.	.
<i>Viola odorata</i>	E1	17	.	.	.	20	25	.	.	14	2	.	.	.
<i>Alliaria petiolata</i>	E1	11	33	25	10	100	.	10
<i>Chaerophyllum aureum</i>	E1	6	.	25	30	20
<i>Lamium maculatum</i>	E1	6	.	.	10	40	.	10	.	.	.	25	.	86	2	.	9	.
<i>Parietaria officinalis</i>	E1	6	50	.	.	20	.	10
<i>Galium aparine</i>	E1	.	17	50	.	60	.	30	.	.	.	25	.	.	19	.	.	.
<i>Impatiens glandulifera</i>	E1	.	.	25	.	60	25
<i>Helianthus tuberosus</i>	E1	20	.	10
<i>Solidago gigantea</i>	E1	80	.	40	.	.	.	13	9	43	4	3	.	50

Successive number (Zaporedna številka)	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
<i>Urtica dioica</i>	E1	.	.	.	80	.	20	11	.	.	25	55	71	35	28	9	67
<i>Impatiens parviflora</i>	E1	.	.	.	40	.	10	.	.	.	18	14
<i>Petasites hybridus</i>	E1	90	100
<i>Solidago canadensis</i>	E1	25	3	.	.
SM <i>Stellarietea mediae</i>																	
<i>Allium vineale</i>	E1	17	.	.	.	25	20	.	14
<i>Stellaria media</i> s.str.	E1	6	.	25	.	20
<i>Lapsana communis</i>	E1	.	.	25
<i>Chelidonium majus</i>	E1	.	.	.	40	14
<i>Bromus sterilis</i>	E1	.	.	.	40
<i>Erigeron annuus</i> subsp. <i>annuus</i>	E1	.	.	.	20	3	.	.
<i>Solanum nigrum</i>	E1	25	2	3	.	.
<i>Plantago major</i> subsp. <i>major</i>	E1	0	10
<i>Aethusa cynapium</i>	E1	10
TR <i>Thlaspietea rotundifolii</i> , <i>Asplenietea trichomanis</i>																	
<i>Cardaminopsis arenosa</i>	E1	6
AT <i>Polypodium vulgare</i>	E1	6	.	.	.	25
<i>Petasites paradoxus</i>	E1	10
<i>Peucedanum verticillare</i>	E1	10
O Other species (Druge vrste)																	
<i>Robinia pseudoacacia</i>	E3b	83	33	.	10	20	25	.	11	14
<i>Robinia pseudoacacia</i>	E3a	17	.	10
<i>Robinia pseudoacacia</i>	E2b	11
<i>Robinia pseudoacacia</i>	E2a	6	.	.	.	25
<i>Prunus insititia</i>	E3b	6
<i>Prunus insititia</i>	E3a	.	17	.	20	.	25	.	.	.	13
<i>Prunus insititia</i>	E2b	.	17	11	14
<i>Prunus insititia</i>	E2a	22	33	22	.	25
<i>Ficus carica</i>	E1	6
<i>Narcissus pseudonarcissus</i>	E1	6	17
<i>Prunus</i> sp.	E2a	6
<i>Spiraea japonica</i>	E2a	6	33
<i>Malus domestica</i>	E3a	.	.	10	2	.	.	.
<i>Hesperis matronalis</i>	E1	.	.	.	20
<i>Allium</i> sp.	E1	25
<i>Mentha</i> sp.	E1	10
<i>Viola</i> sp.	E1	10	27	.
<i>Quercus rubra</i>	E3b	11
<i>Vitis vinifera</i>	E3a	14
<i>Carex</i> sp.	E1	14	.	13	.	57
<i>Juniperus communis</i>	E2a	14
<i>Trachycarpus fortunei</i>	E1	14
<i>Potamogeton</i> sp.	E1	13
<i>Platanus x hispanica</i>	E3b	8	6	.	.
<i>Hottonia palustris</i>	E1	6	.	.
M Mosses (Mahovi)																	
<i>Ctenidium molluscum</i>	E0	17
<i>Plagiommium undulatum</i>	E0	6	.	.	.	20	50	60	22	14	.	9	29	.	.	45	50
<i>Brachythecium rutabulum</i>	E0	6
<i>Homalothecium lutescens</i>	E0	6
<i>Isoetecium alopecuroides</i>	E0	6
<i>Atrichum undulatum</i>	E0	10
<i>Climacium dendroides</i>	E0	10
<i>Conocephalum conicum</i>	E0	20
<i>Brachythecium</i> sp.	E0	29
<i>Polytrichum formosum</i>	E0	18
<i>Plagiommium cuspidatum</i>	E0	18
<i>Plagiommium affine</i>	E0	18
<i>Sphagnum squarrosum</i>	E0	9
<i>Sphagnum</i> sp.	E0	9
<i>Mnium hornum</i>	E0	9

OrClo *Ornithogalo pyraicae-Carpinetum betuli lamietosum orvalae*, the Vipava Valley, this article, Table 7, relevés 20–39

LoAg-Vd *Lamio orvalae-Alnetum glutinosae*, the Vipava Valley, this article, Table 7, relevés 14–19

LoAgb-R *Lamio orvalae-Alnetum glutinosae* var. *Scilla bifolia*, the Reka Valley, this article, Table 9, relevés 6–9

LoAgcb-R *Lamio orvalae-Alnetum glutinosae* var. *Cardamine bulbifera*, the Reka Valley, this article, Table 9, relevés 10–19

LoAgb1-R *Lamio orvalae-Alnetum glutinosae* var. *Scilla bifolia*, the Reka Valley, this article, Table 9, relevés 1–5

LoAg1-Vd *Lamio orvalae-Alnetum glutinosae*, the Vipava Valley, this article, Table 7, relevés 10–13

LoAg-ZP *Lamio orvalae-Alnetum glutinosae* s. lat., the Soča and Idrija Valleys, Dakskobler, (2016, mscr.)

PsCb1a-Vd *Pseudostellario-Carpinetum betuli leucojetosum aestivi*, the Vipava Valley, Lijak, this article, Table 7, relevés 1–9

Fra-Is *Rusco aculeati-Fraxinetum angustifoliae* nom. prov., Istria, Dakskobler & Sadar (2016, mscr.)

CacAg-R *Carici acutae-Alnetum glutinosae* nom. prov., the Reka Valley, Dakskobler (2016, mscr.)

CraA-Md *Carici randalpinae-Alnetum glutinosae* Martinčič 2007 nom. prov., Dolenjska, the Rašica Valley, Dakskobler (2016, mscr.)

CelAggr-A *Carici elongatae-Alnetum glutinosae* var. *Geum rivale*, Dolenjska, the Kočevsko region, Accetto (1994, Table 3)

SnAg-A *Stellario-Alnetum glutinosae* var. geogr. *Knautia drymeia*, Dolenjska, Accetto (1994, Table 4)

ChAg-It *Corno hungaricae-Alnetum glutinosae*, N-Italy, Sburlino et al. (2011, Table 1)

CelAg-It *Carici elatae-Alnetum glutinosae*, N-Italy, Sburlino et al. (2011, Table 2)

CelAg1a-A *Carici elongatae-Alnetum glutinosae* var. *Leucojum aestivum*, Dolenjska, the Krka Valley, Accetto (1994, Table 2)

CelAgcr-A *Carici elongatae-Alnetum glutinosae caricetosum ripariae*, the Mura region (Pomurje), Accetto (1994, Table 1, relevés 1–6).

NOTULA

Nomenclature correction of the name and rank of the association
Chamaecytiso hirsuti-Quercetum petraeae Dakskobler 2014 nom. illeg.

Nomenklaturni popravek imena in ranga asociacije *Chamaecytiso*
hirsuti-Quercetum petraeae Dakskobler 2014 nom. illeg.

Igor DAKSKOBLER

Some years ago we published a phytosociological analysis of *Quercus petraea* stands, whose herb layer is dominated by *Erica carnea* in the flysch hills of Vipavska Brda and on the margins of the Vrhe plateau (southwestern Slovenia) – DAKSKOBLER (2014). We determined that they are a long-term degradation stage on beech forest sites from the association *Seslerio autumnalis-Fagetum* M. Wraber ex Borhidi 1963. Based on comparisons with similar sessile oak stands from the associations *Melampyro vulgati-Quercetum petraeae* Puncer et Zupančič 1979, *Seslerio autumnalis-Quercetum petraeae* Poldini (1964) 1982 and *Erico carnea-Quercetum petraeae* Krause et Ludwig ex Horvat 1959, the studied stands were classified into the new association *Chamaecytiso hirsuti-Quercetum petraeae*, into the alliance *Carpinion orientalis* Horvat 1958 and into the order *Quercetalia pubescenti-petraeae* Klika 1933. We had overlooked, however, that the same name, *Cytiso hirsuti-Quercetum petraeae* (Stefanović 1964) Pallas 2003 (BOHN & NEUHÄUSL 2003) or *Cytiso hirsuti-Quercetum petraeae* (Stefanović 1964) Pallas in Bohn et Neuhäusl 2004 (STUPAR et al. 2015) was already used

for different *Quercus petraea* woods in Bosnia and Herzegovina that belong to the alliance *Castaneo-Quercion* Soó 1964 and the order *Quercetalia roboris* Tüxen 1931. According to the Code of phytosociological nomenclature (WEBER et al. 2000, Art. 31, 753–754) the name *Chamaecytiso hirsuti-Quercetum petraeae* Dakskobler 2014 is a later homonym and illegitimate, while *Cytisus hirsutus* is a synonym for *Chamaecytisus hirsutus*.

As it is not possible to find a proper new name for our association, we decided for a change of rank. We subordinated the association *Chamaecytiso hirsuti-Quercetum petraeae* Dakskobler 2014 nom. illeg. to the association *Seslerio autumnalis-Quercetum petraeae* Poldini (1964) 1982 as a new subassociation *Seslerio autumnalis-Quercetum petraeae ericetosum carnea* subass. nov. hoc loco. This also means that we consider the association *Seslerio autumnalis-Quercetum petraeae* in a broader sense, not only as a primary sessile oak community of the Vipava Valley, Karst and Istria, but also including secondary *Quercus petraea* communities of this area on potential beech sites.

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NAVODILA AVTORJEM

Folia biologica et geologica so znanstvena revija IV. razreda SAZU za naravoslovne vede. Objavljajo naravoslovne znanstvene razprave in pregledne članke, ki se nanašajo predvsem na raziskave v našem etničnem območju Slovenije, pa tudi raziskave na območju Evrope in širše, ki so pomembne, potrebne ali primerljive za naša preučevanja.

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Znanstvena razprava zajema celovit opis izvirne raziskave, ki vključuje teoretični pregled tematike, podrobno predstavlja rezultate z razpravo in zaključki ali sklepi in pregled citiranih avtorjev. V izjemnih primerih so namesto literaturnega pregleda dovoljeni viri, če to zahteva vsebina razprave.

Razprava naj ima klasično razčlenitev (uvod, material in metode, rezultati, diskusija z zaključki, zahvale, literatura idr.).

Dolžina razprave, vključno s tabelami, grafikoni, tablami, slikami ipd., praviloma ne sme presegati 2 avtorskih pol oziroma 30 strani tipkopisa. Zaželeno so razprave v obsegu ene avtorske pole oziroma do dvajset strani tipkopisa.

Razpravo ocenjujeta recenzenta, od katerih je eden praviloma član SAZU, drugi pa ustrezní tuji strokovnjak. Recenzente na predlog uredniškega odbora revije *Folia biologica et geologica* potrdi IV. razred SAZU.

Razprava gre v tisk, ko jo na predlog uredniškega odbora na seji sprejmeta IV. razred in predsedstvo SAZU.

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Razprava ali članek sta lahko pisana v slovenščini ali katerem od svetovnih jezikov. V slovenščini zlasti tedaj, če je tematika lokalnega značaja.

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6. POVZETEK

Za razprave ali članke, pisane v slovenščini, mora biti povzetek v angleščini, za razprave ali članke v tujem jeziku ustrezen slovenski povzetek. Povzetek mora biti dovolj obširen, da je tematika jasno prikazana in razumljiva domačemu in tujemu bralcu. Dati mora informacijo o namenu, metodi, rezultatu in zaključkih. Okvirno naj povzetek zajema 10 do 20 % obsega razprave oziroma članka.

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Izveček mora podati jedrnato informacijo o namenu in zaključkih razprave ali članka. Napisan mora biti v slovenskem in angleškem jeziku.

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Število ključnih besed naj ne presega 10 besed. Predstaviti morajo področje raziskave, podane v razpravi ali članku. Napisane morajo biti v slovenskem in angleškem jeziku.

9. NASLOV RAZPRAVE ALI ČLANKA

Naslov razprave ali članka naj bo kratek in razumljiv. Za naslovom sledi ime/imena avtorja/avtorjev (ime in priimek).

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Pod ključnimi besedami spodaj je naslov avtorja/avtorjev, in sicer akademski naslov, ime, priimek, ustanova, mesto z oznako države in poštno številko, država, ali elektronski poštni naslov.

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Uvod se mora nanašati le na vsebino razprave ali članka.

12. ZAKLJUČKI ALI SKLEPI

Zaključki ali sklepi morajo vsebovati sintezo glavnih ugotovitev glede na zastavljena vprašanja in razrešujejo ali nakazujejo problem raziskave.

13. TABELE, TABLE, GRAFIKONI, SLIKE IPD.

Tabele, table, grafikoni, slike ipd. v razpravi ali članku naj bodo jasne, njihovo mesto mora biti nedvoumno označeno, njihovo število naj racionalno ustreza vsebini. Tabele, table, slike, ilustracije, grafikoni ipd. skupaj z naslovi naj bodo priloženi na posebnih listih. Če so slike v

digitalni obliki, morajo biti pripravljene u zapisu **.tiff** v barvni skali **CMYK** in resoluciji vsaj **300 DPI/inch**. Risa-ne slike pa v zapisu **.eps**.

Pri fitocenoloških tabelah se tam, kjer ni zastopana rastlinska vrsta, natisne pika.

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Uporabljeno literaturo citiramo med besedilom. Citirane avtorje pišemo v kapitelkah. Enega avtorja piše-mo » (Priimek leto)« ali »(Priimek leto: strani)« ali »Priimek leto« [npr. (BUKRY 1974) ali (OBERDORFER 1979: 218) ali ... POLDINI (1991) ...]. Če citiramo več del istega avtorja, objavljenih v istem letu, posamezno delo ozna-čimo po abecednem redu »Priimek leto mala črka« [npr. ...HORVATÍĆ (1963 a)... ali (HORVATÍĆ 1963 b)]. Avtor-jem z enakim priimkom dodamo pred priimkom prvo črko imena (npr. R. TUXEN ali J. TUXEN). Več avtorjev istega dela citiramo po naslednjih načelih: delo do treh avtorjev »Priimek, Priimek & Priimek leto: strani« [npr. (SHEARER, PAPIKE & SIMON 1984) ali PEARCE & CANN (1973: 290-300)...]. Če so več kot trije avtorji, citiramo »Priimek prvega avtorja et al. leto: strani« ali »Priimek prvega avtorja s sodelavci leto« [npr. NOLL et al. 1996: 590 ali ...MEUSEL s sodelavci (1965)].

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– Razprava ali članek:

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KAJFEŽ, L. & A. HOČEVAR, 1984: *Klima. Tlatvorni činitelji*. V D. Stepančič: *Komentar k listu Murska Sobota*. Osnovna pedološka karta SFRJ. Pedološka karta Slovenije 1:50.000 (Ljubljana): 7–9.

LE LOEUFF, J., E. BUFFEAUT, M. MARTIN & H. TONG, 1993: *Decouverte d'Hadrosauridae (Dinosauria, Ornithischia) dans le Maastrichtien des Corbieres (Aude, France)*. C. R. Acad. Sci. Paris, t. 316, Ser. II: 1023–1029.

– Knjiga:

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Če sta različna kraja založbe in tiskarne, se navaja kraj založbe.

– Elaborat ali poročilo:

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– Atlasi, karte, načrti ipd.:

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LETNO poročilo meteorološke službe za leto 1957. Hidrometeorološki zavod SR Slovenije. Ljubljana.

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Latinska imena rodov, vrst in infraspecifičnih tak-sonov se pišejo kurzivno. V fitocenoloških razpravah ali člankih se vsi sintaksoni pišejo kurzivno.

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Članek naj bo pisan v formatu RTF z medvrstičnim razmikom 1,5 na A4 (DIN) formatu. Uredniku je treba oddati izvornik in kopijo ter zapis na disketi 3,5 ali na CD-ROM-u. Tabele in slike so posebej priložene tekstu. Slike so lahko priložene kot datoteke na CD-ROM-u, za podrobnosti se vpraša uredništvo.

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It is the entire description of novel research including the theoretical review of the subjects, presenting in detail the results, conclusions, and the survey of literature of the authors cited. In exceptional cases the survey of literature may be replaced by sources, if the purport requires it.

It should be composed in classic manner: introduction, material and methods, results, discussion with conclusions, acknowledgments, literature, etc.

The treatise should not be longer than 30 pages, including tables, graphs, figures and others. Much desired are treatises of 20 pages.

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The treatise or article ought not to be published previously in other periodicals or books.

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When the treatise or article is written in Slovenian, the summary should be in English. When they are in foreign language, the summary should be in Slovenian. It should be so extensive that the subjects are clear and understandable to domestic and foreign reader. It should give the information about the intention, method, result, and conclusions of the treatise or article. It should not be longer than 10 to 20% of the treatise or article itself.

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It should give concise information about the intention and conclusions of the treatise or article. It must be written in English and Slovenian.

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The address of author/authors should be at the bottom of the page: academic title, name, surname, institution, town and state mark, post number, state, or e-mail of the author/authors.

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KAJFEŽ, L. & A. HOČEVAR, 1984: *Klima. Tlatvorni činitelji*. V D. Stepančič: *Komentar k listu Murska Sobota*. Osnovna pedološka karta SFRJ. Pedološka karta Slovenije 1:50.000 (Ljubljana): 7–9.

LE LOEUFF, J., E. BUFFEAUT, M. MARTIN & H. TONG, 1993: *Découverte d'Hadrosauridae (Dinosauria, Ornithischia) dans le Maastrichtien des Corbieres (Aude, France)*. C. R. Acad. Sci. Paris, t. 316, Ser. II: 1023-1029.

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GORTANI, L. & M. GORTANI, 1905: *Flora Friuliana*. Udine.

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LETNO poročilo meteorološke službe za leto 1957. Hidrometeorološki zavod SR Slovenije. Ljubljana.

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Latin names for order, series, and infraspecific taxa are to be written in italics. All syntaxa written in phytocoenological treatises or articles are to be in italics.

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